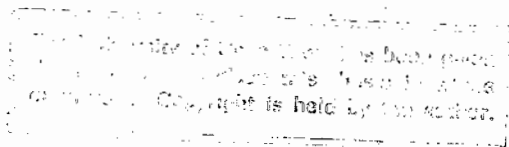


SOLUTION EQUILIBRIA OF THE METHYLDOPA-COPPER(II) SYSTEM
AND THE
CRYSTAL AND MOLECULAR STRUCTURES OF SELECTED COMPOUNDS

A thesis submitted to
THE UNIVERSITY OF CAPE TOWN
in fulfilment of the requirements for the degree of
DOCTOR OF PHILOSOPHY

by

M. ROBERT W. WRIGHT



Department of Physical Chemistry,
University of Cape Town,
Rondebosch, 7700,
Republic of South Africa.

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SUMMARY

The contents of this thesis are essentially in two parts. The first deals with solution studies relating to the methyldopa-copper(II) system and its relevance to medicinal chemistry, a full abstract of which is given on page 1 of the main text. The second part describes five single crystal x-ray investigations into some interesting compounds isolated at this university. The abstracts to these topics may be found preceding the relevant sections as they appear in the text on pages 94, 118 and 146, respectively.

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PART ISOLUTION EQUILIBRIA OF THE METHYLDOPA-COPPER(II)
SYSTEM

ABSTRACT

The system copper(II) ions in aqueous solution in the presence of L-3-(3',4'-dihydroxyphenyl)-2-methylalanine (methyldopa), at 25°C and $I = 0,150 \text{ mol dm}^{-3} \text{ Na}^+\text{ClO}_4^-$ has been characterised using glass electrode potentiometry. Both mononuclear and oligonuclear complexes were found to occur, the former including a series of successively deprotonated species in the higher ligand to metal ratio titrations, the latter including a cyclic dimer in the 1:1 ligand to metal solutions. Interpretation of the results was facilitated by (i) ultra-violet/visible spectrophotometry of solutions containing copper(II), methyldopa or alanine or catechol at various pH's, (ii) ^1H NMR measurements of methyldopa in $^2\text{H}_2\text{O}$ solutions, (iii) glass electrode potentiometric measurements on alanine and catechol in the presence of copper(II) ions. Formation constants are given for 3 proton - and 11 copper(II) - complexes of methyldopa, 2 proton and 2 copper(II) complexes of alanine and 1 proton and 2 copper(II) complexes of catechol.

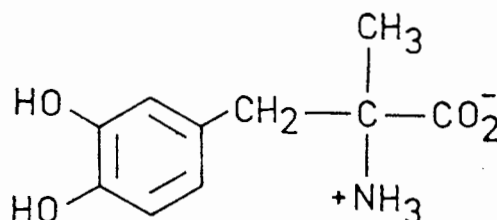
1. INTRODUCTION

1.1 Importance of methyldopa as an anti-hypertensive agent in vivo and its proposed mode of action.

Essential hypertension i.e. that condition of high blood pressure where the specific causes cannot be identified, can be classified into two types. Firstly, early or labile hypertension, where the haemodynamic pattern is characterised by increased cardiac output with the total peripheral resistance not significantly different from that of controls and generally diagnosed in relatively young hypertensive patients; and secondly, established hypertension, in which the haemodynamic pattern is a normal cardiac output but an increased total peripheral resistance. The mechanisms by which the transition from labile to established hypertension are brought about, as well as the fraction of patients that undergo it, as yet remains unknown¹.

The importance of L-3-(3',4'-dihydroxyphenyl)-2-methylalanine (methyldopa - see Fig 1.1.1) as a potential antihypertensive agent in remedying this condition was first reported in 1960 by Oates *et al.*² Since then, numerous experiments have confirmed its usefulness in this area^{3,4} and today its clinical applications in man are becoming increasingly common. Of the two enantiomorphs of methyldopa, it has been shown that it is only the L-(-) isomer which has hypotensive activity⁵. Fig 1.1.2 depicts the Fischer projection representation of the active form. The absolute con-

Fig 1.1.1



figuration was determined by chemical methods⁶ and optical rotatory dispersion data⁷.

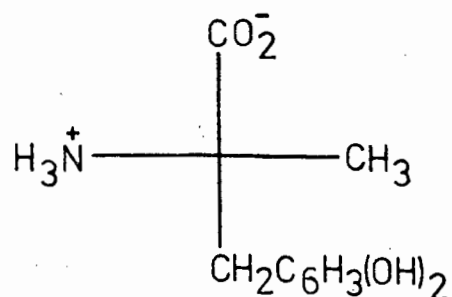


Fig 1.1.2

Various modes of action have been postulated to explain the drug's effect in vivo, the most important of which will now be briefly discussed.

(i) Decarboxylase inhibitor.

Methyldopa was initially thought to inhibit the decarboxylation of the related compound 3-(3',4'-dihydroxyphenyl)-2-alanine (dopa) to dopamine, and hence the subsequent formation of noradrenaline. The decreased rate of formation of noradrenaline (in hypertension there is a tendency for excessive noradrenaline to be released at the sympathetic nerve endings of arteriolar smooth muscle⁸) would then result in a fall in arterial blood pressure. This explanation was considered incomplete as:

- (a) the antihypertensive effect persists longer than the enzymatic inhibition i.e. there is no causal relationship between the administration of methyldopa as a decarboxylase inhibitor and the expected fall in blood pressure;⁹
- (b) several decarboxylase inhibitors which are more

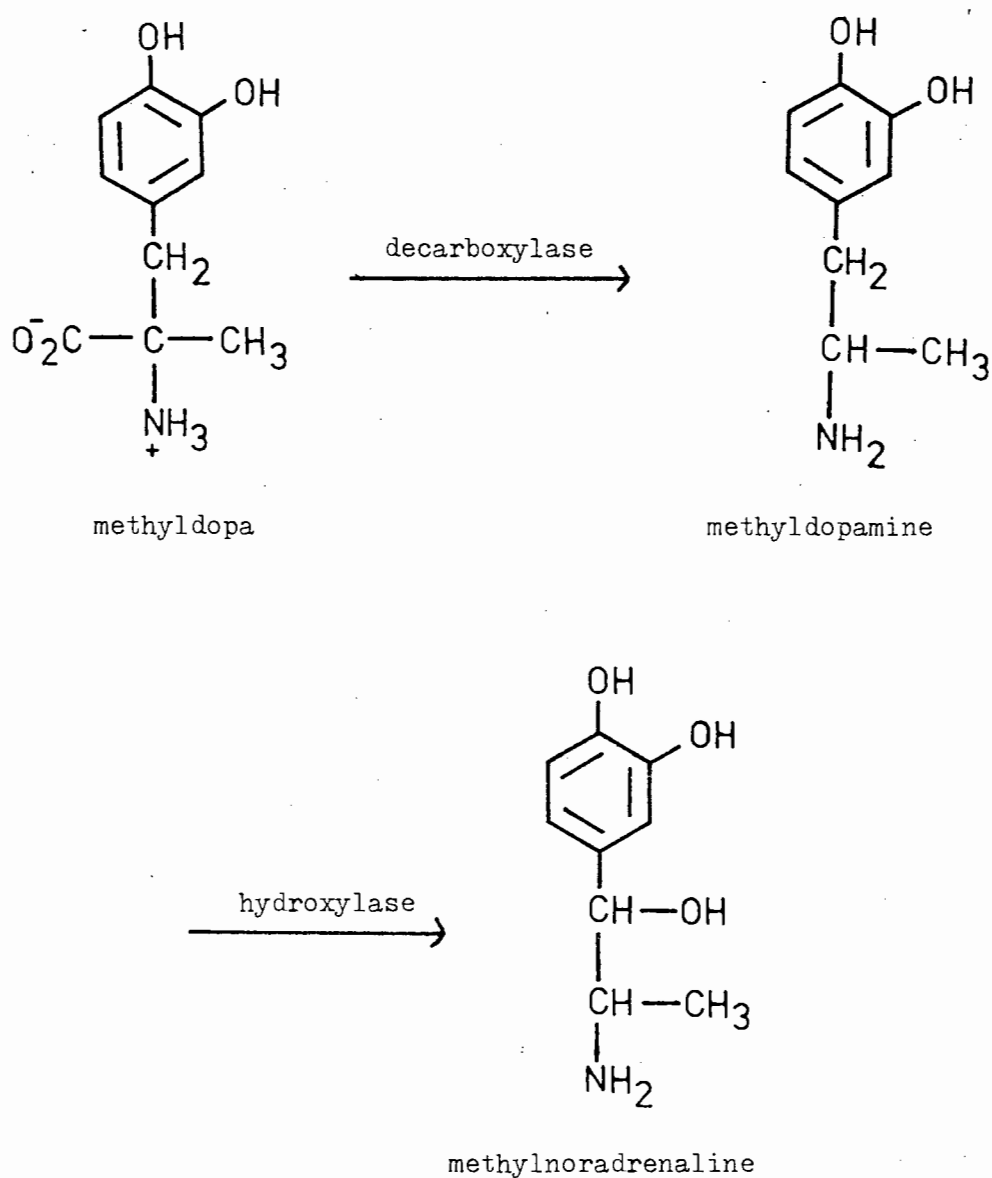


Fig 1.1.3

potent than methyl dopa (methyl dopa hydrazine, 4-bromo-3-hydroxybenzyloxyamine) do not lower the blood pressure of hypertensive patients;¹⁰

(ii) False-transmitter theory:¹¹

Methyl dopa is metabolised in the body via the enzymes dopa decarboxylase and dopamine- β -hydroxylase into methyl dopamine and methyl noradrenaline (Fig 1.1.3), analogous to the biotransformation of dopa into dopamine and noradrenaline.

A mixture of stored methylnoradrenaline and noradrenaline is released from nerve endings as a result of peripheral adrenergic activity. If it is assumed that both transmitters compete for receptors in the target organ (in this case mainly the blood vessels), and that methylnoradrenaline is a less effective receptor stimulant than noradrenaline, this might explain the observed decrease in blood pressure. However, this false transmitter theory has also been proved to be unsatisfactory. Besides the unconvincing causal relationships detected, it is generally thought that the adrenergic properties of methylnoradrenaline and noradrenaline, although different in some species, is probably not different in all in which methyldopa is effective e.g. in man.¹²

(iii) Central Hypotensive Action.

There is increasing evidence that replacement of noradrenaline in sympathetic neurons by the less potent false-transmitter methylnoradrenaline does not completely account for the hypotensive action of methyldopa.^{13,14} Experiments have confirmed that the hypotensive response due to methyldopa administered into the central nervous system, e.g. the cerebrospinal fluid, is significantly larger than systemic administration and that it is methylnoradrenaline which is the active metabolite. The medullary region of the lower brainstem contains α -adrenergic receptors which activate inhibitory neurons that diminish peripheral adrenergic activity.^{15,16} The antihypertensive effect of methyldopa may thus be explained in terms of methylnoradrenaline coming into

contact with these sites and being a more potent stimulant than noradrenaline, also present in this region. Adequate amounts of dopa decarboxylase and dopamine- β -hydroxylase are present in this region for the biotransformation of methyldopa into the active species to take place.

(iv) Suppression of Renin Release.

The ability of methyldopa to suppress renin release from the kidneys of patients whose hypertensive state is brought about by this condition has also been demonstrated.¹⁷ Whether this property is due solely to a false-transmitter type effect of the drug on the post ganglionic renal adrenergic nerve, or whether a centrally mediated renin-suppressing action is also involved, awaits final demonstration.¹⁸

As a summary, it seems that the mode of action of methyldopa in relieving hypertension has not yet been established with certainty. The relative predominance of the possible modes depends to a large extent on the method of administration and the type of species being studied. However it is generally accepted that the centrally mediated action is the most important.

The use of methyldopa has its accompanying side effects. Amongst those detected are autoimmune haemolytic anaemia,¹⁹ reversible abnormalities of liver function tests and low grade lactation.²⁰ The potency of tricyclic anti-depressant drugs (e.g. desipramine) administered concurrently with methyldopa has also been found to be reduced.²¹

1.2. Metal ion interactions in vivo:

It has been well established that metal ions, although contributing negligibly to total body weight, nevertheless play an important role in the healthy functioning of the living organism. Their importance is becoming increasingly apparent as a result of research into fields such as enzymology and neurology, where it has been found that deviation of total metal ion concentrations outside comparatively narrow limits can lead to a rapid deterioration in essential bodily functions.²² Wilson's disease (a nervous disorder due to an excessive concentration of copper in the liver, brain and tissues²³) and anaemia (caused by iron deficiency²⁴) are but two of numerous examples in which the imbalance of metal ions in vivo is directly implicated in the breakdown of physiological processes. In most cases it is not known whether imbalances are the cause or the effect of the disease and the interplay of metal ion concentrations resulting from such imbalances is not well understood.

Nature has selected metal ions, and in particular those of the transition series, for the special role they play in life processes because of their ability to exist as stable ions, their capacity to form stereochemical bonds having low reaction energies and the comparative ease with which they are incorporated into redox systems. Pearson²⁵ in the early sixties developed the HSAB principle which classified acids and bases as hard or soft depending on their electronegativity, oxidisability and polarisability. Hard acids prefer hard bases and soft prefer soft. Given the vast range of species present in living systems, the HSAB principle applied to metal ion equilibria in vivo goes a long way in qualitatively explaining

the bond strengths and sites of co-ordination of complexes that have been observed. In addition, the specificity of most of the reactions occurring can be understood on this basis.

The ions of sodium, potassium, calcium, magnesium, manganese, iron, cobalt, copper and zinc are generally accepted as part of the list of 25 elements essential for healthy life. Table 1.2.1. shows briefly some of the more important biological processes in which they participate.

Table 1.2.1.

<u>The Important Role of Metal ions in the healthy organism.²²</u>	
Na^+, K^+	Maintaining osmotic pressure on either side of the cell wall, muscle tone.
$\text{Ca}^{2+}, \text{Mg}^{2+}$	Nerve impulse transmission, formation of bones and teeth, blood clotting.
$\text{Mn}^{2+}/\text{Mn}^{3+}$	Enzyme activation.
$\text{Fe}^{2+}/\text{Fe}^{3+}$	Protein formation, redox reactions, oxygen transport.
$\text{Co}^{2+}/\text{Co}^{3+}$	Manufacture of vitamin B_{12} , enzyme activation.
$\text{Cu}^+/\text{Cu}^{2+}$	Production of haemoglobin, oxygen storage and transport.
Zn^{2+}	Enzyme activation.

As can be seen from the table, metal ions figure predominantly in enzyme reactions. Enzymes promote

physiological reactions by providing alternative pathways with lower activation energies and hence faster reaction rates. They have the ability of positioning the substrate adjacent to an active centre, where the three dimensional geometry and the energetics of the system bring about an enhanced biological reaction. Carboxy peptidase A is an enzyme which hydrolyses the c-terminal amino acid residue from a protein or peptide chain. Consisting of a polypeptide chain of 307 amino acid residues, it contains one Zn(II) ion per molecule, which if removed, renders the enzyme inactive. Some transition metal ions (Fe(II), Mn(II), Co(II), Ni(II)), by occupying the site formerly held by Zn(II), are able to regenerate peptidase activity in the apoenzyme.²⁶ Carbonic anhydrase oxaloacetate decarboxylase and alkaline phosphatase are other examples of enzymes which depend on bound metal ions for their activity.²⁷⁻²⁹ Haemoglobin, an important oxygen transporter in the blood, binds its metal ion more tightly. In this case, iron is covalently bonded within a protected cavity formed by the surrounding molecule and is therefore well sheltered from outside attack. In fact, metal ions in blood plasma can conveniently be subdivided into four fractions depending on the type of chelation with co-existing ligands.

- (i) Tightly bound by proteins and therefore not exchangeable under mild chemical conditions. Examples are haemoglobin and the cobaltic complex of vitamin B₁₂.
- (ii) Loosely bound by proteins and therefore in labile equilibrium. Included here are most of the enzymes and the metal ion carriers e.g. iron in transferrin and copper in serum albumen.

- (iii) Complexed to numerous low molecular weight ligands such as amino acids, carboxylates, phosphates, nucleosides and nucleotides.
- (iv) Non-chelated and existing in its free or aquated form.

For a given metal, the great proportion exists in the first two fractions, and it has been tentatively theorised that the second fraction can act as a so-called metal ion buffer. This idea has been shown to be useful in the calculation of the distribution of metal ions in physiological solutions by computer simulation.³⁰

The importance of metal ions in transport systems across cell membranes also warrants mention. For example, the concentration difference of Na^+ ions on either side of certain cell membranes has been shown to be intimately involved in the active transport of amino acids and sugars into cells.³¹ The concentration gradient is believed to be brought about by a so-called ion pump using ATP as the energy source. Chelates of dopa with metal ions, in particular Cu(II) and Zn(II), are transported across the blood/brain barrier at a faster rate than the uncomplexed ligand.³² The metal ion, by being complexed to the amine group of dopa is thought to inhibit the decarboxylation of dopa to dopamine by enzymes present in the precerebral areas.

Despite the mammoth amount of work which has been done on metal ion interactions in vivo, quantitative knowledge concerning the selective preferences of these metal ions for the many varied ligand species present in the body is still lacking. A comprehensive understanding of the role of metal

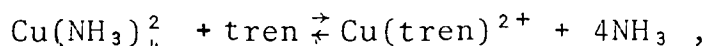
ions in life processes, with its resultant implications as far as nutrition, therapeutics and pollution are concerned, must of necessity await further progress.

1.3. Previous work on dopa-metal ion interactions and related systems.

Mention should be made of the more important factors influencing the magnitude of formation constants of metal ion-ligand interactions in general. Under a given set of conditions these depend on:

- (i) The nature of the metal ion. For the transition series, the well established Irving-Williams order is often followed i.e. $\text{Mn(II)} < \text{Fe(II)} < \text{Co(II)} < \text{Ni(II)} < \text{Cu(II)} > \text{Zn(II)}$.³³ Their position is a function of the electronic configuration of the outermost electrons, and is a manifestation of the crystal field stabilisation energy (CFSE) which the metal ion undergoes on complex formation.³⁴
- (ii) The number of metal ion-ligand bonds per ligand. The so-called chelate effect, predicting the normal order of increasing stability (monodentate < bidentate < terdentate) can be explained using thermodynamic and statistical arguments.

For example, in the system,

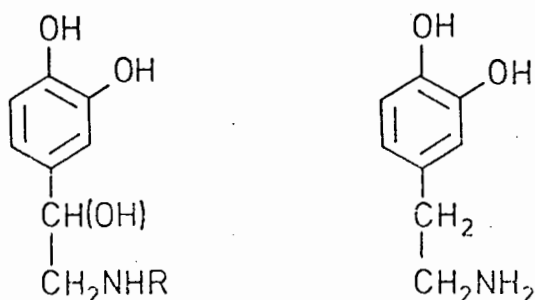


the equilibrium lies well over to the right for all but extremely low tren concentrations (tren \equiv triethylenetetramine).³⁵

- (iii) The acidity of the donor atoms. A ligand which binds protons strongly will have a high affinity for other cations.³⁶ A contributing factor here is the nature of

- any adjacent functional groups. Electron withdrawing groups will depress the pKa of a potential donor atom, making it a less potent complexing agent, whereas an electron donating group will have the opposite effect.
- (iv) Steric factors. Metal ion-amino acid chelates tend to form 5-membered planar ring structures. A distortion with accompanying bond strain brought about by bulky substituents on a participating atom will tend to have a destabilising influence.³⁷
- (v) Neutralisation of charge. When two oppositely charged species complex to form a neutral species, solvent molecules are released from the inner co-ordination spheres of the reactants. The resultant increased freedom of translational motion of these solvent molecules generally results in a positive entropy change for the reaction, and hence an enhanced stability for the complex. Clearly exothermic reactions favour complex formation and endothermic do not.
- (vi) The type of bond formed. An amine nitrogen is able to form a σ type bond with a transition metal ion, whereas a pyridyl or imadazole group has additional π -acceptor qualities. Electron density on the metal ion is reduced through the back donation of d electrons into vacant π orbitals of appropriate ligands, leading to a higher formation constant for the species involved than one would normally expect.³⁸
- (vii) The nature of the donor atoms. In a review, Sigel has noted that $\text{Cu}(\text{bipyridyl})^{2+}$ binds ligands with oxygen donors more firmly than those with nitrogen donors. Mixed N,O ligands have constants with intermediate values.³⁹

Exploratory work on dopa related catecholamines such as adrenaline, noradrenaline and dopamine (Fig 1.3.1) with metal ions has aided the understanding of the metal binding characteristics of dopa itself.



R = CH₃ : adrenaline

R = H : noradrenaline

dopamine

Fig 1 3.1

Weber *et al*⁴⁰ found in such systems that with zinc (II), cadmium(II) and lead(II) as the metal ion, only 1:1 catechol type complexes were present, and these were much less stable than the corresponding complexes of copper(II) and nickel(II). These last two metal ions were able to form species with two ligands per metal ion. Below a ligand:copper(II) ratio of 2:1, Jameson and Neillie⁴¹ working with adrenaline and noradrenaline reported the formation of tetramers (Fig 1.3.2) with not only the phenolic groups but also the side chain participating in complexation. In the case of catechol, this side chain participation is not possible, and the formation of the 2:1 complex on further addition of base is postulated, with the eventual precipitation of copper hydroxide. The formation of 1:1 and 2:1 catechol-copper(II) chelates has also

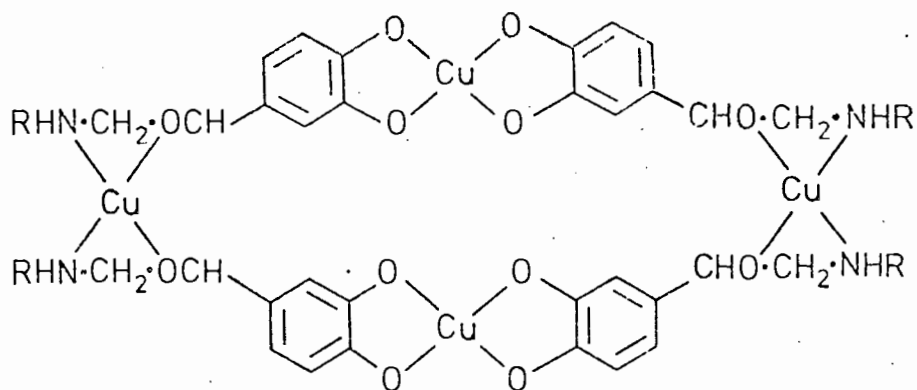


Fig 1.3.2

been reported by other workers.^{42,43}

Interest in dopa as a complexing ligand stems from the fact that, in addition to the terminal carboxylic acid-amine structure common to all amino acids, it possesses potential metal ion binding sites at its opposite end in the form of the two hydroxyl groups. Of all the dopa-metal ion systems that have been investigated, those involving copper(II) as the metal ion have received the most attention. This is likely due to this metal ion's kinetic lability and the high stability in aqueous media of many of its complexes.⁴⁴ In addition, copper(II) has been shown to play an important role in the *in vivo* reactions of catecholamines.⁴⁵

Although there is general agreement on the mode of complexation predominant at high and low pH values in the dopa-copper(II) system, inconsistencies as to the structures and stoichiometries of the species involved still remain. Experimental data obtained from potentiometry, ESR, UV/visible spectrophotometry and cyclic voltametry indicate that the bonding site of copper(II) to dopa is strongly pH dependent.^{40,32,46-52}

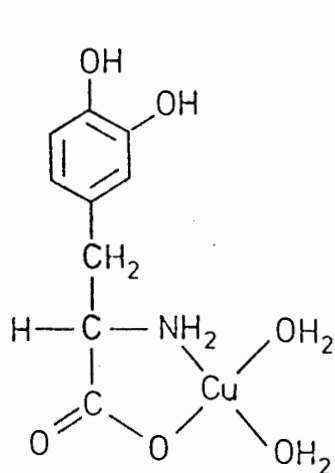
Gorton and Jameson⁴⁶ found that in a 1:1 dopa-copper(II) mixture a monomeric complex with the metal ion bound at the aminocarboxylate end of the ligand at a pH below 4 was replaced by a cyclic dimer involving catechol type bonding as the pH was increased to 6. (Fig 1.3.3)

In a subsequent paper,⁴⁷ the model was extended by introducing polymers. In studies with higher ligand to copper(II) ratios, Gergely and Kiss⁵¹ have determined the stability constants of complexes containing one and two ligands per metal ion, the bonds being of the amino acid type at low pH (<5) and of the catechol type at high pH (>9). A mixed complex with N,O and O,O bonds, as well as cyclic and open chain species in which dopa behaves as a bridging ligand, were evident in the intermediate pH range. Polymers of higher order were also postulated, but were not included in the final model due in part to their very low concentration. Further evidence of the ability of dopa to act as a simple amino acid at lower pH values and as catechol at higher pH values has been provided by the isolation and characterisation of a bluish-purple solid at pH 5 and an olive-green solid at pH 9,5 showing respectively these types of bonding.⁵⁰

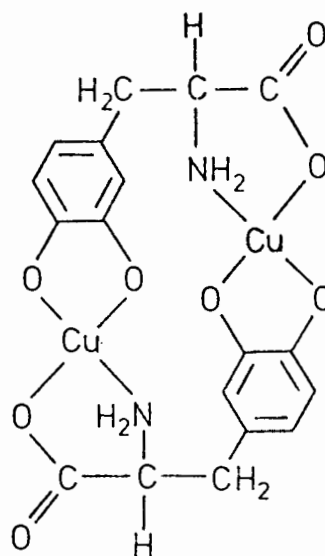
Copper(II) is known to bind simple amino acids strongly in the ratio of one or two ligands per metal ion at pH values between 3 and 5.^{37,53} In addition, the preference of copper(II) for ligands containing conjugated π ring systems for d electron back donation has already been mentioned.³⁸ Studies of catechol-copper(II) systems have shown that complexation with copper(II) occurs on the deprotonation of the catecholic hydroxyls at pH values only above 5, due to the high pKa's of these groups.⁴³ Thus it is reasonable to expect the pH

Fig 1.3.3

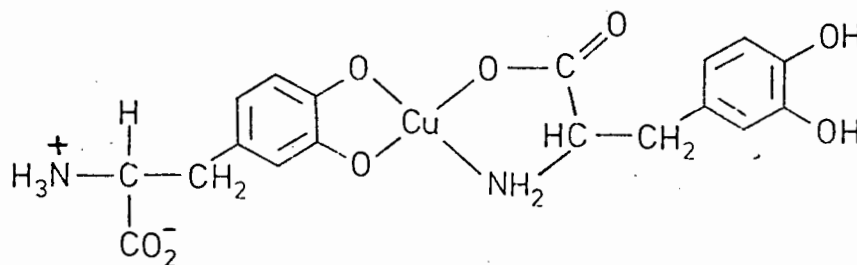
Modes of complexations in the dopa-copper(II) system



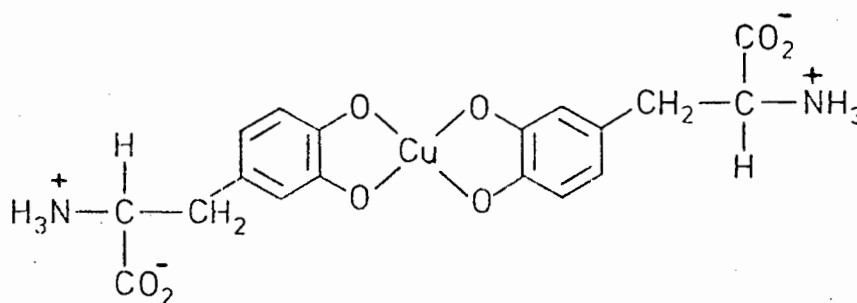
Monomeric amino acid type



Cyclic dimer



Mixed 0,0 and N,0 type



Dimeric catechol type

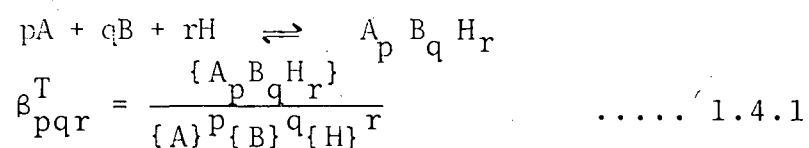
dependent mode of complexation of copper(II) to dopa to be mainly amino acid type chelation at low pH values and catechol type at high pH values.

Dopa complexations with metal ions other than copper(II) have not been as widely studied. Evidence at the present time suggests that nickel(II) forms amino acid type complexes with dopa, whereas magnesium(II) and zinc(II) prefer catechol type.⁵⁴

1.4. Ligand-metal ion equilibria in solution.

The theory of the study of ligand-metal ion equilibria in solution is comprehensively covered in many texts,⁵⁵⁻⁵⁷ and so only a brief account will be given here.

The relative amounts of complexes which exist in an aqueous solution can be calculated from their thermodynamic formation or stability constants, which relate the activity of the complex under investigation to the activities of the free components comprising the system. Thus for a system of 3 components, represented by A (ligand), B (metal ion) and H (proton), in which the following equilibrium is established,



β_{pqr}^T as defined above is the overall or cumulative thermodynamic formation constant. The concentration of water is generally unaffected by the participating equilibria within the system due to its large excess, and hence is omitted in the calculations. When $r = -1$ this refers to a proton removed to a water molecule or a hydroxide ligand added.

The system can also be expressed in terms of stepwise thermodynamic formation constants, K_{pqr}^T , which can be related

to the β_{pqr}^T 's. Hence, in the absence of metal ions, with just one ligand,

$$\begin{aligned}
 K_{101}^T &= \frac{\{AH\}}{\{A\}\{H\}} = \beta_{101}^T \\
 K_{102}^T &= \frac{\{AH_2\}}{\{AH\}\{H\}} = \frac{\beta_{102}^T}{K_{101}^T} \\
 &\vdots \\
 K_{10n}^T &= \frac{\{AH_n\}}{\{AH_{n-1}\}\{H\}} = \frac{\beta_{10n}^T}{\prod_{i=1}^{n-1} K_{10i}^T}
 \end{aligned}$$

Clearly analogous relationships can be derived in extended systems.

In practice, because of the difficulty in measuring activities, complex equilibria are usually expressed in terms of the concentrations of species. This technique entails using a non-complexing background electrolyte of sufficiently high molarity to maintain the activity coefficients of the reacting species effectively constant during the measurements. Hence for a given electrolyte, the activities of the components partaking in the equilibria are determined primarily by the total ionic strength of the medium, and are largely unaffected by a small change which may be caused through a slight perturbation of the equilibria. A limitation to this method is that the stability constants are only applicable to the ionic strength and the medium in which they are measured and their value in others can only be estimated. Nevertheless the stability constants can be measured over a wide range of ionic strengths, and thus extrapolated to zero ionic strength where the activities are represented by the concentrations.

1.5. Objectives.

The importance of metal ions in biological processes involving catecholamines has been made apparent in a number of

cases. For example, copper containing enzymes have been shown to be intimately involved in the metabolism of biogenic amines such as adrenaline and noradrenaline.^{58,59} Dopa, a member of this class, has for a number of years been administered in the treatment of Parkinson's disease, a disorder of the central nervous system. When chelated to certain metal ions, the drug shows an increased rate of transport across the blood/brain barrier in comparison to unchelated dopa.³² Methyldopa in turn has found wide application in the lowering of high blood pressure in man.

This work was undertaken in order to advance investigations concerning the stoichiometry, identity and distribution of methyldopa-copper(II) complexes co-existing in solution in the hope that it might throw further light on the understanding of the drug's clinical usefulness.

The techniques employed were:

- (i) Potentiometry and computer analysis, primarily in stability constant determination and identification of the complex species co-existing in equilibrium under various conditions e.g. concentration of the components and pH.
- (ii) Ultraviolet/visible spectrophotometry and nuclear magnetic resonance (NMR) with the aim of identifying the coordination and protonation sites of the methyldopa molecule.
- (iii) The computer program ECCLES³⁰ was used as an aid in estimating the relative metal ion populations at the two predominant sites of methyldopa.

The major part of the effort went into (i) whilst (ii) and (iii) were used to facilitate the selection of feasible models in (i).

2. TECHNIQUES USED

2.1. Computer programs used in model fitting.

2.1.1. MINIQUAD.

A solution containing components which are potentially capable of forming complexes will consist at equilibrium of a series of species, the stoichiometry and amounts of which will depend on the physical conditions prevalent, the chemical nature of the components and the magnitude of their attraction. In a study to determine their nature, the procedure is to make use of chemical knowledge gained from similar systems, or alternative experimental techniques, in choosing a model which will within reasonable error match the observed experimental data. The method of testing a postulated model on a set of potentiometric titration data as followed by the program MINIQUAD⁶⁰ for a system of 3 components (if need be up to five can be handled) will now be briefly described.

In a mixture containing several species of various types (mononuclear binary, protonated, hydroxo, oligonuclear), the total concentrations of the components can be expressed in terms of the free concentrations of the components and the various stability constants of the N complexes. Thus

$$[A]_T = [A] + \sum_{r=-R}^R \sum_{q=0}^Q \sum_{p=1}^P p \beta_{pqr} [A]^p [B]^q [H]^r \quad \dots 2.1.1.1.$$

$$[B]_T = [B] + \sum_{r=-R}^R \sum_{q=1}^Q \sum_{p=1}^P q \beta_{pqr} [A]^p [B]^q [H]^r \quad \dots 2.1.1.2.$$

$$[H]_T = [H] + \sum_{r=-R}^R \sum_{q=0}^Q \sum_{p=1}^P r \beta_{pqr} [A]^p [B]^q [H]^r \quad \dots 2.1.1.3.$$

A model with estimated formation constants is postulated. In a set of n titration points a total of 3n equations is set up. If only a glass or other $[H^+]$ sensitive electrode is used in the potentiometric measurements, it follows that the free hydrogen ion concentration for each point is known, and so the

number of unknowns is $(2n + m)$, where m is the number of formation constants to be determined. The number of degrees of freedom is $(n - m)$. By an iterative process, using the Gauss-Newton least squares method, the parameters $[A]$ and $[B]$ for each titration point, and the β_{pqr} 's common to all, are adjusted so as to minimize the error square sum

$$U = \sum_{i=1}^{3n} (f_i^{\text{calc}} - f_i^{\text{obs}})^2$$

where f_i^{calc} are the total component concentrations as calculated according to equations 2.1.1.1. to 2.1.1.3, and f_i^{obs} the corresponding experimental values. The model is said to be a possible representation of the true chemical state if the parameters are refined such that the function U is less than a pre-specified value. An important feature of MINQUAD is the optimisation of the parameter shifts, and successful refinement occurs when the shift changes after each cycle become sufficiently small.

Useful statistical outputs of the program include the following functions:

(i) R-factor:

$$R = \frac{\sum_{i=1}^{3n} (f_i^{\text{calc}} - f_i^{\text{obs}})^2}{\sum_{i=1}^{3n} (f_i^{\text{obs}})^2} \dots 2.1.1.4.$$

Generally the higher R is, the less good is the fit. A limiting value of R can be calculated using conservative estimates of the errors in the experimental measurements and following the method of propagation of errors (see Appendix A), and if the R as calculated in 2.1.1.4 is less than this R_{LIM} , the model can be said to be within the maximum allowed experimental error. It may so happen that two models give values R_1 and R_2 which are both less than this R_{LIM} . Vacca *et al*⁶¹ have applied

a method developed by Hamilton involving the ratio R_1/R_2 whereby one can decide whether they are significantly different at the α confidence level. If it is found that they are not, the simplest of the models matching the experimental data should be taken.

(ii) χ^2 - chi squared

This function as it appears in the program allows one to decide how closely the $(f_i^{\text{calc}} - f_i^{\text{obs}})$ values follow a normal Gaussian distribution about its zero mean, an indication as to the distribution of residuals on either side of the mean.

(iii) The residuals:

A scan of the residuals $(f_i^{\text{calc}} - f_i^{\text{obs}})$ on the mass balance equations may aid the user in postulating alternative models in subsequent runs, or perhaps to detect erroneous experimental points.

2.1.2. ZPLOT.

This program calculates the formation curve parameters \bar{Z} and $P[A]$, assuming only mononuclear binary complexes are formed, where

$$\begin{aligned} \bar{Z} &= \text{average no. of ligand molecules bound to metal} \\ &\quad \text{per metal ion} \\ &= \frac{[A \text{ bound to B}]}{[B]_T} \quad \dots 2.1.2.1 \end{aligned}$$

and

$$P[A] = -\log[\text{fully deprotonated ligand}] \quad \dots 2.1.2.2$$

The protonation formation curve parameter \bar{Z}_H , analogous to \bar{Z} , is calculated as

$$\bar{Z}_H = \frac{[\text{bound H}]}{[A \text{ not bound to B}]} \quad \dots 2.1.2.3$$

$$= \frac{\sum_{j=0}^J j [H]^j \beta_{10j}}{\sum_{j=0}^J [H]^j \beta_{10j}}$$

From equation 2.1.2.1

$$\bar{Z} = \frac{[A]_T - [A \text{ not bound to B}]}{[B]_T} \quad \dots\dots 2.1.2.4$$

Substituting from equation 2.1.2.3, we obtain

$$\begin{aligned} \bar{Z} &= \frac{[A]_T - [\text{bound H}]/\bar{Z}_H}{[B]_T} \\ &= \frac{[A]_T - ([H]_T - [H] + K_w [H]^{-1})/\bar{Z}_H}{[B]_T} \quad \dots\dots 2.1.2.5 \end{aligned}$$

To calculate the value of [A] at this point, use is made of equation 2.1.2.4 expressed in an equivalent form, i.e.

$$\bar{Z} = \frac{[A]_T - [A] \sum_{j=0}^J [H]^j \beta_{10j}}{[B]_T}$$

Solving for [A],

$$[A] = \frac{[A]_T - \bar{Z}[B]_T}{\sum_{j=0}^J [H]^j \beta_{10j}} \quad \dots\dots 2.1.2.6$$

The program thus presumes no knowledge of the ligand-metal ion species in solution or their stability constants, and computes \bar{Z} and P[A] (equations 2.1.2.5 and 2.1.2.6) solely from a knowledge of the protonation constants of the ligand, the free hydrogen ion concentration from potentiometric data, and the total concentrations of the components.

The equivalent theoretical expression for \bar{Z} is

$$\bar{Z} = \frac{\sum_{j=0}^N j [A]^j \beta_{j10}}{\sum_{j=0}^N [A]^j \beta_{j10}}$$

This shows \bar{Z} to be a function of $[A]$ only. If it is found that for a number of titrations at varying concentrations and ratios that non-overlapping of the formation curves occurs, then one or more of other types of complexes e.g. protonated, hydroxo, oligonuclear etc. are present in the mixture as well.⁶²

2.1.3. PSEUDOPLOT:

This program is essentially a marriage between the programs HALTAFALL and ZPLOT, an idea proposed by D.R. Williams of UWIST, Cardiff University.⁶³ The former computes the equilibrium concentrations of the various constituents in solution, including the free hydrogen ion concentration, by an iterative process using equations 2.1.1.1 to 2.1.1.3. The information required comprises the total concentration of each component, the composition of all possible complexes postulated in a given model to describe the system, and their various stability constants. The refined free hydrogen ion concentrations for each point are then used to calculate \bar{Z} and $P[A]$ using the ZPLOT part of the program, which outputs the 'theoretical' formation curve. A comparison between this and the analogous experimental formation curve serves as a further check as to the validity of a postulated model.

2.1.4. ECCLES:

This program, developed at the University of Cape Town by P.M. May,³⁰ can compute the equilibrium concentrations of metal-ligand complexes in aqueous equilibrium systems at various free

hydrogen ion concentrations and, like HALTAFALL, requires the formation constants for all the complexes and the total or free concentrations of the components present in the mixture.

In a system containing more than two components in addition to hydrogen e.g. two ligands and a metal ion, there is a strong possibility of the formation of ternary complexes. This possibility is enhanced statistically when the probability of such complexes being formed is considered. An additional factor is the influence which energetically favourable (or unfavourable) effects might have on the magnitude of these ternary formation constants as compared to the binary formation constants. ECCLES contains a subroutine whereby these considerations are utilised in the estimation of ternary formation constants, based on values for the binary complexes and on stabilisation factors. The presence of these ternary complexes are then taken into account in the computation of species present at equilibrium.

2.1.5. GEPOLYC.

GEPOLYC⁴⁷ is a program, developed at the University of Dundee, for treating a 1:1 ligand to metal equilibrium mixture, where the ligand has two possible bidentate coordination sites, one at each end of the molecule, and where the allowable species consist of an infinite series of closed chain polymers, the formation constant K of each in the series being defined such that they are equal and independent of the number of units n , and an infinite set of open chain polymers of formation constant K_c defined analogously. The ligand must have four dissociable protons, two at each bidentate site. A unit is considered as one ligand bound to a metal ion at one end. Units are added to the chain one at a time; thus the possible infinite set of all

straight open chain polymers is treated as a subset of the simplest set, the dimer, with one associated chain forming constant. The invariance of the formation constants with n is a reasonable assumption if the concentration of polymers of large n is small and the addition of a moiety at one end is considered to be negligibly affected by what is complexed at the other end due to it being far from the site in question.

It is to be appreciated that polymers become a necessity for the 1:1 methyl dopa-copper(II) system in which the four dissociable protons are neutralised at a pH well below the pKa of the most basic, if hydroxo or six coordinate complexes are to be excluded.

The GEPOLYC treatment was applied successfully by Gorton and Jameson to the dopa-copper(II) system,⁴⁷ and hence its adaptability to the structurally related methyl dopa-copper(II) system was tested.

In the calculations, the parameters \bar{z}' and $[L']$ are introduced as the formation curve parameters. They are experimentally defined as:

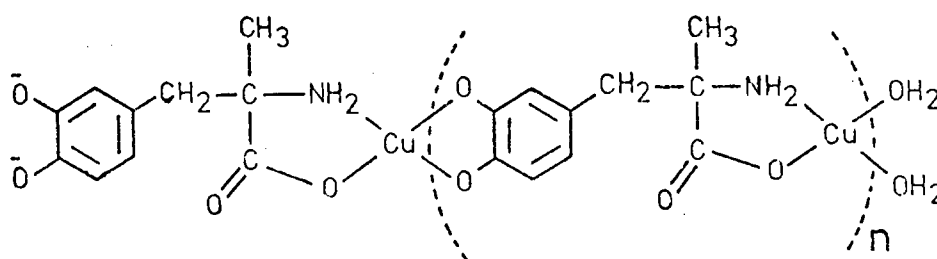
$$\bar{z}' = \frac{[\text{OH}]_T - 2[\text{B}]_T}{2[\text{B}]_T} \quad \dots 2.1.5.1$$

and

$$[L'] = \frac{2[\text{B}]_T - [\text{OH}]_T/2}{\beta'_{102}[\text{H}]^2} \quad \dots 2.1.5.2$$

$[\text{OH}]_T$ is the total concentration of proton removed from the ligand, calculated from a defined point in the titration and obtained from the amount of hydroxide added and the initial mineral acid concentration. $[\text{B}]_T$ is again the total metal ion concentration. \bar{z}' hence represents the fraction of proton pairs replaced by copper(II) from the catechol end of the molecule.

β'_{102} is the second overall formation constant for methyldopa when the fully deprotonated ligand is regarded as being the trianon. $[H]$ is the free hydrogen ion concentration. L' thus has no definite structure, but is related to a series of open chain polymers in which one end is complexed to copper (alanine type), whilst the other is catechol type, deprotonated but uncomplexed (Fig 2.1.5)



$n=0,1,2,\dots$

Fig 2.1.5

In the methyldopa system in the absence of copper(II), the second proton to be added to the fully ionised molecule is attached to the amine moiety and not at a hydroxy position as suggested by the expression above for $[L']$. However, since copper is already assumed to be complexed at the alanine end and $[L']$ is introduced merely to facilitate the calculation, this can be overlooked.

In summary, the functions \bar{z}' and $[L']$ can be visualised as the formation curve parameters for the system after the 1:1 alanine type complex formed initially begins to polymerise through interaction with its catechol end; i.e. at $\bar{z}' = 0$, the ligand exists totally in the form ABH, but progressively begins to form long chain polymers as the titration proceeds and the catechol protons become neutralised.

Theoretical expressions for \bar{Z}' and $[L']$ containing the constants K and K_c will now be derived. The total ligand concentration can be approximated as:

$$[A]_T = \sum_{n=1}^{\infty} n[A_n B_n H_{2-n}] + \sum_{n=2}^{\infty} n[A_n B_n H_{-n}] \quad \dots 2.1.5.3.$$

The first term is the concentration of the open chain polymers, the second the closed. Towards the lower pH values, the total ligand effectively exists as ABH. Under these conditions the uncomplexed ligand is taken to be stoichiometrically negligible. As the pH is raised, so the different polymeric species become significant and contribute towards \bar{Z}' .

From these considerations, it follows that

$$\bar{Z}' [A]_T = \sum_{n=1}^{\infty} (n-1) [A_n B_n H_{2-n}] + \sum_{n=2}^{\infty} n [A_n B_n H_{-n}] \quad \dots 2.1.5.4.$$

Let the formation of a closed chain polymer be represented by K , such that

$$K = \frac{[A_n B_n H_{-n}] [ABH]}{[A_n B_n H_{2-n}] [ABH^*_{-1}]}$$

ABH^*_{-1} is ABH deprotonated twice at the catechol end and its concentration is stoichiometrically zero as appreciable ionisation of the catechol group protons does not occur in the pH region studied.

Let the formation of an open chain polymer be represented by K_c , such that

$$K_c = \frac{[A_n B_n H_{2-n}]}{[A_{n-1} B_{n-1} H_{3-n}] [ABH^*_{-1}]}$$

If both K and K_c are assumed to be independent of n , then in general

$$[A_n B_n H_{2-n}] = (K_c [ABH^*_{-1}])^{n-1} [ABH] \quad \dots 2.1.5.5$$

and

$$[A_n B_n H_{-n}] = (K_c [ABH_{-1}^*])^{n-1} K [ABH_{-1}^*] \quad \dots 2.1.5.6.$$

Continuing, letting

$$X = K_c [ABH_{-1}^*] \text{ and substituting in 2.1.5.3.,}$$

one obtains

$$[A]_T = \sum_{n=1}^{\infty} n X^{(n-1)} [ABH] + \sum_{n=2}^{\infty} n X^{(n-1)} K [ABH_{-1}^*] \quad \dots 2.1.5.7.$$

Similar treatment of equation 2.1.5.4. yields

$$\bar{Z}' [A]_T = \sum_{n=1}^{\infty} (n-1) X^{(n-1)} [ABH] + \sum_{n=2}^{\infty} n X^{(n-1)} K [ABH_{-1}^*] \quad \dots 2.1.5.8.$$

Solving for [ABH] in 2.1.5.7. and substitution in 2.1.5.8.

yields

$$\begin{aligned} \bar{Z}' [A]_T = & \frac{\sum_{n=1}^{\infty} (n-1) X^{(n-1)} \left[[L]_T - \sum_{n=2}^{\infty} n X^{(n-1)} K [ABH_{-1}^*] \right]}{\sum_{n=1}^{\infty} n X^{(n-1)}} \\ & + \sum_{n=2}^{\infty} n X^{(n-1)} K [ABH_{-1}^*] \quad \dots 2.1.5.9. \end{aligned}$$

The numerator of the expression for [L'] (equation 2.1.5.2) refers to the concentration of that series of complexes which still has a proton pair bound at the catechol site. It must therefore be equal to the difference between the total ligand concentration and that fraction of the total ligand concentration which has been deprotonated, i.e.

$$[L'] = \frac{[A]_T (1 - \bar{Z}')}{K' [H]^2} \quad \dots 2.1.5.10.$$

where

$$K' = \frac{[ABH]}{[ABH_{-1}^*] [H]^2} \quad \dots 2.1.5.11.$$

β'_{102} and K' are equal if complexation at the alanine site does not affect the protonation constants at the catechol site.

Substituting for $K' [H]^2$, (equation 2.1.5.11), and $[A]_T (1 - \bar{Z}')$, (equation 2.1.5.4 subtracted from equation 2.1.5.3), in equation 2.1.5.10, one obtains,

$$\begin{aligned}
[L'] &= \frac{([ABH] + [A_2B_2] + [A_3B_3H_{-1}] + \dots) [ABH_{-1}^*]}{[ABH]} \\
&= \frac{\sum_{n=1}^{\infty} X^{(n-1)} [ABH] [ABH_{-1}^*]}{[ABH]} \\
&= \sum_{n=1}^{\infty} X^{(n-1)} [ABH_{-1}^*] \quad \dots 2.1.5.12.
\end{aligned}$$

Experimental values of \bar{Z}' and $-\log[L']$ using equations 2.1.5.1. and 2.1.5.2. are first calculated from potentiometric data. Estimated values of $\log K$ and $\log K_c$ are then submitted to the program GEPOLYC, which outputs the theoretical formation curve parameters (equations 2.1.5.9. and 2.1.5.12). These values are adjusted until the best fit with the experimental curve is achieved. The program allows the maximum size of the closed chain species to be predetermined, but this is not possible with the open chain species as the relevant summation expression is in an explicit form.

The concentration of any particular species can be calculated at any point in the titration.

From equation 2.1.5.12,

$$\begin{aligned}
[L'] &= \frac{[ABH_{-1}^*]}{1-X} = \frac{[ABH_{-1}^*]}{1-K_c [ABH_{-1}^*]} \\
\therefore [ABH_{-1}^*] &= \frac{[L']}{1 + [L'] K_c} \quad \dots 2.1.5.13.
\end{aligned}$$

From equations 2.1.5.7 and 2.1.5.8,

$$\begin{aligned}
[A]_T (1 - \bar{Z}') &= \sum_{n=1}^{\infty} X^{(n-1)} [ABH] \\
\therefore [ABH] &= [A]_T (1 - \bar{Z}') (1 - X) \quad \dots 2.1.5.14.
\end{aligned}$$

$[ABH_{-1}^*]$ and $[ABH]$ may be calculated from equations 2.1.5.13 and 2.1.5.14 respectively. Hence $[A_n B_n H_{2-n}]$ and $[A_n B_n H_{-n}]$ may be calculated using equations 2.1.5.5. and 2.1.5.6.

2.1.6. MAGEC.

MAGEC is a program developed at this laboratory to calibrate the $[H^+]$ sensitive glass electrodes used in this work.⁶⁴ It uses as input data ml and mV readings obtained from the titration of a weak monoprotic acid with a strong base. Estimates of the seven parameters necessary to define the system are then used to solve for the value of base added at each point, V'_{calc} . The seven parameters are: β_H , the protonation constant of the acid; F_n and E° , the slope and intercept respectively of the Nernst equation (equation 2.4.1); K_w , the ionic product of water at the background ionic strength; $[AH]_0$, the initial concentration of the acid used; $[NaOH]$, the concentration of the titrant; and $[V]_0$, the initial volume of titrate. Those parameters most likely to contain errors in their estimates are adjusted iteratively by the simplex routine in minimising the sum of the squares of the individual residuals, $\sum_{i=1}^n (V'_{calc} - V'_{obs})^2$. A scan of the sense of the residuals for systematic trends provides a check on the validity of the assumption that equation 2.4.1 is linear for the electrode used.

All the programs were run in FORTRAN V on the University of Cape Town Univac 1106.

2.2. Applications of ultraviolet/visible spectrophotometry.

In general, solution potentiometry can provide useful information as to the stoichiometry of the complexes formed and the strengths of the bonds formed in any ligand-metal ion interactions. However, structural information such as bonding sites must generally be obtained by other methods. The structure of methyl dopa is such that if it could be cleaved at the position shown in Fig 2.2.1, the one half would be identical to catechol whilst the other would closely resemble alanine.

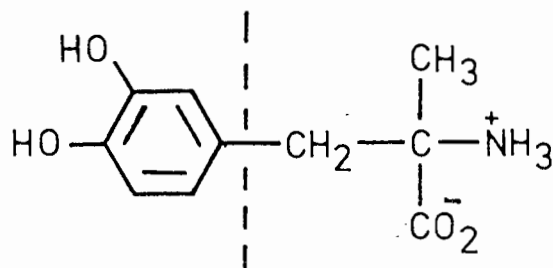


Fig 2.2.1

Hence given an absorption spectrum of a methyl dopa-copper(II) solution which shows strong similarities to a spectrum obtained from an alanine-copper(II) solution under the same conditions, one could with some justification presume that the copper atom giving rise to the absorption maxima in the two solutions exists in similar chemical environments. An analogous deduction must follow for similarities in the spectra of a methyl dopa-copper(II) solution and a catechol-copper(II) solution. An intermediate spectrum of a methyl dopa-copper(II) solution which contained characteristics of both 'pure' spectra hence would indicate both types of bonding to

be present. Clearly the more distinct and simple the spectra arising from the different chemical environments, the easier it would be to estimate copper(II) populations at the two complexing sites in question.

2.3. Application of nuclear magnetic resonance (NMR).

The pH dependence in the chemical shift of a proton in close proximity to a dissociable proton can in cases be used to give a rough estimate of the pKa of the dissociable proton. Hence that region where the observable rate of change of chemical shift with pH is a maximum can be taken to approximately equal the pKa of the nearest group with a dissociable proton, for at this point the chemical surroundings influencing the shielding of the proton nucleus being observed is changing most rapidly.

The technique described was attempted in the determination of the protonation sites of the methyl dopa dianion (Fig 5(2)).

2.4. Potentiometric approach : apparatus and materials.

The apparatus as shown in Fig 2.4.1. was used to titrate solutions containing various concentrations of ligand, copper(II) perchlorate and perchloric acid or sodium hydroxide against either sodium hydroxide or perchloric acid, the ionic strength being maintained at 0,15 M ClO_4^- by added sodium perchlorate. The reaction was followed using a combination of a Russell pH Ltd. glass electrode (SF 75/C14) and a calomel electrode in which the KCl had been replaced by NaCl. The reference electrode and the titration vessel (Metrohm Cat. No. EA876-20) were thermostated at $25^\circ \pm 0,1^\circ\text{C}$ using a Lauda (Type K4RD) thermostat. Burettes (Metrohm Cat.No. E274) and

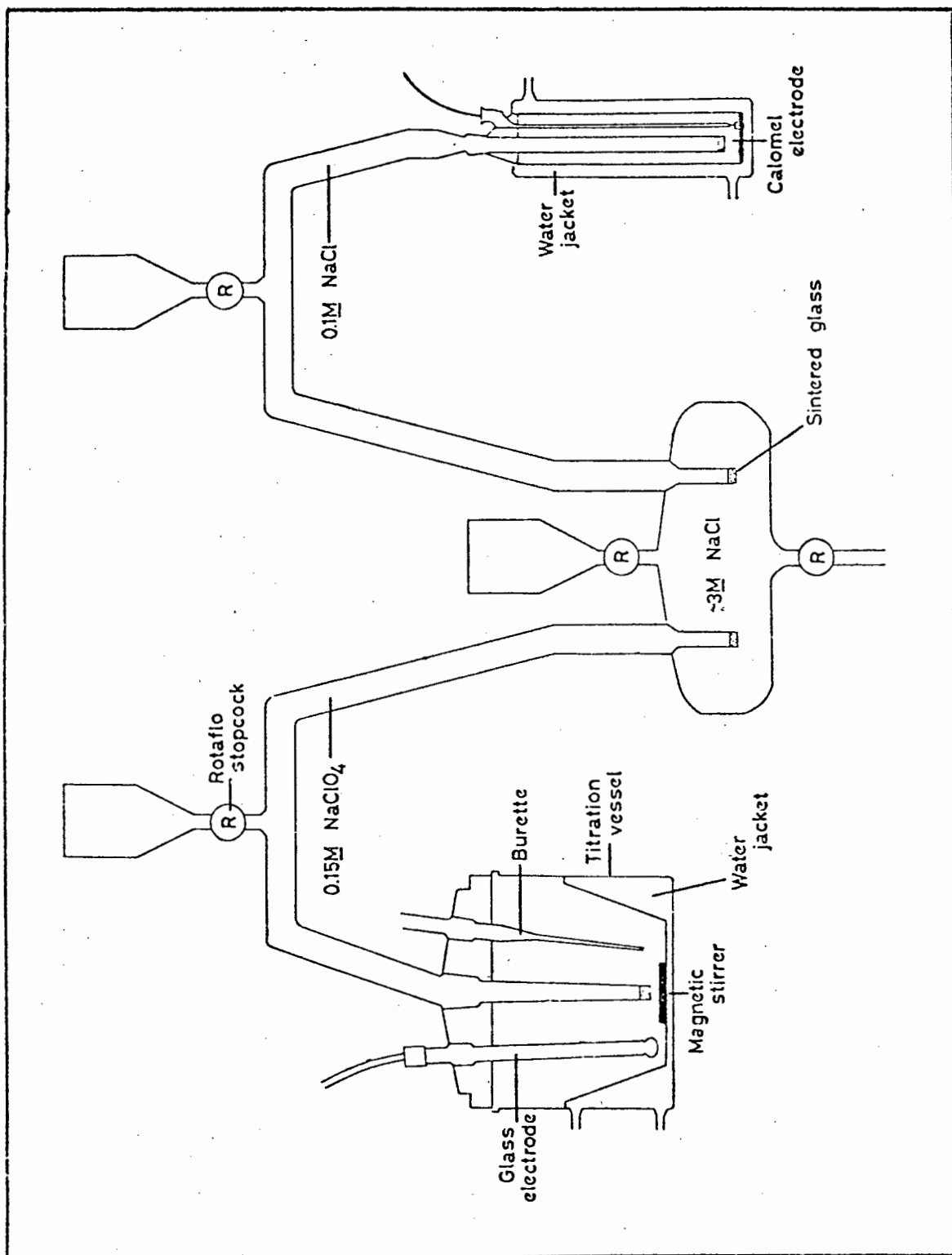


Diagram of apparatus used in the determination of formation constants. The nitrogen bubbler, thermometer and potentiometer are not shown.

Fig 2.4.1

pipettes were calibrated before use. The e.m.f. readings were taken using a Radiometer PHM 64 digital pH-meter. The construction and composition of the salt bridge were such as to minimise diffusion effects yet facilitate the renewal of the liquid junction potentials between titrations. The high purity nitrogen (AFROX) under which the reactions were studied was further purified by bubbling through a sequence of gas wash bottles before being admitted to the titration vessel.

- (i) 50% KOH to remove CO_2 ;
- (ii) 15 g pyrogallol in 100 ml 50% NaOH to remove O_2 ; ⁶⁵
- (iii) lead wool immersed in an alkaline (200 g dm^{-3} KOH) 1% sodium 1,2-naphthoquinone-4-sulphonate solution to remove O_2 ; ⁶⁶
- (iv) an excess of zinc amalgam in contact with an acidic (50 ml dm^{-3} conc. H_2SO_4) chromous/chromic sulphate (300 g dm^{-3} $\text{Cr}_2(\text{SO}_4)_3 \cdot 15\text{H}_2\text{O}$) solution to remove O_2 ; ⁶⁶
- (v) an empty wash bottle;
- (vi) the support electrolyte ($0,15 \text{ M NaClO}_4$) thermostated at 25°C .

Back diffusion after passing through the titration vessel was prevented by releasing the nitrogen to the atmosphere via an ionic background solution trap.

Glass distilled CO_2 -free water was used in the preparation of all solutions. NaClO_4 (Merck) was further purified by filtering through a $8,0 \mu$ PVC membrane (Sartorius). The pH was adjusted to 12 and left for 3 days to precipitate any metal impurities present, and then refiltered through a $0,45 \mu$ membrane. Acidification and boiling to remove CO_2 then followed. After cooling, the solution was finally adjusted to pH 7. The NaClO_4 concentration was determined gravimetrically by evaporating a pipetted volume to dryness under vacuum. The

$\text{Cu}(\text{ClO}_4)_2$ (K and K Laboratories) stock solution was standardised by EDTA titration.⁶⁷ The NaOH was prepared from ampoules (Merck) and standardised with potassium hydrogen phthalate. HClO_4 (Merck) concentration was determined by titration with the NaOH and checked using borax which had been recrystallised and stored over a sugar/salt solution. Methyldopa (Fluka, Puriss) was used as supplied (Found : C, 50,3; H, 6,8; N, 6,0. $\text{C}_{10}\text{H}_{13}\text{O}_4\text{N}\cdot 1\frac{1}{2}\text{H}_2\text{O}$ requires C, 50,4; H, 6,7; N, 5,9%). Catechol (Fluka, Puriss) after being triply sublimed and alanine (Sigma Chemical Co.) were found to be >99% pure. The ligand was weighed directly into the titrating vessel containing 20 ml of pipetted acidified $\text{Cu}(\text{ClO}_4)_2$ solution at the required concentration before each titration. The mixture was allowed to equilibrate for $\frac{1}{2}$ hour before titrating was commenced. Reproducibility in titrations was improved if the glass electrodes were allowed to 'recover' after a titration by soaking them in an acidified (0,1 M HClO_4) solution at the ionic background concentration before being used again. Hence two electrodes were used in rotation in the experiments.

The $[\text{H}^+]$ value at 25°C was calculated according to the equation,

$$E = E^\circ + 59,16 \log [\text{H}^+] \quad \dots 2.4.1.,$$

E is a function of the potential difference in mV between the glass and reference electrodes as read by the potentiometer, and varies according to the difference in the concentration of hydrogen ions on either side of the glass electrode. E° is a constant dependent on the apparatus design. Before each titration the value of E° was evaluated using a solution of known $[\text{H}^+]$ (section 3.3), and the linear relationship

(equation 2.4.1) was assumed to hold over the complete pH range studied. Hence mV readings versus known additions of titrant were obtained and used with the E° value as input data to the computer programs employed in the model fitting.

3. CALIBRATION AND ASSESSMENT OF POTENTIOMETRIC TECHNIQUE.

Before investigating an unknown system using untested apparatus, it is necessary to analyse known systems and compare the results obtained with those from the literature. Agreement allows one to place confidence in any readings one might obtain from future experiments. With this in mind, potentiometric titrations using our apparatus were carried out on solutions of acetic acid, glycine-copper(II) and perchloric acid at 0,15 M NaClO₄ and 25°C, the purpose being to:

- (i) test the linearity of the glass electrodes used in the pH range to be studied;
- (ii) assess the reproducibility of the apparatus;
- (iii) obtain formation constants for the relevant systems, literature values for which are well established, and in so doing to calibrate the apparatus.

3.1. Acetic acid protonation constant.

Solutions of acetic acid (Merck) varying in concentration from 0,005 M to 0,05 M were titrated potentiometrically with NaOH to a pH of 5.5. The results were processed using the program MAGEC (see section 2.1.6).

A FIX label required that the value given be held constant throughout the calculation. FIND means that the variable is refined. FREE means the value is initially fixed but is permitted later to vary along with the FIND variables.

The optimised [HOAc]₀ values (Table 3.1.1) agreed favourably with those obtained by a Gran plot⁶⁸, these being 0,00592 M, 0,00999 M and 0,0503 M respectively. The protonation constant of acetic acid at 25°C in 0,2 M KNO₃ has been reported by Feldman and Koval⁶⁹. An estimation of its

value at 0,15 M ionic strength was calculated using the Davies relationship (equation 4.6.1) and found to be $4,22 \times 10^4$. From the study, the linearity of the electrodes and the reproducibility of the apparatus were found to be satisfactory.

Table 3.1.1.

Optimised parameter values for acetic acid titration (0,15 M NaClO ₄ , 25°C)				
E°	317,6	316,4	315,4	FIND
β ^H	$3,3807 \times 10^4$	$3,3947 \times 10^4$	$3,2825 \times 10^4$	FREE
Slope	59,16	59,16	59,16	FIX
Kw	$1,73 \times 10^{-14}$	$1,73 \times 10^{-14}$	$1,73 \times 10^{-14}$	FIX
[HOAc] ₀	$5,9204 \times 10^{-3}$	$9,9416 \times 10^{-3}$	$5,0172 \times 10^{-2}$	FIND
[NaOH]	$1,988 \times 10^{-2}$	$1,988 \times 10^{-2}$	$3,009 \times 10^{-1}$	FIX
[V] ₀	20,0	20,0	20,0	FIX

3.2. Glycine-proton and glycine-copper(II) formation constants.

Formation constants of glycine (BDH) were determined by titrating 0,05 M ligand solutions in 0,05 M HClO₄ with 0,03 M NaOH in the absence of metal ions up to a pH of 10. The refined values (MINIQUAD) appear in Table 3.2.1. The first protonation constant is assigned to the amine nitrogen of glycine, the second to the carboxyl group.

The glycine-copper(II) stability constants were determined by titrating 0,05 M glycine, 0,05 M HClO₄ and 0,005 M Cu(ClO₄)₂ with 0,03 M NaOH. The protonation constants included in the refinement (MINIQUAD) were those in Table 3.2.1, and were not permitted to vary. The results appear in Table 3.2.2.

Table 3.2.1.

Logarithms of protonation constants for glycine (0,15 M NaClO ₄ , 25 °C) No. of titration points = 129 R = 0,0018				
Complex	log β_{por}	log K_{por}	std.dev.in log β_{por}	log β_{por} (lit) ⁷⁰
AH	9,536	9,536	0,007	9,62
AH ₂	11,912	2,376	0,011	12,06

The small value of log K_{310} as compared to log K_{110} and log K_{210} , corresponding to a third ligand molecule complexing to the metal, is consistent with the prediction that copper(II) accepts a third bidentate ligand relatively unwillingly in rearranging itself from a square planar coordination geometry into a distorted octahedral environment (Jahn-Teller effect).

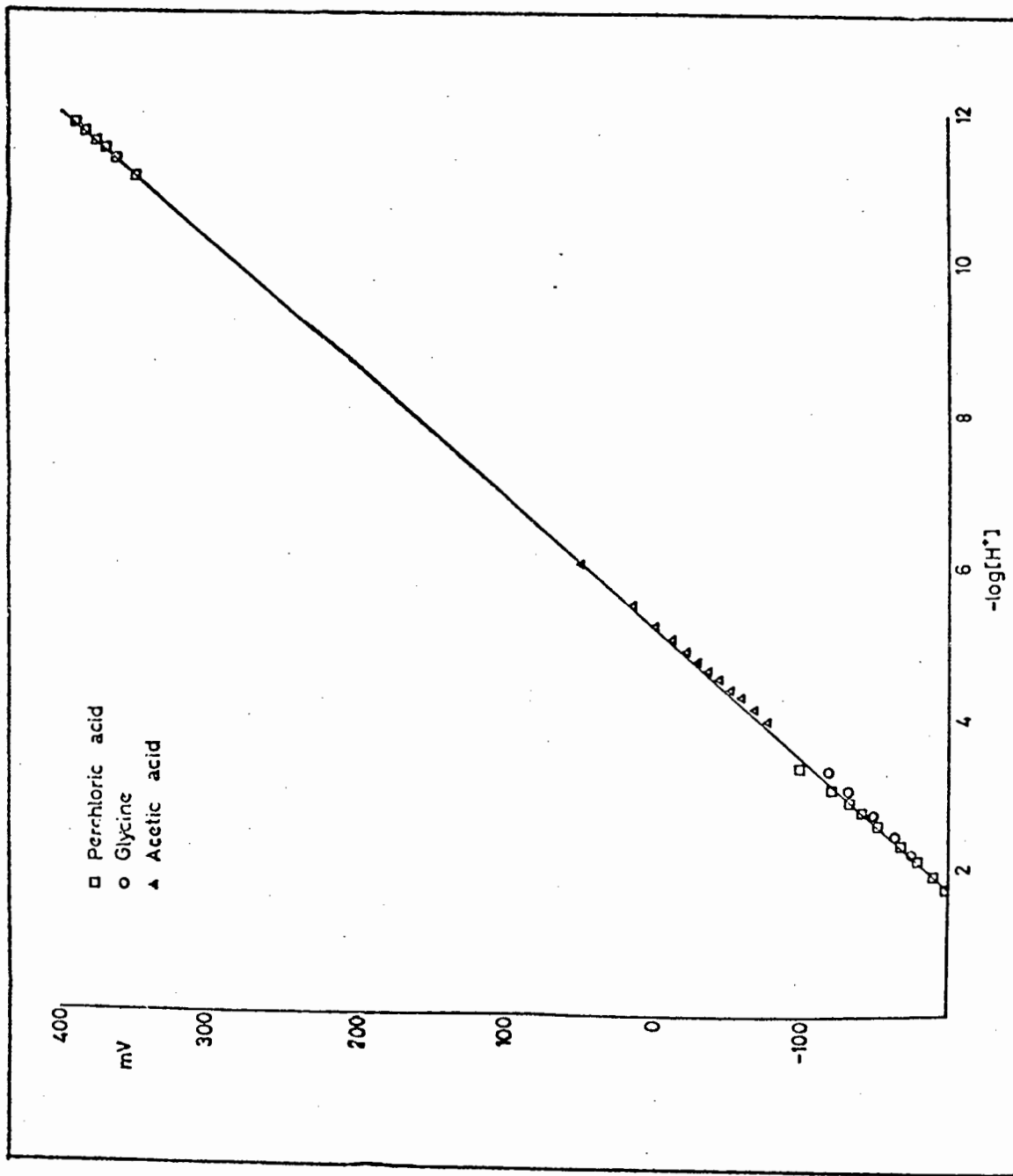
Table 3.2.2.

Logarithms of formation constants for the glycine- copper(II) system (0,15 M NaClO ₄ , 25 °C) No. of titration points = 109 R = 0,0033				
Complex	log β_{pgo}	log K_{pgo}	std.dev.in log β_{pgo}	log β_{pgo} (lit) ⁷⁰
AB	8,250	8,250	0,021	8,18
A ₂ B	15,072	6,822	0,055	15,02
A ₃ B	16,300	1,228	0,107	-

3.3. Calibration of the apparatus.

From mass balance and electrical neutrality considerations, $[H^+]$ can be calculated at any point in the titration of a weak acid by a strong base, provided the dissociation constant of the weak acid is known under the conditions of the titration. $-\log[H^+]$ can then be plotted against the potentiometer readings obtained and the result checked to see whether equation 2.4.1 is obeyed. An example of such calculations on readings obtained from titrations of glycine, acetic acid and perchloric acid with NaOH using one electrode can be seen in Fig 3.3.1. The acids were chosen so as to incorporate as wide a range as possible in $-\log[H^+]$. The linearity of the electrode used is evident.

The method for determining E° in a titration in which $[H^+]$ was unknown is now described. A buffer solution of known $[H^+]$ made up of acetic acid and NaOH in 0,15 M $NaClO_4$ was placed in the titration vessel and the potentiometer reading noted using a glass electrode whose linearity over the range to be studied had been established employing the methods described above. Equation 2.4.1. was solved for E° , a value that was then used in the titration to follow in the calculation of $[H^+]$ at each point. The procedure was repeated before each subsequent titration with the electrode, since E° can be expected to vary slightly due to the renewal of the liquid junction carried out between titrations and changes in the potential of the glass electrode membrane.



Electrode calibration

Fig 3.3.1

4. STUDY OF METHYLDOPA-PROTON AND METHYLDOPA-COPPER(II) EQUILIBRIA: RESULTS

4.1. Ultraviolet/visible spectrophotometric results.

All spectra were recorded in 10mm cuvettes at 25°C and $I = 0,150 \text{ mol dm}^{-3} \text{ Na}^+ \text{ClO}_4^-$ against water as a reference using a Beckman DK2A recording spectrophotometer. Spectra (1000nm - 350nm) at progressively increasing pH values were obtained from solutions in which the total ligand concentration ranged from 0,088 M to 0,010 M, the ligand:copper(II) ratio varying from 6:1 to 1:1. With the stated aim in mind (section 2.2), the analysis was initially directed more at positional maxima in the spectra than their absolute absorbance.

(i) Alanine-copper(II):

Spectra of a solution at various pH values in which the total concentration of alanine was 0,010 M and that of copper(II) was 0,004 M are shown in Fig 4.1.1. The absorption maximum at pH 4 shows a hypsochromic shift with an increase in intensity as the pH is brought up to 6. Above this value the band remains essentially constant at $\approx 610 \text{ nm}$, the variation in absorbance being non-uniform. An asymmetric absorption maximum in the range 600 to 700 nm is typical of the d-d transitions found in copper(II) complexes.^{34,71,72}

(ii) Catechol-copper(II):

Fig 4.1.2 shows the spectra obtained at various pH values from a solution containing initially 0,010 M catechol and 0,004 M copper(II) respectively. Towards the high pH region, the spectra consist of two broad bands. One band centred at about 700 nm at pH 6, first increases and then decreases slightly in intensity with a hypsochromic shift as the pH is raised, and is

probably of the type mentioned above involving the d-d transitions of copper(II). The other band at approximately 415 nm increases in intensity in undergoing the same pH change, the position of the absorption maximum remaining essentially constant. Charge transfer bands in this latter region are evidently characteristic of copper(II) - O,O bonds⁷³.

(iii) Methyldopa-copper(II):

The spectra at different pH values for a solution in which both the methyldopa and copper(II) concentrations were 0,010 M is shown in Fig 4.1.3. The spectra appear to be an approximate overlap of those described above. The conclusion hence is that below pH4, copper(II) is predominantly bound to the alanine-type moiety of methyldopa, with complexation at the catechol-type site being minimal. As the pH increases, binding to both the alanine and catechol ends becomes evident. Due to the complicated pH dependence of the absorbances, however, a detailed estimate of the relative coordination distribution appears to be not feasible.

An interesting observation detected during the methyldopa-copper(II) spectra measurements was the increased competitiveness for the catechol site of the ligand by copper(II) as the concentration of the components was increased, an effect paralleled for decreasing values of the ligand:metal ion ratio at constant ligand concentration. This was made apparent by the lower pH at which the band at 415 nm first showed itself in these solutions.

Spectra of alanine-copper(II) complexes. Concentrations
(10^{-3} mol dm^{-3}) of A and B are 10,0 and 4,0.
Reference, water.

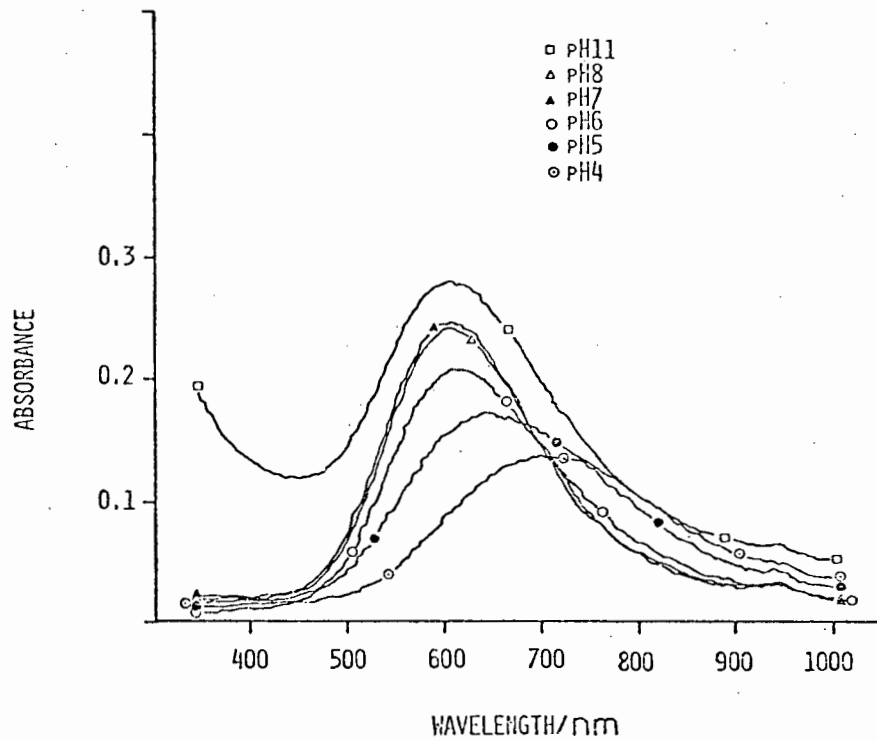


Fig 4.1.1

Spectra of catechol-copper(II) complexes. Concentrations
(10^{-3} mol dm^{-3}) of A and B are 10,0 and 4,0.
Reference, water.

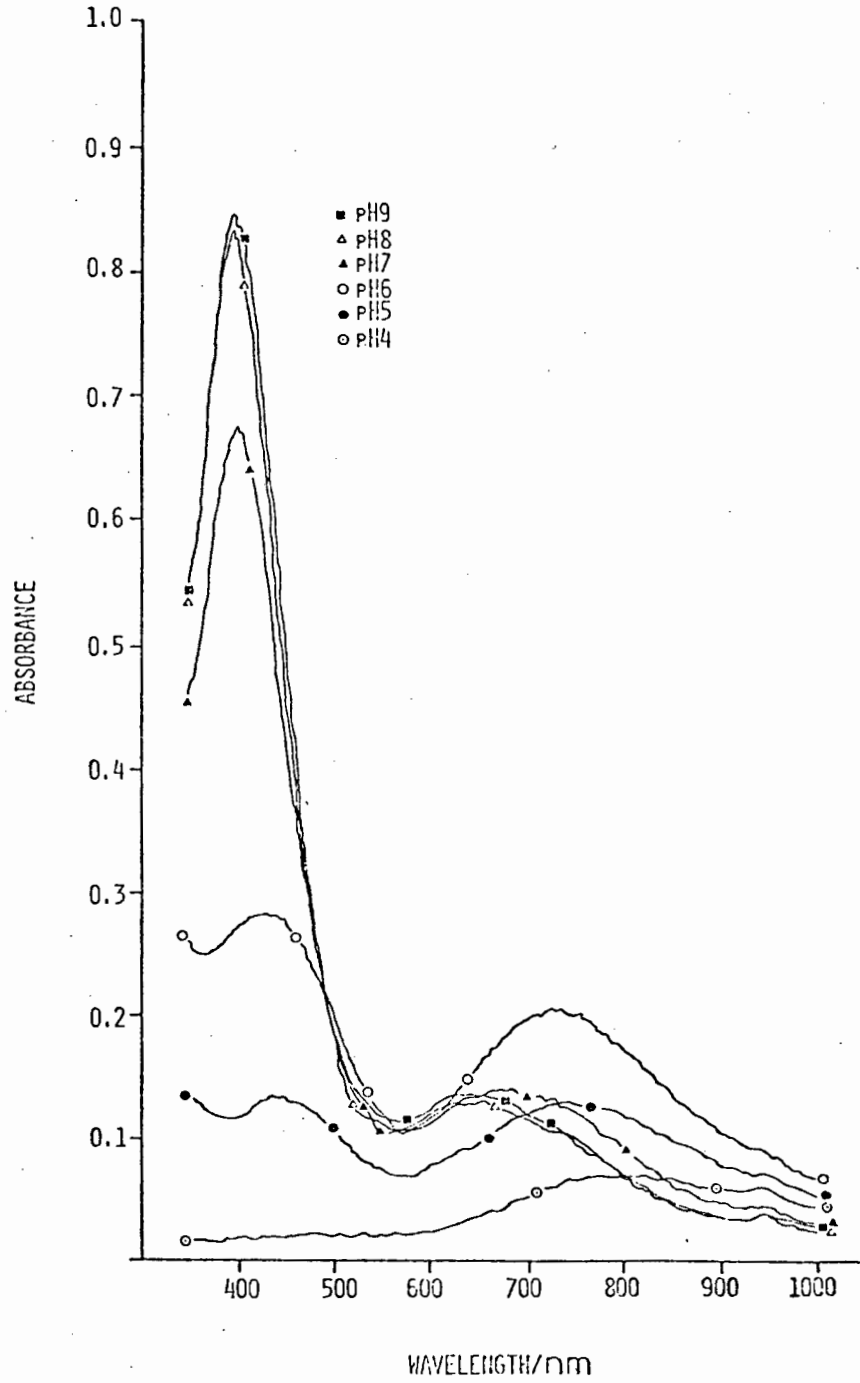


Fig 4.1.2

Spectra of methyl dopa-copper(II) complexes. Concentrations
(10^{-3} mol dm^{-3}) of A and B are 10,0 and 10,0.

Reference, water.

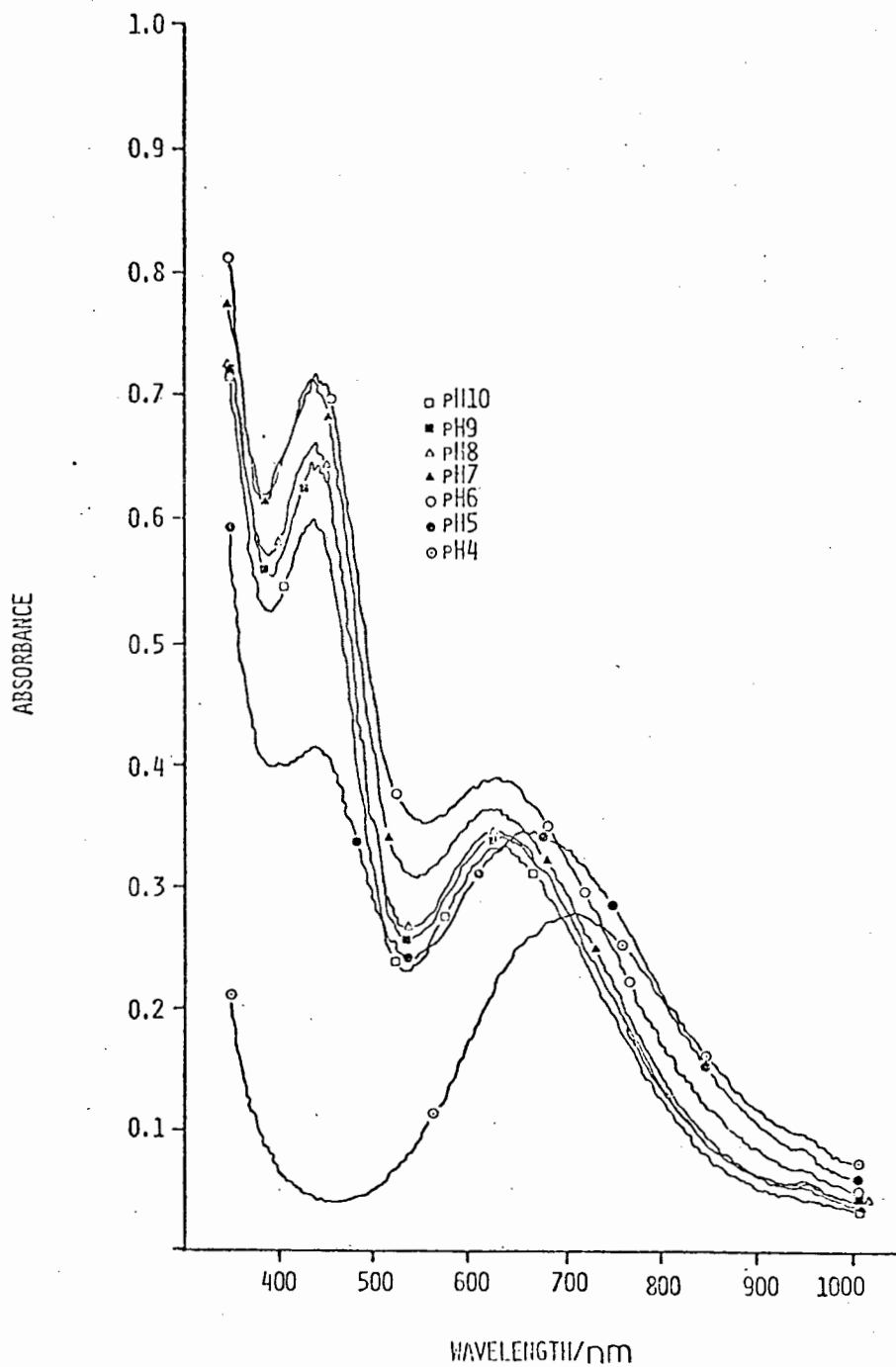


Fig 4.1.3

4.2. NMR results.

Spectra were recorded at 90 MHz on a Bruker WH-90DS spectrometer operating in the Fourier transform mode. Ligand solutions (0,05 mol dm⁻³) in ²H₂O were adjusted to the required pH with NaO²H and ²HCl. The pH of the solutions were calculated from pD = pH + 0,4. The sodium salt of 3-trimethylsilyl propane sulphonic acid was used as internal reference.

The chemical shifts for the protons on C(3), C(2)-CH₃, C(2'), C(5') and C(6') were measured as a function of pH in the methyldopa molecule (Fig 5(1)). The pH corresponding to the maximum rate of change of chemical shift for C(3) and C(2)-CH₃ was observed in the range 9,80-9,96, and corresponds to the approximate pKa of the protonated amino group. For the aromatic protons the apparent pKa was in the range 9,1-9,2.

It has been found that in the phenol/phenoxide and substituted phenol systems in which there is a unique site of deprotonation, the protons in the ortho and para positions show much larger upfield shifts than the meta protons.^{74,75} In the methyldopa molecule however, the aromatic proton shifts are: C(2') 20 Hz, C(5') 24 Hz and C(6') 26 Hz. As an explanation it appears that the remaining proton on the catechol mono-anion moiety either is in rapid exchange between two sites or is held in a 'symmetric' hydrogen bonded bridged structure.

A NMR study consisting of T₁ experiments⁷⁶ at pH 4 and pH 8,5 with ligand:metal ion ratios in the vicinity of 100:1 failed to provide further information as regards the pH dependence of copper(II) bonding at the two coordination sites of methyldopa. This arose due to the fact that the changes in

T_2 were very much faster than those in T_1 . The consequent overlapping of the aromatic resonances made quantitative measurement impossible.

4.3. ECCLES results.

In order to determine the feasibility of 'mixed bonding' occurring in the methyldopa-copper(II) system involving both N,O and O,O bonds, a model consisting of copper(II) in solution with molecules in which the potential donor groups of methyldopa occur in different ligands was inputted to the program ECCLES. The ligands used were alanine and catechol. The treatment necessitated a knowledge of the protonation and copper(II) stability constants of the ligands concerned. These were determined potentiometrically by the same method used for methyldopa. Titration data were in each case submitted to the program MINQUAD for refinement of the β 's, the final values for which appear in Tables 4.3.1 and 4.3.2 respectively.

In the ECCLES calculations, total alanine concentration was initially equal to total catechol concentration at 0,02 M. Total copper(II) initial concentration was 0,004 M. Employing a stabilisation factor of 0,40³⁰, the stability constant of the ternary alanine-copper(II)-catechol complex as computed by ECCLES was 19,65. The distribution with pH of complexes as calculated by ECCLES is shown in Fig 4.3.1. The designation Ala-Cu(II) on the graph includes both the 1:1 and 2:1 alanine-copper(II) complexes, as does Cat-Cu(II) in the case of the catechol-copper(II) complexes. The significant contribution of the ternary complex towards the total bound copper(II) at pH 7,5 substantiates the proposal that a 'mixed' mode of complexation is present in the methyldopa-copper(II)

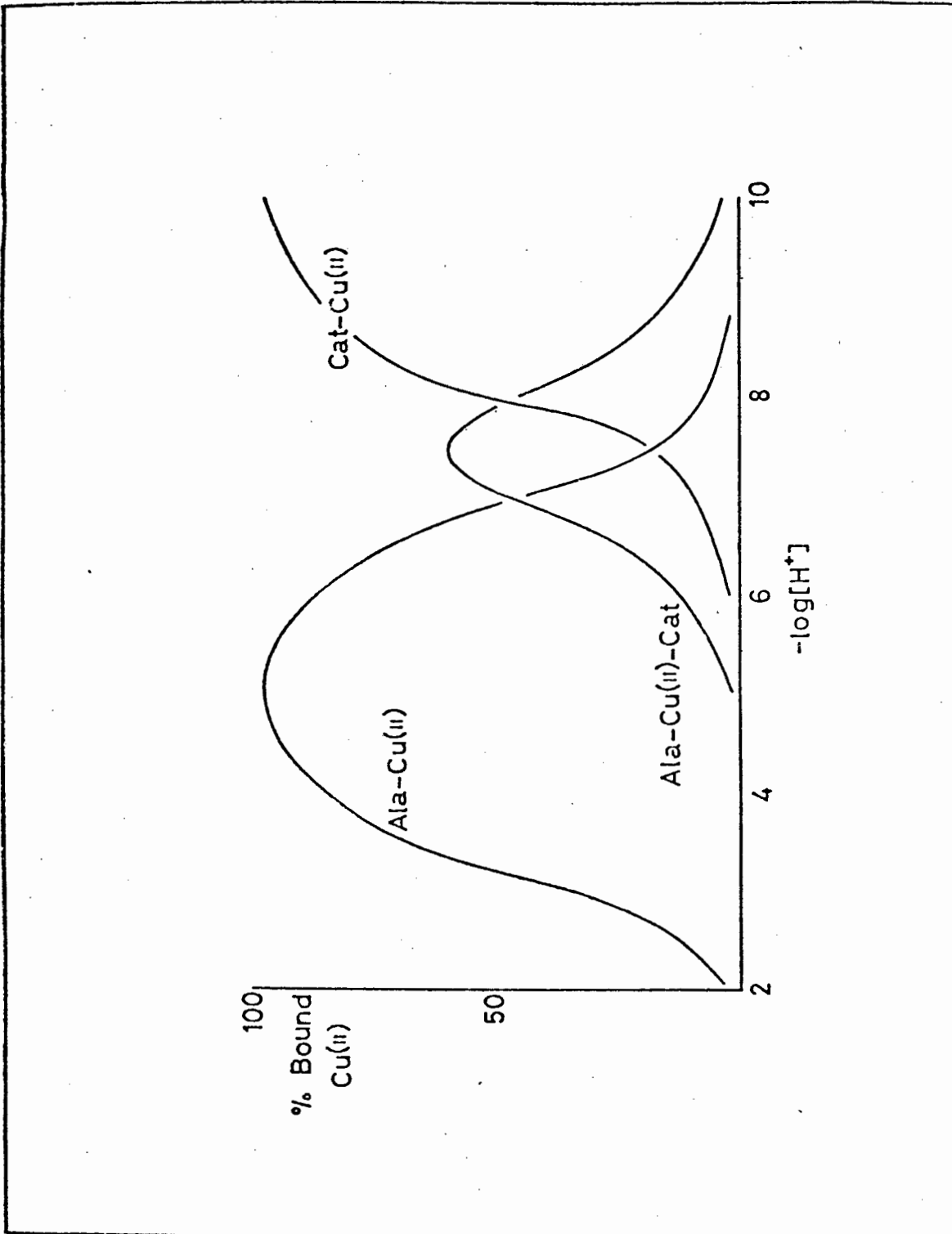
system in the same region. Also noteworthy is the obvious preference of copper(II) for N,O type bonding at low pH values and O,O type bonding at high pH values, given equal concentrations of both options.

Table 4.3.1.

Logarithms of protonation and formation constants for the alanine-copper(II) system (0,15 M NaClO ₄ , 25°C) No. of titration points = 78 R = 0,0062			
Complex	log β_{pqr}	std. dev. in log β_{pqr}	log β_{pqr} ⁷⁷ I = 0,1 M NaClO ₄
AH	9,659	0,007	9,83
AH	11,970	0,032	12,09
AB	8,128	0,072	8,25
A ₂ B	14,956	0,062	15,30

Table 4.3.2.

Logarithms of protonation and formation constants for the catechol-copper(II) system (0,15 M NaClO ₄ , 25°C) No. of titration points = 61 R = 0,0145			
Complex	log β_{pqr}	std. dev. in log β_{pqr}	log β_{pqr} ⁷⁷ I = 0,10 M NaClO ₄
AH	9,091	0,055	9,32
ABH ₋₁	0,835	0,076	0,96
A ₂ BH ₋₂	-1,041	0,159	-0,97



Distribution of complexes in alanine-copper(II)-catechol system
calculated by ECCLES.

Fig 4.3.1

4.4. Methyldopa protonation constants.

The logarithms of the overall protonation constants of methyldopa as determined by MINIQUAD appear in Table 4.4.1. The experimental (ZPLOT) and theoretical (PSEUDOPLOT) formation curves appear in Fig 4.4.1.

Table 4.4.1.

Logarithms of the overall protonation constants of methyldopa (0,15 M NaClO ₄ , 25°C) R = 0,0123				
Complex	Site	log β_{por}	std.deviation in log β_{por}	No. of exp. observations
AH	NH ₂	9,982	0,005	289
AH ₂	O ⁻	18,886	0,006	439
AH ₃	CO ₂ ⁻	21,104	0,014	288

The refinement was carried out on data points obtained from forward and backward titrations varying in initial ligand concentration from 0,008 M to 0,037 M, up to a pH of approximately 11,6. Slight oxidation effects, made evident by the discolouration of the titration mixture in the high pH range, proved to be insignificant in that the formation curves for both forward and backward titrations overlapped almost exactly. It was found impractical, considering the comparatively low background electrolyte ionic strength used in this work, to determine potentiometrically the protonation constant of the trianion of methyldopa due to its expected value lying close to the ionic product of water. (However, see Section 4.6). It has been suggested that the very strong

PSEUDOPLOT curves calculated from the methyl dopa-proton values in Table 4.4.1 plotted on experimental ZPLOT points.

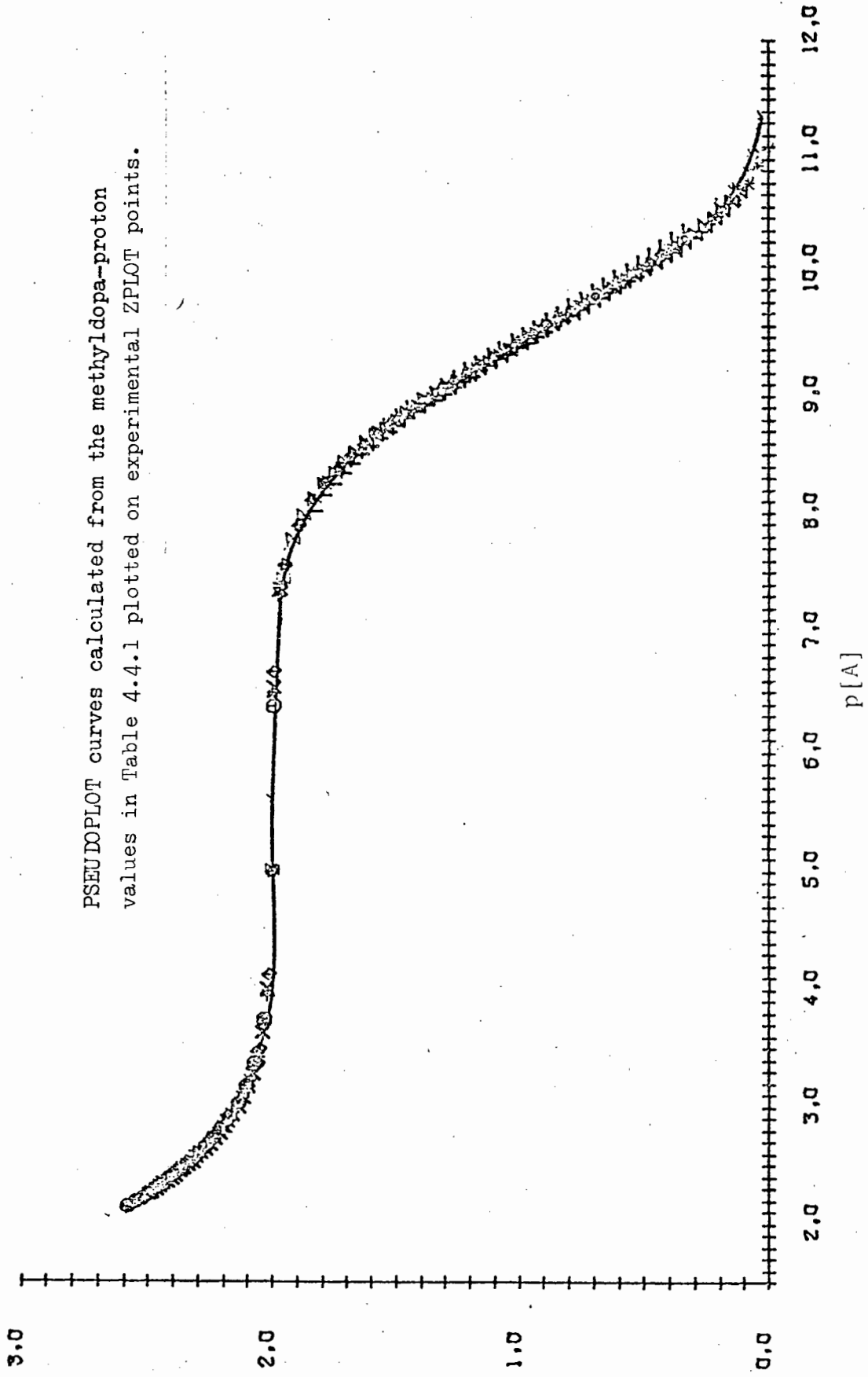


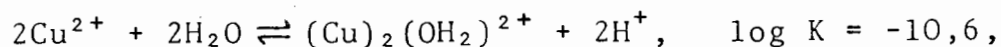
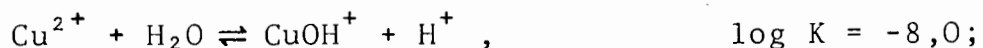
Fig 4.4.1

basic nature of one of the catechol oxygen atoms is caused by hydrogen bonding between the hydroxyl and phenoxide groups.⁷⁸ The ligand A unless otherwise stated hence refers to the doubly charged methyldopa anion resulting from the ionisations of the carboxylate group, one phenolic group and the amino group (Fig 5(2)). The assignments of the formation constants to their respective protonation sites by NMR measurements is described in Section 4.2.

4.5. Methyldopa-copper(II) formation constants (MINIQUAD):

Table 4.5.1 shows the most consistent set of complexes found, together with the respective logarithms of the formation constants, as refined by MINIQUAD. The validity of the model is illustrated by the excellent matching of the experimental $\bar{Z}_{\text{obs}}/p[A]$ points in Fig 4.5.1 to the corresponding theoretical formation curves computed by PSEUDOPLOT, using the formation constants in Table 4.5.1. Fig 4.5.2(a)-(f) are the HALTAFALL plots of the concentrations of complexes as a function of pH for the 10:1, 2½:1 and 1:1 ligand: metal ion ratios studied. The 5:1 distributions are similar to the 10:1 and are not shown.

The MINIQUAD computations were carried out on a series of forward and backward titrations varying in total reactant concentration from 0,004 to 0,037 mol dm⁻³. In all refinements the protonation formation constants of the ligand were fixed and not allowed to vary, and the hydrolysis reactions,⁸⁵



were incorporated into every trial. The high degree of non-overlapping together with the fanning back pattern observed in the experimental formation curves for the various total

concentrations of ligand and metal (Fig 4.5.1) indicate that not only mononuclear binary but also other types of complex, e.g. protonated, hydroxo and polynuclear, are formed. Attempts to refine all ratios together failed (see chapter 5), and this necessitated successive division and grouping of the titration data in various ways according to the ligand:metal ratios and the range of pH in order to obtain a consistent chemical model to explain the entire set of potentiometric data. Thus the final formation constants of the species ABH , A_2BH_2 and ABH_2 were obtained through refinement using titration data points from all ratios, but restricted to those corresponding to a pH less than 3,5. In subsequent refinements at higher pH values when additional complexes were included in the model, β_{111} , β_{212} and β_{112} were fixed together with the ligand protonation and hydrolysis constants and not varied in the refinement process.

Referring to Fig 4.5.2(a), the 10:1 (and 5:1) ratio solutions in particular are characterised by complexes corresponding to successive deprotonations of the species A_2BH_2 , with no polynuclear species present.

The presence of a series of open chain polymers and a cyclic dimer in the 1:1 ligand:metal solutions was indicated using results obtained from the program GEPOLYC (see Section 4.6) and substantiated through successful refinement of this model with the program MINIQAD. Based on the GEPOLYC results, the concentration of the open chain polymers $A_nB_nH_{2-n}$ becomes negligible for $n \geq 5$. In agreement with this is the finding from MINIQAD calculations that the linear dimer, the trimer and the tetramer are the only significant representatives of these polymers in the 1:1 ratio solutions.

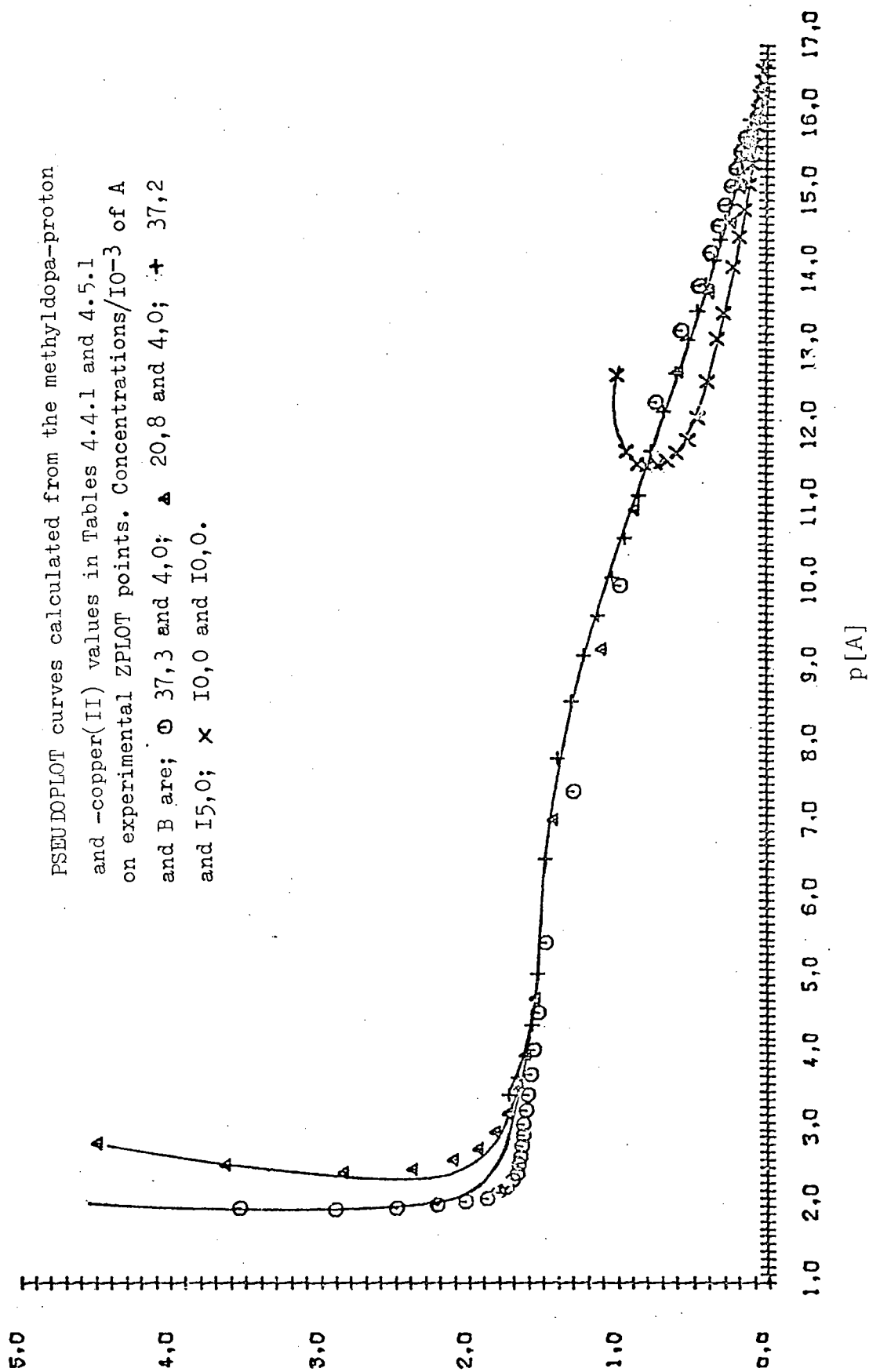


Fig 4.5.1

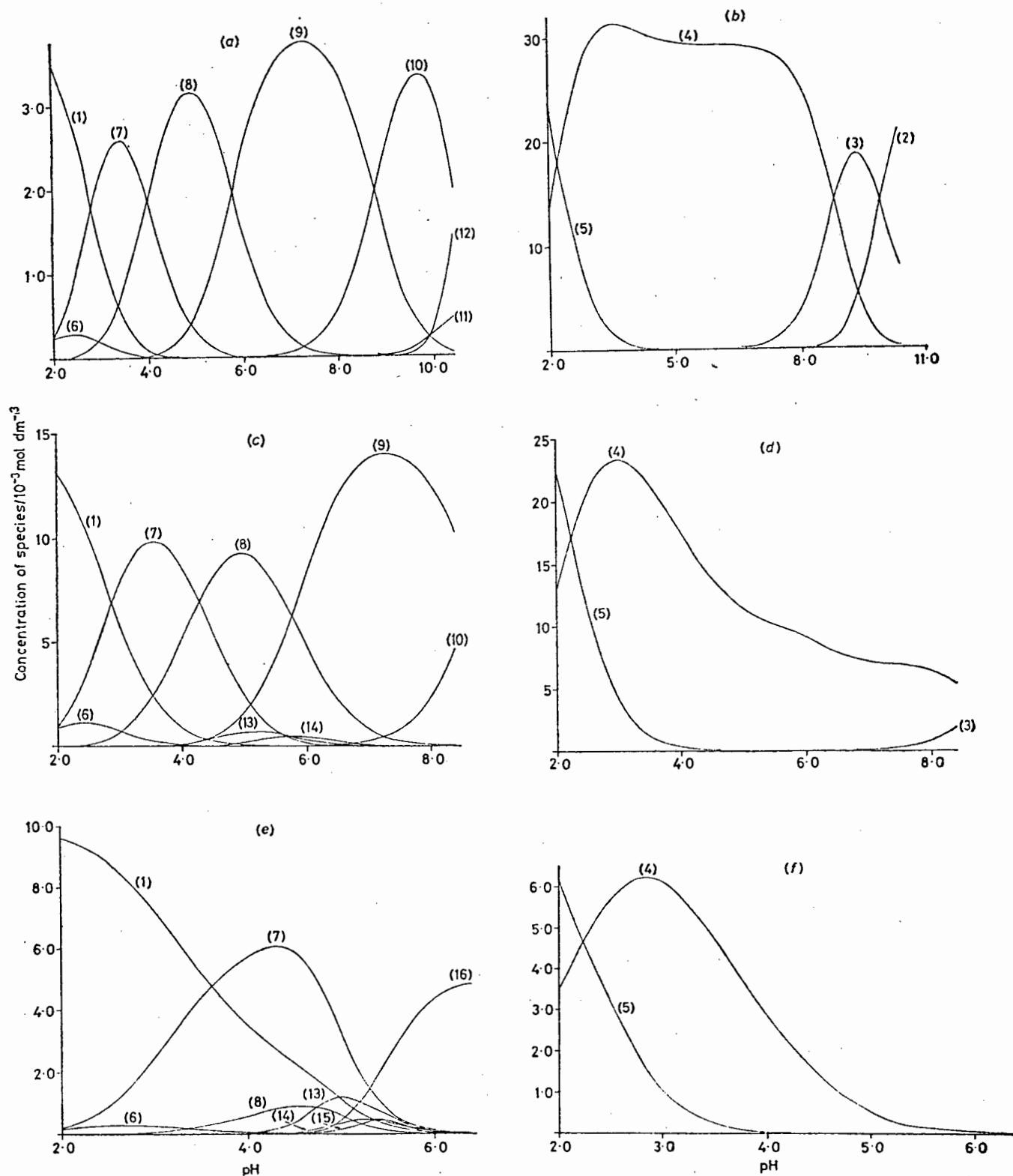
Table 4.5.1

Logarithms of the formation constants for the copper(II) complexes of methyldopa (0,15 M NaClO ₄ , 25°C)			
Complex	$\log \beta_{pqr}$	Std.deviation in $\log \beta_{pqr}$	No.of experimental observations
ABH	17,613	0,005	167
A ₂ BH ₂	34,004	0,031	75
ABH ₂	19,568	0,085	167
A ₂ BH	28,234	0,011	66
A ₂ B	19,487	0,018	105
A ₂ BH ₋₁	8,447	0,264	48
A ₂ BH ₋₂	-1,454	0,094	44
A ₂ B ₂	27,290	0,035	107
A ₃ B ₃ H ₋₁	36,850	0,136	89
A ₄ B ₄ H ₋₂	46,625	0,129	82
A ₂ B ₂ H ₋₂	16,894	0,012	87

R = 0,0030 for the 10:1, 5:1 and 2½:1 ratio solutions and 0,0044 for the 1:1 solutions.

The observation that the formation curve of the 1:1 solution resides in a very different region to that of the other ratios is probably indicative of the higher fraction of oligonuclear species which are apparent in these solutions and is likely due to the relatively large proportion of metal ion present. The concentration of these oligonuclear species is seen to be particularly high between pH 5 and 6, a region in which the ligand and protonated ligand concentrations are extremely small (Fig 4.5.2(f)).

The $2\frac{1}{2}$:1 ratio solutions were found to be of intermediate character between the 10:1 and 5:1 ratios on the one hand and the 1:1 ratio on the other. Thus the species A_2BH and A_2B are important in the 10:1, 5:1 and $2\frac{1}{2}$:1 solutions but not in the 1:1, whilst the species A_2B_2 and $A_3B_3H_{-1}$ are significant only in the $2\frac{1}{2}$:1 and 1:1 solutions but not in the others. In fact, the distributions of complexes in the 1:1 solutions are seen to be distinctly different from those with A:B ratios of 10:1 and 5:1.



HALTAFALL plots of the concentrations of complexes present as a function of pH for the methyl dopa-copper(II) system when the total concentrations ($10^{-3} \text{ mol dm}^{-3}$) of A and B(Cu^{2+}) are: 37.3 and 4.00 [(a), (b)]; 37.2 and 15.0 [(c), (d)]; and 10.0 and 10.0 [(e), (f)]. Species: Cu^{2+} (1), A^{2-} (2), HA^- (3), H_2A (4), H_3A^+ (5), $[\text{Cu}(\text{H}_2\text{A})]^{2+}$ (6), $[\text{Cu}(\text{HA})]^+$ (7), $[\text{Cu}(\text{AH})_2]$ (8), $\text{CuA}(\text{HA})^-$ (9), $[\text{CuA}_2]^{2-}$ (10), $[\text{CuA}(\text{H}_-1\text{A})]^{2-}$ (11), $[\text{Cu}(\text{AH}_-1)_2]^{4-}$ (12), $[\text{Cu}_2\text{A}_2]$ (13), $[\text{Cu}_3\text{A}_3\text{H}_-1]^-$ (14), $[\text{Cu}_4\text{A}_4\text{H}_-2]^{2-}$ (15), $[\text{Cu}_2\text{A}_2\text{H}_-2]^{2-}$ (16)

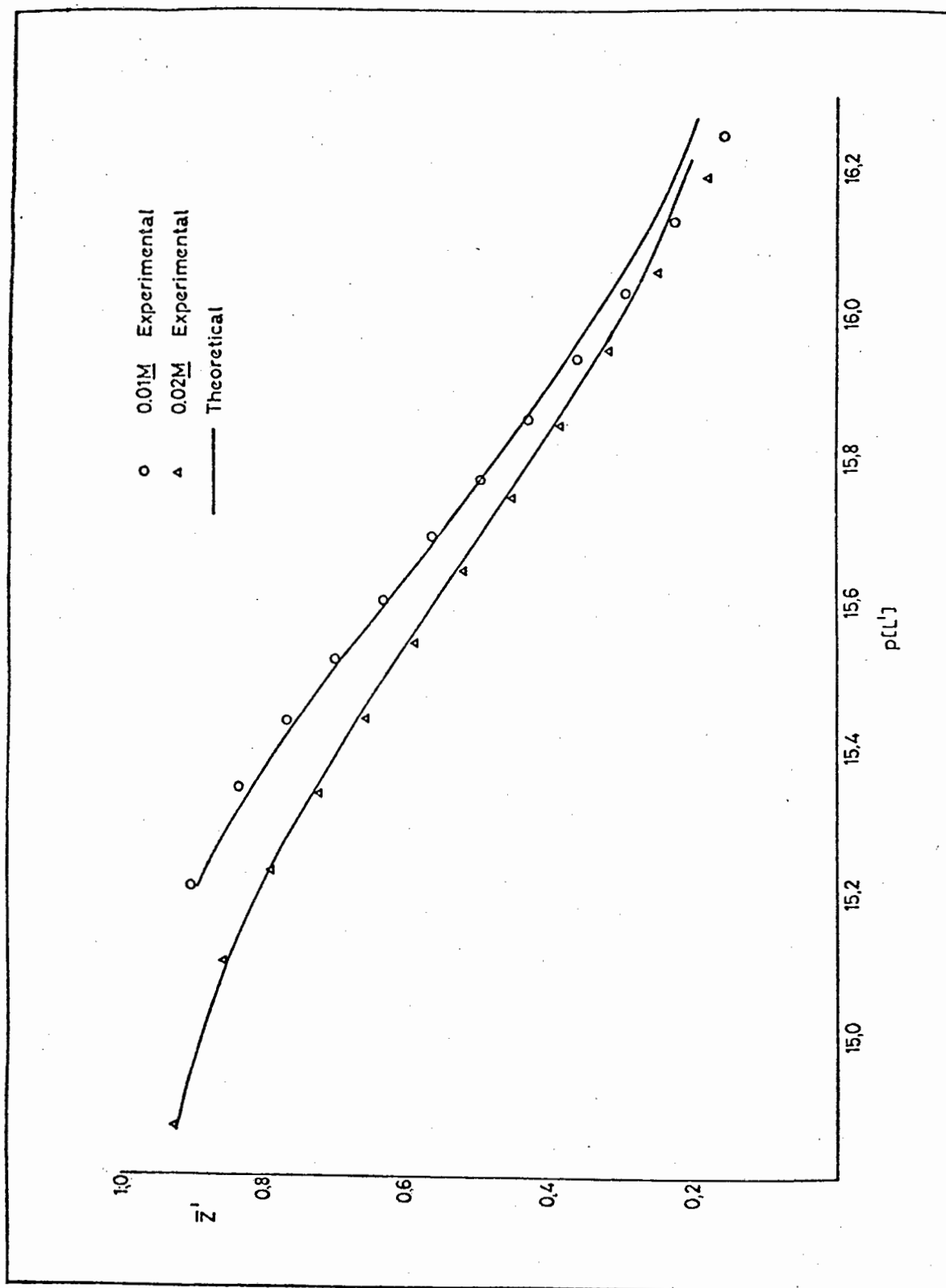
Fig 4.5.2

4.6. Methyldopa-copper(II) formation constants (GEPOLYC).

Application of the program GEPOLYC to the 1:1 ligand: metal ion solutions studied necessitated a knowledge of the protonation constant of the trianion of methyldopa (eqn. 2.1.5.2), a value found too high to determine experimentally in the present study owing to the low background electrolyte used. An attempt was made to find a good estimate of this first protonation constant at 0,15 M ionic strength using the Davies equation,⁷⁹

$$-\log f = AZ^2 \left(\frac{I^{\frac{1}{2}}}{1+I^{\frac{1}{2}}} - CI \right) \quad \dots\dots 4.6.1$$

based on the value for dopa measured at $I = 1,00$ by Gorton and Jameson.⁴⁶ An average value of C was estimated by comparing K'_{102} and K'_{103} for dopa and methyldopa at the two respective ionic strengths, and this was then used to estimate the first protonation constant at 0,15 M ionic strength for methyldopa. K'_{102} and K'_{103} refer to the second and third stepwise protonation constants of the ligand respectively when the fully deprotonated ligand is the trianion. The value for K'_{101} obtained is $10^{13,84}$. The GEPOLYC approach was applied on two 1:1 mixtures of 0,02 M and 0,01 M initial concentration. The best fit was a model which included the series of open chain polymers ($A_n B_n H_{2-n}$) but only the cyclic dimer ($A_2 B_2 H_{-2}$) representing the closed chain species, with values of 13,30 for $\log K$ and 15,55 for $\log K_c$. The formation curves for both concentrations appear in Fig 4.6.1. The computed concentrations of the various species present at selected pH values are shown in Table 4.6.1.



Formation curves for methyl dopa-copper(II) 1:1 solutions (GEFOLYC).

Fig 4.6.1

Table 4.6.1

Distribution of complexes (% of total ligand) in 0,02 M 1:1 solution as a function of pH (GEPOLYC)			
Complex	pH 4,9	pH 5,1	pH 5,5
ABH	51,24	28,53	8,14
A ₂ B ₂	13,63	12,24	5,32
A ₃ B ₃ H ₋₁	3,62	5,20	3,48
A ₄ B ₄ H ₋₂	0,96	2,21	2,27
A ₅ B ₅ H ₋₃	0,26	0,94	1,49
A ₆ B ₆ H ₋₄	0,07	0,40	0,97
A ₂ B ₂ H ₋₂	2,44	6,45	16,01

5. GENERAL DISCUSSION OF RESULTS.

This work has indicated that the complexing characteristics of copper(II) with methyldopa are very similar to those reported for the closely related dopa-copper(II) system.^{40,32,46-52} Below pH 5 predominantly amino acid type chelation exists, changing progressively (to some extent via mixed catechol and amino acid type complexes) to catechol type as the pH is increased. As regards the applicability of this to medicinal chemistry, one can say with some assurance that copper(II) is capable of bonding at both sites at physiological pH, an important structural consideration when the role played by copper(II) in the in vivo metabolism of methyldopa is discussed. The distribution of complexes in a solution containing methyldopa as the only ligand and copper(II) as the only metal ion at concentrations found in the blood of a patient undergoing treatment (typically $0,5$ to $1,9 \times 10^{-5}$ mol dm⁻³ for methyldopa⁸⁰ and 10^{-11} to 10^{-19} mol dm⁻³ for free copper(II) ions⁸¹) would probably closely resemble that in Fig 4.5.2(a). Considering the charged nature of $[\text{CuA}(\text{AH})]^-$ which predominates at the physiological pH of 7.4 in this figure, it is unlikely that this species facilitates the passive transport of methyldopa across biological membranes.

It is realised however that neutral ternary species involving other low molecular weight ligand are likely to be present in human blood plasma and future computations simulating these conditions using the program ECCLES⁸¹ might help to clarify the picture.

Before arriving at the results as presented in section

4.5, an enormous amount of computing was done. This was because it was frequently found that a model that had been successfully refined from titration data grouped in a particular way gave unsuccessful refinement of a solution which was mathematically satisfactory but chemically unsound when applied to the full set of titration data points. That this is a result of unreliability in experimental data is unlikely as reproducibility was fully checked in all cases. The complex nature of the ligand is an important factor as is the wide range of pH and ligand:metal ion ratios over which the titrations were carried out. Clearly for this system, the type of complex species is strongly pH dependent, and even at a given pH the solution composition can be expected to change from one concentration ratio to another. Thus polymer formation seems to be important in the 1:1 ligand:metal ion solutions, but less so in other ratios. As regards refining the complete set together, the observed phenomenon may be explained by proposing that some species important in a minority of concentration ratios tend to be rejected whilst those significant in the majority tend to be retained. Thus the 'weight' of the more predominant species appear to overwhelm the contribution of the less so in the optimisation process. Division and grouping of the titration data according to these considerations hence became necessary. In summary it can be said that the model as reported in section 4,5 is the result of exhaustive efforts at resolving this difficulty, and likely represents the best solution

consistent with the available data.

Tables 5.1 and 5.2 contain protonation constants for dopa and complexation constants for the dopa-copper(II) system respectively as obtained by other workers. Although determined under different conditions, the constants where applicable compare favourably with the values found for the methyl-dopa-copper(II) system during the work presented here.

Important in the evaluation of a model is the assignment of structures to the species refined. The hypothesis that ABH and A₂BH₂ refer to the 1:1 and 2:1 ligand:metal ion complexes respectively, with co-ordination through the aminocarboxylate end of methyl-dopa, can be inferred from the following.

The stepwise stability constant for the formation of ABH can be defined as:

$$\begin{aligned} K_{111} &= \frac{[\text{ABH}]}{[\text{AH}][\text{B}]} \\ &= \frac{[\text{ABH}]}{[\text{A}][\text{B}][\text{H}]} \frac{[\text{A}][\text{H}]}{[\text{AH}]} \end{aligned}$$

$$\text{i.e. } \log K_{111} = \log \beta_{111} - \log \beta_{101}$$

Similarly, K₂₁₂ for the binary complex A₂BH₂ can be defined as:

$$\begin{aligned} K_{212} &= \frac{[\text{A}_2\text{BH}_2]}{[\text{AH}][\text{ABH}]} \\ &= \frac{[\text{A}_2\text{BH}_2]}{[\text{A}]^2[\text{B}][\text{H}]^2} \frac{[\text{AH}][\text{B}]}{[\text{ABH}]} \frac{[\text{A}]^2[\text{H}]^2}{[\text{AH}]^2} \end{aligned}$$

Table 5.1

Logarithms of protonation constants of methyl dopa and dopa

Ligand	Ionic strength/ medium	Temp. (°C)	$\log \beta_{101}$	$\log \beta_{102}$	$\log \beta_{103}$	Ref.
Methyl dopa	0,15 M <u>NaClO₄</u>	25	9,982	18,866	21,104	this work
Lopa	1,0 M <u>KNO</u>	25	9,74	18,45	20,76	46
Dopa	1,0 M <u>KNO</u>	25	9,78	18,58	20,98	32
Dopa	0,37 M <u>NaNO</u>	20	9,87	18,68	20,72	40
Dopa	0(KCl)	25	10,629	19,786	22,004	82
Dopa	0,2 M <u>KCl</u>	25	9,83	18,63	20,85	51

Table 5.2.

Logarithms of formation constants for the copper(II) complexes of methyl-dopa and dopa

Methyl-dopa		Dopa O/°C, I/mol dm ⁻³ , log β_{pqr}	Ref.
Complex	log β_{pqr}		
ABH	17,613	25, 1,0 (KNO ₃), log β_{111} = 16,86, log β_{212} = 32,89, log β_{11-1} = -0,41, log β_{21-2} = -1,86, log β_{22-2} = 14,59	46
A ₂ BH ₂	34,004		
ABH ₂	19,568	25, 1,0 (KNO ₃), log β_{111} = 17,38, log β_{11-1} = 3,08	32
A ₂ BH	28,234	20, 0,37 (NaNO ₃), log β_{111} = 17,47, log β_{212} = 34,25	40
A ₂ B	19,487		
A ₂ BH ₋₁	8,447	25, 0,2 (KCl), log β_{111} = 17,35, log β_{212} = 33,81, log β_{211} = 27,01, log β_{210} = 18,53, log β_{21-1} = 9,03, log β_{21-2} = -1,33, log β_{220} = 26,55, log β_{22-2} = 15,10	51
A ₂ BH ₋₂	-1,454		
A ₂ B ₂	27,290		
A ₃ B ₃ H ₋₁	36,850		
A ₄ B ₄ H ₋₂	46,625		
A ₂ B ₂ H ₋₂	16,894		

i.e. $\log K_{212} = \log \beta_{212} - \log K_{111} - 2 \log \beta_{101}$

According to Sigel,³⁹ discussing mono- and bis-complexes, since more coordination positions are available for bonding of the first ligand to a metal ion than for the second ligand, one would expect $\log K_{212} < \log K_{111}$, and this difference should be between 1 and 2 log units for a bidentate ligand.

Hence, using the refined values from Table 4.5.1,

$$\begin{aligned} \log K_{212} - \log K_{111} &= \log \beta_{212} - 2 \log \beta_{111} \\ &= -1,22 \end{aligned}$$

In addition, potentiometric work carried out concurrently at this laboratory to determine the stability constants of the methyltyrosine-copper(II) system under similar conditions yielded complexes of the type ABH and A₂BH₂ with log β values of comparative magnitude to those obtained in the methyldopa-copper(II) system (18,05 and 35,01 respectively). Methyltyrosine differs from methyldopa in having only one hydroxy group on the phenyl ring, this in the 3' position. Since the only mode of binding likely with methyltyrosine is through its alanine end, complexes of the type ABH and A₂BH₂ obtained in the methyldopa-copper(II) system must represent similar modes of ligand-metal binding.

Based on the conclusion that in bis(glycinato)Cu(II), where the two planar rings are attached to copper(II) in a *cis* arrangement,³⁵ the geometrical structure of A₂BH₂ in the present system is assumed to be of a similar type.

The species A₂BH in the methyldopa-copper(II) system could be regarded as being formed from A₂BH₂ through the deprotonation of a hydroxy group of one of the ligands. This being the case, if it is assumed that the dissociation quotient $[A_2BH][H]/[A_2BH_2]$ can be approximated to $K_{102}^{-1} = [AH][H]/[AH_2]$

(K_{102} is the second stepwise protonation constant for the ligand), then

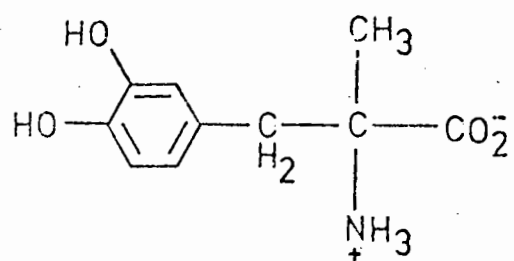
$$\begin{aligned}\beta_{211} &= \frac{[A_2BH]}{[A]^2 [B] [H]} \\ &= \frac{[A_2BH_2]}{[A]^2 [B] [H]^2} \frac{[A_2BH] [H]}{[A_2BH_2]} \\ &\approx \frac{[A_2BH_2]}{[A]^2 [B] [H]^2} \frac{[AH] [H]}{[AH_2]}\end{aligned}$$

$$\text{i.e. } \log \beta_{211} \approx \log \beta_{212} - \log K_{102}$$

The value obtained is 25,12, which is markedly different from the experimental value of 28,234. Thus the structure assigned to A_2BH on this basis is probably incorrect. In accordance with the enhanced stability of A_2BH , the structure is more likely to be a complex involving copper(II) in a mixed mode of bonding, i.e. complexed to the alanine end of one ligand and the catechol end of the other as depicted in Fig 5(3). The implied proposal that neither phenoxide of the methyl dopa ligand is protonated when this moiety is complexed to copper(II) is in agreement with results obtained from potentiometric titrations on the catechol-copper(II) system (section 4.3), where species in which this was not the case were not detectable. Hence it appears that the phenolic proton of the methyl dopa dianion (Fig 5(2)) becomes displaced upon 0,0 coordination to copper(II).

The next in the series of successively deprotonated bis-complexes is A_2B . By similar reasoning as with A_2BH , this complex could be regarded as being derived from A_2BH through neutralisation of one of its uncomplexed catechol protons. In

Fig 5

AH₂

A

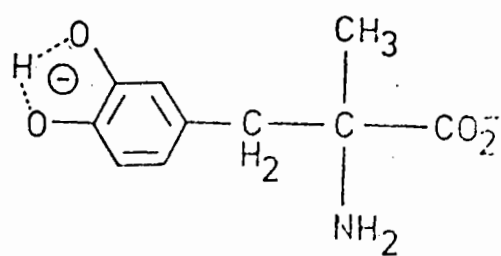
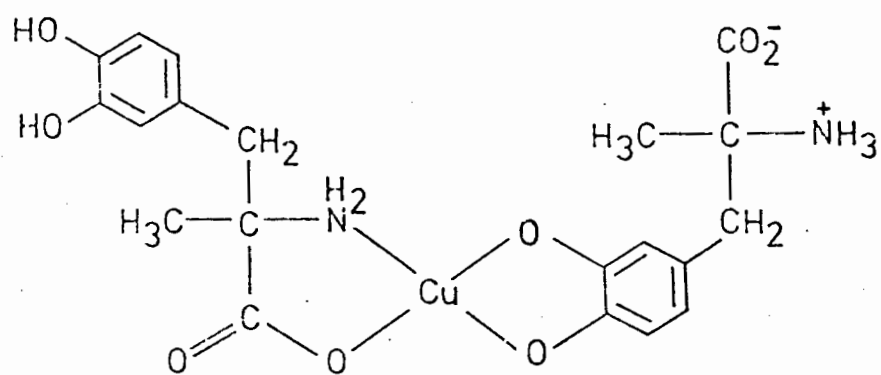
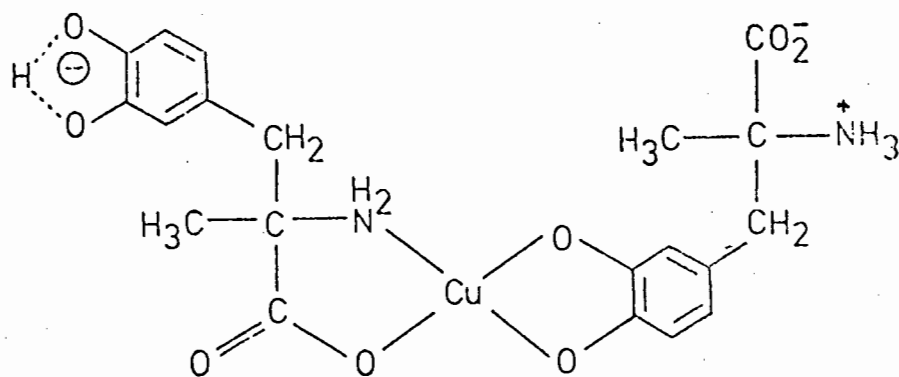
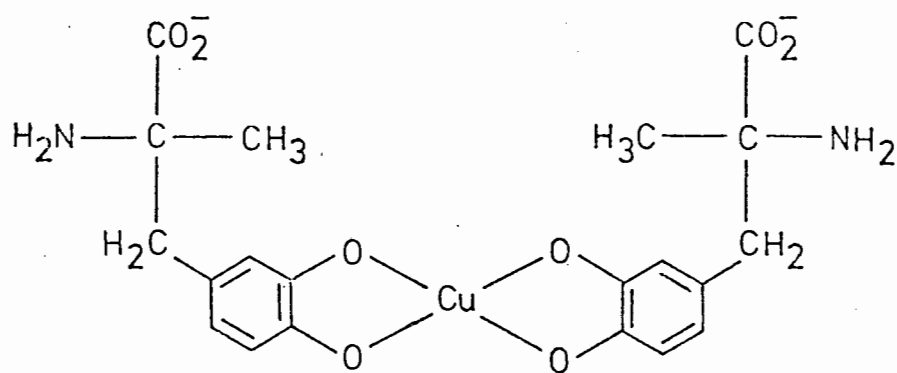
A₂BH

Fig 5 (contd.)

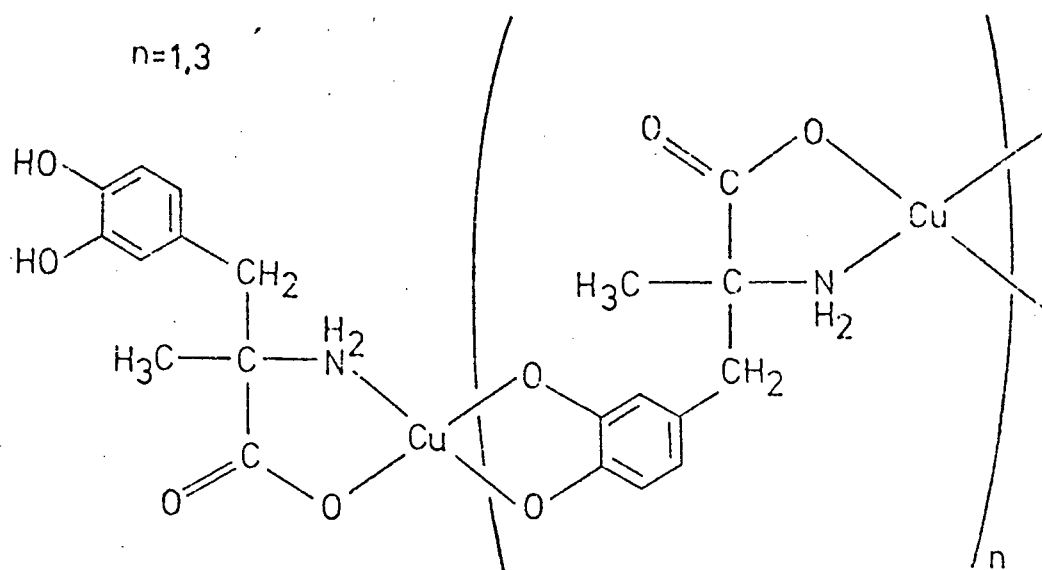
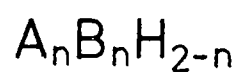
A₂B

(4)

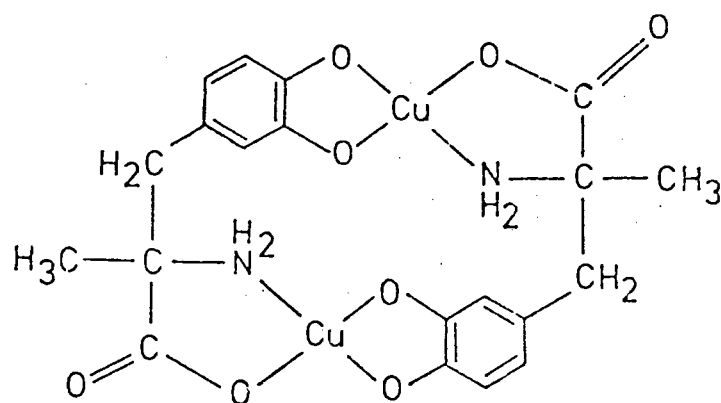
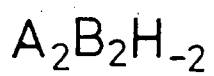
A₂BH₋₂

(5)

Fig 5 (contd.)



(6)



(7)

this case if the quotient $[A_2B][H]/[A_2BH]$ can be approximated to by $[AH][H]/[AH_2]$, then $\log \beta_{210} \approx \log \beta_{211} - \log K_{102}$. The value obtained is 19,35, which is close to the observed value of 19,487. On this basis the structure assigned to the species is as depicted in Fig 5(4). This conclusion is in disagreement with the structure proposed by Gergely and Kiss⁵¹ for the analogous A_2B species detected in the dopa-copper(II) system, in which the bis-complex discussed is assigned a structure in which the metal ion is involved in O,O binding with both ligands.

Considering the high pKa for the phenolic proton on the methyl dopa dianion i.e. 13 to 14, and that A_2BH_{-2} begins to form at about pH 9, the only feasible structure for this complex is that shown in Fig 5(5). A_2BH_{-1} can hence be thought of as being formed either from the deprotonation of the uncomplexed amino group of (4) or the protonation of one of the uncomplexed amino groups of (5). In the former case, the estimated $\log \beta_{21-1}$ value is $\log \beta_{210} - \log \beta_{101} = 9,505$. In the latter $\log \beta_{21-1} \approx \log \beta_{21-2} + \log \beta_{101} = 8,528$. Since the observed value for $\log \beta_{21-1}$ is 8,447, the second implied structure is perhaps the more likely, though not definite.

The structures of the series of open chain polymers $A_n B_n H_{2-n}$ found in MINIQUAD calculations on the 1:1 ligand:metal ion ratio solutions are as shown in Fig 5(6), and involve copper(II) in a 'mixed' mode of bonding as required by the GEPOLYC approach. In support of this assignment was the failure to detect species of analogous stoichiometry in the methyltyrosine-copper(II) and catechol-copper(II) solutions studied, where such polymer formation is structurally impossible.

During model-fitting using MINIQUAD on the 1:1 solutions, it was found that equally successful refinement occurred when the species $A_2B_2H_{-2}$ was substituted for by ABH_{-1} . A complex of stoichiometry ABH_{-1} would by necessity require either the amino group or one of the phenolic protons to be neutralised, or have the copper ion in a hydrolysed form. However, its presence in significant amounts at a pH of 4,5 and the fact that a corresponding species could again not be detected in the methyltyrosine-copper(II) solutions indicated that ABH_{-1} in the methyldopa-copper(II) system was perhaps a fortuitous coincidence in the refinement. The preferred species is hence $A_2B_2H_{-2}$ (Fig. 5(7)), and represents the cyclic dimer suggested by the GEPOLYC calculations. Its formation can be thought to occur via the terminal copper(II) atom in A_2B_2 bending back onto the catechol end of the same molecule with accompanying release of two protons. The feasibility of this species was initially questioned as a Dreiding model showed the copper(II) to aromatic ring distance for the dimer to be only about 3 Å, with the necessary absence of an apical aquo ligand. This difficulty has however been partially resolved in that in the crystal structure of bis-(L-tyrosinato) copper(II),⁸³ a weak interaction of similar magnitude has been observed between the phenolic rings and a copper(II) ion.

A relationship can be derived between the GEPOLYC constants K_c and K and those as refined by MINIQUAD for the 1:1 ligand: metal ion ratio solutions.

From Section 2.1.5

$$\frac{K_c}{K} = \frac{[A_n B_n H_{2-n}]^2}{[A_{n-1} B_{n-1} H_{3-n}] [A_n B_n H_{-n}] [ABH]}$$

Putting $n = 2$ ($K = 0$ for $n \geq 3$ in the final model and undefined for $n = 1$), one obtains

$$\begin{aligned} \frac{K_c}{K} &= \frac{[A_2 B_2]^2}{[ABH]^2 [A_2 B_2 H_{-2}]} \\ &= \frac{[A_2 B_2]^2}{[A]^4 [B]^4} \frac{[A]^2 [B]^2 [H]^2}{[ABH]^2} \frac{[A]^2 [B]^2 [H]^{-2}}{[A_2 B_2 H_{-2}]} \end{aligned}$$

i.e. $\log K_c - \log K = 2 \log \beta_{220} - 2 \log \beta_{111} - \log \beta_{22-2}$.

Substituting for the relevant constants from Sections 4.5 and 4.6 one obtains the difference between the left hand side and right hand side of the above equation to be 0,21 log units, indicating the compatibility of the two approaches.

APPENDIX

A. Calculation of R_{LIM} based on maximum experimental errors incurred in a typical titration.

A calculation of R_{LIM} based on propagation of error theory⁸⁴ using maximum tolerances of apparatus used both in the standardisation of solutions and in the titration itself appears below. The calculations are applied to a hypothetical titration involving the addition of 5 ml of 0,18 M sodium hydroxide to a solution comprising 0,02 M methyldopa, 0,019 M perchloric acid and 0,004 M copper perchlorate. The method will also illustrate the preparation of the solutions. The tolerances of the apparatus used appear in Table A.1.⁶⁵ The error in weighing was assumed to be negligible. In the preparation of some of the solutions, buretted volumes greater than 20 ml were required which necessitated the filling of the burette at least twice. This is accounted for in the calculations.

Table A.1

Nominal capacity of apparatus (ml)				
Tolerance \pm ml	10	20	250	500
Burette (Class A)	0,02	0,04		
Pipette (Class B)	0,04	0,05		
Vol. flask (Class B)			0,2	0,3

After addition of 5 ml of sodium hydroxide, the total hydrogen concentration is:

$$\begin{aligned}
 T_3 &= [\text{Total hydrogen}] \\
 &= \frac{[\text{HClO}_4]_o V_o}{V_o + v} + \frac{2[\text{AH}_2]_o V_o}{V_o + v} - \frac{[\text{NaOH}]_o v}{V_o + v} \\
 &= \frac{(0,019)(20,0)}{25,0} + \frac{(2)(0,02)(20,0)}{25,0} - \frac{(0,18)(5,0)}{25,0} \\
 &= 0,0112 \text{ M}
 \end{aligned}$$

The perchloric acid in the titration vessel is made up of contributions from an acidified copper perchlorate stock solution and from a perchloric acid stock solution.

$$[\text{HClO}_4] = \frac{[\text{HClO}_4]'V'}{250} + \frac{[\text{HClO}_4]''V''}{250}$$

The molarity of the perchloric acid stock solution was determined using borax.

$$\begin{aligned}
 [\text{HClO}_4]'' &= \frac{10^3(\text{mass of borax})/(\text{equiv. wt. of borax})}{(\text{standardisation vol. of perchloric acid})} \\
 &\approx \frac{(10^3)(0,2500)/190,72}{9,0} \approx 0,15 \text{ M}
 \end{aligned}$$

$$\begin{aligned}
 \delta[\text{HClO}_4]'' &= \left[\left(\frac{(10^3)(0,2500)/190,72}{(9,0)^2} \right)^2 (0,02)^2 \right]^{\frac{1}{2}} \\
 &= 3,2 \times 10^{-4}
 \end{aligned}$$

The molarity of the sodium hydroxide titrating solution was determined using potassium hydrogen phthalate.

$$\begin{aligned}
 [\text{NaOH}]_o &= \frac{(10^3)(\text{mass of KHPHTH})/(\text{mol. wt. of KHPHTH})}{(\text{standardisation vol. of sodium hydroxide})} \\
 &\approx \frac{(10^3)(0,2600)/204,22}{7,0} \approx 0,18 \text{ M}
 \end{aligned}$$

$$\begin{aligned}\delta[\text{NaOH}]_o &= \left[\left(-\frac{(10^3)(0,2600)/204,22}{(7,0)^2} \right)^2 (0,02)^2 \right]^{\frac{1}{2}} \\ &= 5,2 \times 10^{-4}\end{aligned}$$

The molarity of the perchloric acid in the copper perchlorate stock solution was determined using the above sodium hydroxide solution.

$$\begin{aligned}[\text{HClO}_4]' &= \frac{[\text{NaOH}]_o (\text{vol. of sodium hydroxide need to neutralise})}{(\text{volume of copper perchlorate stock solution taken})} \\ &\approx \frac{(0,18)(1,0)}{10,0} \approx 0,02 \text{ M}\end{aligned}$$

$$\begin{aligned}\delta[\text{HClO}_4]' &= \left[\left(\frac{1,0}{10,0} \right)^2 (5,2 \times 10^{-4})^2 + \left(\frac{0,18}{10,0} \right)^2 (0,02)^2 + \left(-\frac{(0,18)(1,0)}{(10,0)^2} \right)^2 (0,04)^2 \right]^{\frac{1}{2}} \\ &= 3,7 \times 10^{-4}\end{aligned}$$

$$\text{Let } X = \frac{[\text{HClO}_4]'V'}{250} \approx \frac{(0,02)(42,0)}{250} \approx 0,003 \text{ M}$$

$$\begin{aligned}\delta X &= \left[\left(\frac{0,02}{250} \right)^2 (0,06)^2 + \left(\frac{42,0}{250} \right)^2 (3,7 \times 10^{-4})^2 + \left(-\frac{(0,02)(42,0)}{(250)^2} \right)^2 (0,2)^2 \right]^{\frac{1}{2}} \\ &= 6,23 \times 10^{-5}\end{aligned}$$

$$\text{Let } Y = \frac{[\text{HClO}_4]''V''}{250} \approx \frac{(0,15)(26,0)}{250} \approx 0,016 \text{ M}$$

Note: $[\text{HClO}_4]_o = X + Y = 0,019 \text{ M}$

$$\begin{aligned}\delta Y &= \left[\left(\frac{0,15}{250} \right)^2 (0,06)^2 + \left(\frac{26,0}{250} \right)^2 (3,2 \times 10^{-4})^2 + \left(-\frac{(0,15)(26,0)}{(250)^2} \right)^2 (0,2)^2 \right]^{\frac{1}{2}} \\ &= 5,06 \times 10^{-5}\end{aligned}$$

$$V_o + v = 25,0 \text{ ml}$$

$$\delta(V_o + v) = \{(0,05)^2 + (0,02)^2\}$$

$$= 5,4 \times 10^{-2}$$

$$\begin{aligned}
 [\text{AH}_2]_0 &= \frac{(10^3)(\text{mass of methyl dopa})/(\text{mol. wt. of methyl dopa})}{V_0} \\
 &= \frac{(10^3)(0,1000)/238,0}{20,0} \approx 0,02 \text{ M}
 \end{aligned}$$

$$\begin{aligned}
 \delta[\text{AH}_2]_0 &= \left[\left(-\frac{(10^3)(0,1000)/238,0}{(20,0)^2} \right)^2 (0,05)^2 \right]^{\frac{1}{2}} \\
 &= 5,25 \times 10^{-5}
 \end{aligned}$$

At this stage it is convenient to write T_3 as:

$$T_3 = \frac{V_0 X}{V_0 + v} + \frac{V_0 Y}{V_0 + v} + \frac{2[\text{AH}_2]_0 V_0}{V_0 + v} - \frac{[\text{NaOH}]_0 V_0}{V_0 + v}$$

$$\text{Let } A = \frac{V_0 X}{V_0 + v}$$

$$\begin{aligned}
 \delta A &= \left[\left(\frac{20,0}{25,0} \right)^2 (6,23 \times 10^{-5})^2 + \left(\frac{0,003}{25,0} \right)^2 (0,05)^2 + \left(-\frac{(20,0)(0,003)}{(25,0)^2} \right)^2 (5,4 \times 10^{-2})^2 \right]^{\frac{1}{2}} \\
 &= 5,04 \times 10^{-5}
 \end{aligned}$$

$$\text{Let } B = \frac{V_0 Y}{V_0 + v}$$

$$\begin{aligned}
 \delta B &= \left[\left(\frac{20,0}{25,0} \right)^2 (5,06 \times 10^{-5})^2 + \left(\frac{0,016}{25,0} \right)^2 (0,05)^2 + \left(-\frac{(20,0)(0,016)}{(25,0)^2} \right)^2 (5,4 \times 10^{-2})^2 \right]^{\frac{1}{2}} \\
 &= 5,85 \times 10^{-5}
 \end{aligned}$$

$$\text{Let } C = \frac{2[\text{AH}_2]_0 V_0}{V_0 + v}$$

$$\begin{aligned}
 \delta C &= \left[\left(\frac{(2)(20,0)}{25,0} \right)^2 (5,25 \times 10^{-5})^2 + \left(\frac{(2)(0,02)}{25,0} \right)^2 (0,05)^2 + \left(-\frac{(2)(20,0)(0,02)}{(25,0)^2} \right)^2 (5,4 \times 10^{-2})^2 \right]^{\frac{1}{2}} \\
 &= 1,35 \times 10^{-4}
 \end{aligned}$$

$$\text{Let } D = \frac{[\text{NaOH}]_o v}{V_o + v}$$

$$\begin{aligned} \delta D &= \left[\left(\frac{0,18}{25,0} \right)^2 (0,02)^2 + \left(\frac{5,0}{25,0} \right)^2 (5,2 \times 10^{-4})^2 + \left(- \frac{(0,18)(5,0)}{(25,0)^2} \right)^2 (5,4 \times 10^{-2})^2 \right]^{\frac{1}{2}} \\ &= 1,94 \times 10^{-4} \end{aligned}$$

$$\begin{aligned} \text{Thus } \delta T_3 &= \left((\delta A)^2 + (\delta B)^2 + (\delta C)^2 + (\delta D)^2 \right)^{\frac{1}{2}} \\ &= 2,49 \times 10^{-4} \end{aligned}$$

The error in the total copper molarity of the titration vessel solution, δT_2 , is now calculated.

$$[\text{EDTA}] = \frac{(10^3)(\text{mass of EDTA})/(\text{mol. wt. of EDTA})}{(\text{total vol. of solution})}$$

$$\approx \frac{(10^3)(1,9700)/372,24}{500} \approx 0,01 \text{ M}$$

$$\delta [\text{EDTA}] = \left[\left(- \frac{(10^3)(1,9700)/372,24}{(500)^2} \right)^2 (0,3)^2 \right]^{\frac{1}{2}}$$

$$= 6,35 \times 10^{-6}$$

$$[\text{Cu}(\text{ClO}_4)_2]' = \frac{[\text{EDTA}](\text{titration vol. of EDTA})}{(\text{Vol. of copper perchlorate stock solution taken})}$$

$$\approx \frac{(0,01)(24,0)}{10,0} \approx 0,24 \text{ M}$$

$$\delta [\text{Cu}(\text{ClO}_4)_2]' = \left[\left(\frac{0,01}{10,0} \right)^2 (0,06)^2 + \left(\frac{24,0}{10,0} \right)^2 (6,35 \times 10^{-6})^2 + \left(- \frac{(0,01)(24,0)}{(10,0)^2} \right)^2 (0,04)^2 \right]^{\frac{1}{2}}$$

$$= 1,14 \times 10^{-4}$$

$$[\text{Cu}(\text{ClO}_4)_2]_o = \frac{[\text{Cu}(\text{ClO}_4)_2]' (V')}{250}$$

$$\approx \frac{(0,024)(42,0)}{250} \approx 0,004 \text{ M}$$

$$\delta[\text{Cu}(\text{ClO}_4)_2]_o = \left[\left(\frac{0,024}{250} \right)^2 (0,06)^2 + \left(\frac{42,0}{250} \right)^2 (1,14 \times 10^{-4})^2 + \left(- \frac{(0,024)(42,0)}{(250)^2} \right)^2 (0,2)^2 \right]^{\frac{1}{2}}$$

$$= 2,03 \times 10^{-5}$$

$$T_2 = \frac{[\text{Cu}(\text{ClO}_4)_2]_o (V_o)}{(V_o + v)}$$

$$= \frac{(0,004)(20,0)}{25,0} = 0,0032 \text{ M}$$

$$\delta T_2 = \left[\left(\frac{0,004}{25,0} \right)^2 (0,05)^2 + \left(\frac{20,0}{25,0} \right)^2 (2,03 \times 10^{-5})^2 + \left(- \frac{(0,004)(20)}{(25,0)^2} \right)^2 (5,4 \times 10^{-2})^2 \right]^{\frac{1}{2}}$$

$$= 1,94 \times 10^{-5}$$

The error in the total ligand molarity of the titration vessel solution, δT_1 , is now calculated.

$$T_1 = \frac{[\text{AH}_2]_o (V_o)}{(V_o + v)}$$

$$= \frac{(0,02)(20)}{25} = 0,016 \text{ M}$$

$$\delta T_1 = \left[\left(\frac{0,02}{25,0} \right)^2 (0,05)^2 + \left(\frac{20,0}{25,0} \right)^2 (5,25 \times 10^{-5})^2 + \left(- \frac{(0,02)(20,0)}{(25,0)^2} \right)^2 (5,4 \times 10^{-2})^2 \right]^{\frac{1}{2}}$$

$$= 6,75 \times 10^{-5}$$

$$\text{Hence } R_{\text{LIM}} = \left(\frac{(\delta T_1)^2 + (\delta T_2)^2 + (\delta T_3)^2}{(T_1)^2 + (T_2)^2 + (T_3)^2} \right)^{\frac{1}{2}}$$

$$= \left(\frac{(6,75 \times 10^{-5})^2 + (1,94 \times 10^{-5})^2 + (2,49 \times 10^{-4})^2}{(1,6 \times 10^{-2})^2 + (3,2 \times 10^{-3})^2 + (1,12 \times 10^{-2})^2} \right)^{\frac{1}{2}}$$

$$= 0,0131$$

This is of a somewhat higher order to the R-factor outputted by MINIQVAD in a successful refinement, as might be expected. It must be noted, however, that in this work the preferred criterion in judging a refined model was a comparison of the experimental ZPLOT formation curve with that of the theoretical PSEUDOPLOT. Nevertheless, a correlation in general exists between the value of the R-factor and the degree of overlap between the formation curves.

Symbols used in Appendix A

T_1	=	[Total ligand] in titration vessel after addition of v ml of sodium hydroxide.
T_2	=	[Total metal] in titration vessel after addition of v ml of sodium hydroxide.
T_3	=	[Total hydrogen] in titration vessel after addition of v ml of sodium hydroxide.
V_0	=	initial volume of solution in titration vessel (20 ml).
v	=	volume of sodium hydroxide added (5 ml).
$[AH_2]_0$	=	initial molarity of methyldopa in titration vessel (0,02 <u>M</u>).
$[Cu(ClO_4)_2]_0$	=	initial molarity of copper perchlorate in titration vessel (0,004 <u>M</u>).
$[HClO_4]_0$	=	initial molarity of perchloric acid in titration vessel (0,019 <u>M</u>).
$[NaOH]_0$	=	molarity of sodium hydroxide titrating solution (0,18 <u>M</u>).
$[Cu(ClO_4)_2]'$	=	molarity of copper perchlorate stock solution (0,24 <u>M</u>).
$[HClO_4]'$	=	molarity of perchloric acid in copper perchlorate stock solution (0,02 <u>M</u>).
$[HClO_4]''$	=	molarity of perchloric acid stock solution (0,15 <u>M</u>).
V'	=	volume of copper perchlorate stock solution needed to yield 250 ml of 0,004 <u>M</u> copper perchlorate.

- V'' = volume of perchloric acid stock solution needed to bring the initial perchloric acid molarity of the titration vessel solution to 0,019 M.
- [EDTA] = molarity of ethylenediaminetetra-acetic acid (disodium salt) used in standardisation of copper perchlorate stock solution. (0,01 M).

B. The effect of reactant concentration on formation constants.

For a given background electrolyte concentration, there appears to be a limit to the maximum concentration that participating charged reactants can have, if the stability constants derived therefrom are to be quoted at the electrolyte concentration used. For example, for a background medium of 3,00 M perchlorate ion, the sum of the equivalent concentrations of all positive and negative ions disappearing during a complex forming reaction should not exceed 0,15 M, if the activity coefficients are to be effectively held constant.⁸⁶

Calculations were carried out using data from a 1:1 ligand: metal ion titration. The initial ligand concentration was 0,02 M (initial volume 20 ml) and the sodium hydroxide concentration was 0,142 M. Using the distribution plot of the refined model, the total concentrations of all singly and doubly charged species, anionic and cationic, were computed and two points in the titration, one before any base had been added (i = initial), and the other towards the completion of the titration after 6 ml of base had been added (f = final). Equation B.1.⁵⁵ then yielded the ionic strengths at the two respective points.

$$I = \frac{1}{2} \sum_{i=1}^n C_i Z_i^2 \quad \dots \text{B.1.}$$

The Davies relationship (equation 4.6.1) with $A = 0,509$ and $C = 0,3$ was then used to calculate the activity coefficient of the singly and doubly charged species at the different ionic strengths. For each species, using equation 1.4.1, the ratio $\frac{\beta_{pqr}(i)}{\beta_{pqr}(f)}$ was then calculated (see Table B.1). The deviation of this from unity indicates how the values of the stability constants are affected because of a change in the ionic strength

of the solution due to dilution and shifts in solution composition. These calculations were repeated for a hypothetical titration in which the concentrations of the compounds were doubled, but the background electrolyte remained at 0,15 M. In this it was assumed that the distribution plot remained unchanged. Here the effect on the stability constants of changing the conditions is even more pronounced, as is expected.

Table B.1.

Concentration effect on formation constants during typical 1:1 methyl dopa-copper(II) titrations. (i = initial, f = final).		
Species	$\frac{\beta_{pqr(i)}}{\beta_{pqr(f)}}$ when [A] _T = 0,01 M at start	$\frac{\beta_{pqr(i)}}{\beta_{(pqr)f}}$ when [A] _T = 0,02 M at start
AH	.990	.978
AH ₂	.985	.972
AH ₃	.985	.972
ABH	.981	.956
A ₂ BH ₂	.966	.924
ABH ₂	.985	.972
A ₂ B ₂	.962	.914
A ₃ B ₃ H ₋₁	.948	.883
A ₄ B ₄ H ₋₂	.938	.863
A ₂ B ₂ H ₋₂	.976	.945

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PART II

CRYSTAL AND MOLECULAR STRUCTURES OF SELECTED COMPOUNDS

CHAPTER 1GENERAL EXPERIMENTAL AND COMPUTATIONAL PROCEDURES USED IN
CRYSTALLOGRAPHIC ANALYSIS

1.1 General experimental and computational procedures used in crystallographic analysis.

In the case of all the compounds presented in this work, the procedure followed as regards the preliminary space group determination of the crystals and their data collections was essentially the same, and as outlined below. The stoichiometric composition of each compound was determined by micro-analytical estimates of their carbon, hydrogen and nitrogen content, and their density by flotation in suitable solvents. X-radiation (nickel-filtered $\text{Cu K}\alpha$, $\lambda = 1.5418 \text{ \AA}$) for the photographic work was generated either by the Philips PW 1120 or the PW 1008 models operating at 20 mA and 40 kV, and used in conjunction with Stoe (Heidelberg) Weissenberg and precession goniometers. Preliminary crystallographic information was obtained from single crystal oscillation, Weissenberg and precession photographs taken about the principal axes in order to establish the space group symmetry. The x-ray films used were supplied by Kodak (3M) and the developer and fixer solutions by Medex.

Suitable selected single crystal specimens of each compound were mounted onto glass fibres and sent along with unmounted spares to Dr. G. Gafner at the National Physical Research Laboratory, C.S.I.R. (Pretoria), who very kindly carried out the data collections onto magnetic tapes using a Philips PW 1100 computer controlled four-circle diffractometer. In cases where absorption effects were expected to be especially significant, these were minimised by cutting the crystals into cubes or grinding them into spheres where possible. The x-radiation (graphite monochromated $\text{MoK}\alpha$, $\lambda = 0.7107 \text{ \AA}$) used in the intensity

measurements was supplied by a Philips 1130 3 kW x-ray generator operating at 20 mA and 50 kV. Lattice constants were obtained at room temperature by a least squares fit of the χ , ϕ and 2θ angles of 25 reflections accurately centred on the diffractometer. In all cases, crystal and instrumental stability were monitored by measuring the intensities of three suitable reference reflections approximately every hour. The criterion as to whether a particular reflection was to be considered 'observed' or not in the refinement of a particular structure involved calculation of the standard error $\sigma_{\underline{I}(\text{rel})}$ in the relative integrated intensity $\underline{I}(\text{rel})$ of the reflection, and given by:

$$\sigma_{\underline{I}(\text{rel})} = [(0.02 N_o)^2 + K^2 N_b + \dot{N}_o]^{\frac{1}{2}}$$

N_o is the gross peak count for the reflection, N_b the background count (measured on each side at the peak), and K the ratio of scan to background times. Lorentz-polarization corrections were applied to all reflection data in Pretoria immediately after data collection but absorption effects were not accounted for.

All calculations were carried out on a UNIVAC 1106 computer at the University of Cape Town. The program SHELX^{1.1} was employed for data reduction and in the solution and refinement of the structures. Solutions were effected by Patterson and Fourier techniques, and by use of centrosymmetric direct methods. All structures were refined by full-matrix least-squares, where the function minimized was $w(F_o - F_c)^2$. The functions $\underline{R} = \sum ||F_o| - |F_c|| / \sum |F_o|$ and $\underline{R}_w = \sum w^{\frac{1}{2}} ||F_o| - |F_c|| / \sum w^{\frac{1}{2}} |F_o|$, were used as indicators of the accuracy of each structure determination. The program XANADU^{1.2} supplied bond

distances, angles, torsion angles and least square planes from final atomic coordinates where needed, whilst PLUTO^{1.3} gave the molecular illustration. The subroutine BONDLA, part of the XRAY 72^{1.4} program system, was used to obtain estimated standard deviations in the close contacts reported.

CHAPTER 2

PACKING PATTERNS IN LANTHANIDE-EDTA COMPLEXES: CRYSTAL AND MOLECULAR STRUCTURES OF SODIUM TRIAQUOETHYLENE-DIAMINETETRAACETATODYSPROSIUM(III) PENTAHYDRATE AND CAESIUM DIAQUOETHYLENEDIAMINETETRAACETATOYTTERBIUM(III) TRIHYDRATE

ABSTRACT

$\text{Na}^+ [\text{DyEDTA}(3\text{H}_2\text{O})]^- \cdot 5\text{H}_2\text{O}$ crystallises in the space group Fdd2 with $\underline{a} = 19,34(1)$, $\underline{b} = 35,40(2)$, $\underline{c} = 12,116(6)$ Å, $\underline{z} = 16$. $\text{Cs}^+ [\text{YbEDTA}(2\text{H}_2\text{O})]^- \cdot 3\text{H}_2\text{O}$ crystallises in the space group $\text{P2}_1/\text{c}$ with $\underline{a} = 12,935(6)$, $\underline{b} = 8,997(5)$, $\underline{c} = 17,22(1)$ Å, $\beta = 106,7(2)^\circ$, $\underline{z} = 4$. The structures were determined by Patterson and Fourier methods and refined by full-matrix least-squares to a final R of 0,045 and 0,050 respectively. The decrease in ionic radius from Dy^{3+} to Yb^{3+} results in a change in coordination number of the Ln^{3+} ion from 9 to 8 with concomitant loss of a water molecule. The changed geometry of the lanthanide complex gives rise to a different space group.

2.1 Introduction

During the 1940's, the development of methods for the ion exchange separation of lanthanide fission products instigated intensive research on their complexation properties, and since that time the number of papers concerning this and other aspects of these 4f elements has mushroomed.^{2.1} Having said this, it is nevertheless true to say that the total number of lanthanide species possible is limited when compared to the transition metal ions because of the following factors: the 4f orbitals are significantly hidden beneath the $4d^{10}5s^25p^6$ noble gas atom type of outermost electronic configuration and hence are largely unavailable for ligand formation (the ligand field stabilisation energies are of the order of only 4 kJ mol^{-1}); although the most common oxidation state for the lanthanide ions is 3+, electrostatic attractions are minimised due to the large size of the ion; ligand exchange reactions involving the lanthanide species in solution have been shown to be extremely rapid, limiting the number of isolatable species; there is strong competition by water for coordination sites which precludes coordination by any but the strongest donors and restricts isolatable complexes very largely to those of chelating ligands; and finally the 'hard' nature of the lanthanide ions largely limits the donor atoms to those of similar character, e.g. oxygen and nitrogen.

Although the definite composition of isolated lanthanide complexes in the solid form can be determined, ligand lability precludes any guarantee that these same species will exist in solution. The formation constants of those complexes that have been evaluated in aqueous systems by the standard

techniques have shown that there is a more or less regular increase in stability as one progresses across the series, with the chelates being essentially entropy stabilised.

The reasonably large size of the lanthanide ion (1,02 Å for La^{3+} and 0,85 Å for Lu^{3+}) results in generally high coordination numbers being observed. The spatial positions of the donor atoms with respect to the central atom in a given complex are in turn primarily determined by the coordination number and the nature of the ligand. Hence x-ray studies have established Er^{3+} to be six-coordinated in an octahedral geometry^{2.2} in the complex $[\text{Er}(\text{NCS})_6]^{3-}$, yet nine-coordinated in a tricapped trigonal prismatic geometry in the tris(dimethyl sulphoxide) adduct of erbium nitrate.^{2.3}

Since 1956, when a lanthanum-cyclopentadienyl complex was first synthesised, investigations into the organo-metallic chemistry of the lanthanide elements have become possible. These investigations have provided information on the nature of the metal-carbon bond and the possibility of the participation of f electrons in the metal-ligand bond in these structures. X-ray investigations have shown that the tricyclopentadienyl complexes $\text{M}(\text{C}_5\text{H}_5)_3$, with $\text{M} = \text{Pr}, \text{Pm}, \text{Gd}$ and Tb , crystallise in the same space group, $\underline{\text{Pbcm}}$, and have the same structure as when M is Sm . The arrangement around the central atom is a distorted tetrahedron, with one of the π -bonded cyclopentadienyl rings being effectively shared by two lanthanide ions, the infinite zig-zag $\text{M}(\text{C}_5\text{H}_5)_3$ chains so formed running along the c axis and parallel to (100).^{2.4} Single crystal diffraction studies of $\text{Tm}(\text{C}_5\text{H}_5)_3$ however show that this compound crystallises in the space group $\underline{\text{Pnam}}$ or $\underline{\text{Pna}2_1}$, indicating a change in structure type.^{2.5} It is thought

that this change probably occurs earlier in the lanthanide series, but it is not certain at which element this first becomes apparent.

A similar observation has been reported for the EDTA salts of the lanthanides (EDTA = ethylenediaminetetraacetate). Studies on complexes having the general formula $M\text{LnEDTA}\cdot 8\text{H}_2\text{O}$ show that potassium salts with $\text{Ln} = \text{La}, \text{Nd}$ or Gd , sodium salts with $\text{Ln} = \text{Nd}, \text{Tb}$, or Er , and ammonium salts with $\text{Ln} = \text{Nd}$ or Gd all crystallise in the space group $Fdd2$ with $z = 16$. Consideration of the relative atomic sizes of the central atom has suggested that in the sequence La^{3+} to Lu^{3+} there should be a transition from nine-coordinate $[\text{LnEDTA}(3\text{H}_2\text{O})]^-$ to eight coordinate $[\text{LnEDTA}(2\text{H}_2\text{O})]^-$, with a prediction that Tb would be eight-coordinate.^{2.6} However the structure of $\text{NaTbEDTA}\cdot 8\text{H}_2\text{O}$ has shown Tb to be nine co-ordinate.^{2.7} It was therefore thought interesting to investigate structures of both the Dy and Yb complexes to determine their coordination and packing patterns.

2.2 Crystal preparation and data collection

Crystals of the dysprosium complex $\text{Na}^+[\text{DyEDTA}(3\text{H}_2\text{O})]^- \cdot 5\text{H}_2\text{O}$ were prepared by refluxing 10 mmole of dysprosium oxide and 25 mmole of EDTA in 50 ml water for 15 hours, periodically adding sodium hydroxide pellets until the oxide had dissolved and the pH had reached 5.5. Cooling and evaporation yielded the desired crystals. The ytterbium complex $\text{Cs}^+[\text{YbEDTA}(2\text{H}_2\text{O})]^- \cdot 3\text{H}_2\text{O}$ was prepared in an analogous manner.

Weissenberg photographs of the dysprosium complex (I)

showed the conditions limiting possible reflections to be:

hkl : $h+k, k+l, (l+h) = 2n$
 okl : $k+l = 4n$ ($k, l = 2n$)
 hol : $l+h = 4n$ ($l, h = 2n$)
 hko : ($h, k = 2n$)
 hoo : ($h = 4n$)
 oko : ($k = 4n$)
 ool : ($l = 4n$)

These conditions uniquely determine the space group as $\underline{F}dd2$.^{2.8}

In the case of the ytterbium complex(II), the conditions limiting possible reflections were:

hkl : No conditions
 hol : $l = 2n$
 oko : $k = 2n$

These conditions uniquely determine the space group as $\underline{P}2_1/c$.

Crystal data of the specimens used in the intensity measurements and details of the data collection are listed in Table 2.1.

2.3 Solution and refinement of the structures.

The expected Patterson map for complex (I) with $\underline{z} = 16$ yielded 48 non-origin vector positions each of weight 4. However, because the lattice is face-centred the observed Patterson map can be interpreted by using the 12 non-origin vectors which arise from the general positions of the space group without taking centering into account. The expected positions of such vectors are given in Table 2.2.

The position of the Dy^{3+} atom thus deduced was 0,080; 0,155; 0,000 (the two-fold axis parallel to the Z-axis allows

Table 2.1

Crystal data and experimental and refinement parameters for the crystal structures of the dysprosium-EDTA complex (I) and the ytterbium-EDTA complex (II).		
Crystal data	(I)	(II)
Molecular formula	$\text{Na}^+ [\text{DyEDTA}(\text{3H}_2\text{O})]^- \cdot \text{5H}_2\text{O}$	$\text{Cs}^+ [\text{YbEDTA}(\text{2H}_2\text{O})]^- \cdot \text{3H}_2\text{O}$
M_r	617,8	684,2
Space Group	$Fdd2$	$P2_1/c$
a	19,34(1) Å	12,935(6) Å
b	35,40(2)	8,997(5)
c	12,116(6)	17,22(1)
β		106,7(2) ^o
D_m	1,96 Mg m ⁻³	2,35 Mg m ⁻³
D_c	1,98 for $z = 16$	2,37 for $z = 4$
$\mu(\text{MoK}\alpha)$	3,51 mm ⁻¹	6,47 mm ⁻¹
$F(000)$	4912	1300
<u>Data collection</u>		
Crystal dimensions	0,20x0,30x0,50 mm	0,25x0,20x0,20 mm
Scan mode	$\omega - 2\theta$	$\omega - 2\theta$
Scan width	1,2 ^o θ	1,2 ^o θ
Scan speed	0,03 ^o θ s ⁻¹	0,04 ^o θ s ⁻¹
Range scanned (2θ)	6 - 44 ^o	6 - 50 ^o
Stability of standard reflections	1,7%	0,65%
Number of reflections collected	1423	3614
Number of 'observed' reflections	1192 with $\frac{I}{I_{(rel)}} > 3\sigma I_{(rel)}$	3354 with $\frac{I}{I_{(rel)}} > 2\sigma I_{(rel)}$
<u>Final refinement</u>		
Number of variables	125	120
$R = \sum F_o - F_c / \sum F_o $	0,045	0,050
$R_w = \sum w^{\frac{1}{2}} F_o - F_c / \sum w^{\frac{1}{2}} F_o $	0,043	0,058
Weighting scheme w	$(\sigma^2 F)^{-1}$	$(\sigma^2 F + 2 \times 10^{-3} F^2)^{-1}$
U (methylene H)		0,018 Å ²

	x, y, z	$-x, -y, z$	$\frac{1}{4}-x, \frac{1}{4}+y, \frac{1}{4}+z$	$\frac{1}{4}+x, \frac{1}{4}-y, \frac{1}{4}+z$
x, y, z	$0, 0, 0$	$-2x, -2y, 0$	$\frac{1}{4}-2x, \frac{1}{4}, \frac{1}{4}$	$\frac{1}{4}, \frac{1}{4}-2y, \frac{1}{4}$
$-x, -y, z$	$2x, 2y, 0$	$0, 0, 0$	$\frac{1}{4}, \frac{1}{4}+2y, \frac{1}{4}$	$\frac{1}{4}+2x, \frac{1}{4}, \frac{1}{4}$
$\frac{1}{4}-x, \frac{1}{4}+y, \frac{1}{4}+z$	$-\frac{1}{4}+2x, -\frac{1}{4}, -\frac{1}{4}$	$-\frac{1}{4}, \frac{1}{4}-2y, -\frac{1}{4}$	$0, 0, 0$	$2x, -2y, 0$
$\frac{1}{4}+x, \frac{1}{4}-y, \frac{1}{4}+z$	$-\frac{1}{4}, -\frac{1}{4}+2y, -\frac{1}{4}$	$-\frac{1}{4}-2x, -\frac{1}{4}, -\frac{1}{4}$	$-2x, 2y, 0$	$0, 0, 0$

Table 2.2

the Z-coordinate to be arbitrarily fixed at 0,000). Three cycles of least squares refinement with the dysprosium atom treated as Dy^0 gave an R value of 0,202. Subsequent weighted difference Fourier synthesis yielded the positions of all the remaining non-hydrogen atoms. The final full-matrix least squares refinement was carried out with the dysprosium atom anisotropic and the remaining non-hydrogen atoms isotropic. At no stage of the refinement could hydrogen atoms be located in difference maps and they were therefore omitted from the final model.

A comparison of the expected Patterson map assuming two heavy atoms per asymmetric unit (Table 2.3) with the computed Patterson map yielded possible positions for the Yb^{3+} atom and

x_1' y_1' z_1'	x_1' y_1' z_1'	$-x_1'$ $-y_1'$ $-z_1'$	$-x_1'$ $\frac{1}{2}+y_1'$ $\frac{1}{2}-z_1'$	x_1' $\frac{1}{2}-y_1'$ $\frac{1}{2}+z_1'$	x_2' y_2' z_2'	$-x_2'$ $-y_2'$ $-z_2'$	$-x_2'$ $\frac{1}{2}+y_2'$ $\frac{1}{2}-z_2'$	x_2' $\frac{1}{2}-y_2'$ $\frac{1}{2}+z_2'$
x_1' y_1' z_1'	0 0 0	$-2x_1'$ $\frac{1}{2}$ $\frac{1}{2}-2z_1'$	0 $\frac{1}{2}-2y_1'$ $\frac{1}{2}$	0 $\frac{1}{2}+2y_1'$ $\frac{1}{2}$	$-x_1'+x_2'$ $-y_1'+y_2'$ $-z_1'+z_2'$	$-x_1'-x_2'$ $-y_1'-y_2'$ $-z_1'-z_2'$	$-x_1'-x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}-z_1'+z_2'$	$-x_1'+x_2'$ $\frac{1}{2}-y_1'-y_2'$ $\frac{1}{2}-z_1'+z_2'$
$-x_1'$ $-y_1'$ $-z_1'$	$2x_1'$ $2y_1'$ $2z_1'$	0 0 0	$2x_1'$ $\frac{1}{2}$ $\frac{1}{2}+2z_1'$	$2x_1'$ $-2y_1'$ $2z_1'$	$x_1'+x_2'$ $y_1'+y_2'$ $z_1'+z_2'$	$x_1'-x_2'$ $y_1'-y_2'$ $z_1'-z_2'$	$x_1'+x_2'$ $\frac{1}{2}+y_1'+y_2'$ $\frac{1}{2}+z_1'+z_2'$	$x_1'+x_2'$ $-y_1'-y_2'$ $z_1'+z_2'$
$-x_1'$ $\frac{1}{2}+y_1'$ $\frac{1}{2}-z_1'$	$2x_1'$ $\frac{1}{2}$ $\frac{1}{2}+2z_1'$	0 $\frac{1}{2}-2y_1'$ $\frac{1}{2}$	0 0 0	0 0 0	$x_1'+x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}+z_1'+z_2'$	$x_1'-x_2'$ $\frac{1}{2}-y_1'-y_2'$ $\frac{1}{2}-z_1'-z_2'$	$x_1'+x_2'$ $\frac{1}{2}+y_1'+y_2'$ $\frac{1}{2}-z_1'+z_2'$	$x_1'+x_2'$ $-y_1'-y_2'$ $z_1'+z_2'$
x_1' $\frac{1}{2}-y_1'$ $\frac{1}{2}+z_1'$	0 $\frac{1}{2}+2y_1'$ $\frac{1}{2}$	$-2x_1'$ $2y_1'$ $-2z_1'$	0 $2y_1'$ $-2z_1'$	0 0 0	0 0 0	$-2x_2'$ $-2y_2'$ $-2z_2'$	$-2x_2'$ $\frac{1}{2}$ $\frac{1}{2}-2z_2'$	0 $\frac{1}{2}-2y_2'$ $\frac{1}{2}$
x_2' y_2' z_2'	$x_1'-x_2'$ $y_1'-y_2'$ $z_1'-z_2'$	$-x_1'-x_2'$ $-y_1'-y_2'$ $-z_1'-z_2'$	$-x_1'+x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}-z_1'+z_2'$	$-x_1'+x_2'$ $\frac{1}{2}-y_1'-y_2'$ $\frac{1}{2}-z_1'-z_2'$	0 0 0	$-2x_2'$ $-2y_2'$ $-2z_2'$	$-2x_2'$ $\frac{1}{2}$ $\frac{1}{2}-2z_2'$	0 $\frac{1}{2}-2y_2'$ $\frac{1}{2}$
$-x_2'$ $-y_2'$ $-z_2'$	$x_1'+x_2'$ $y_1'+y_2'$ $z_1'+z_2'$	$-x_1'+x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}-z_1'+z_2'$	$-x_1'+x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}-z_1'+z_2'$	$x_1'+x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}+z_1'+z_2'$	$2x_2'$ $2y_2'$ $2z_2'$	0 0 0	0 $\frac{1}{2}+2y_2'$ $\frac{1}{2}$	$2x_2'$ $\frac{1}{2}$ $\frac{1}{2}+2z_2'$
$-x_2'$ $\frac{1}{2}+y_2'$ $\frac{1}{2}-z_2'$	$x_1'+x_2'$ $\frac{1}{2}-y_1'-y_2'$ $\frac{1}{2}+z_1'+z_2'$	$-x_1'+x_2'$ $y_1'-y_2'$ $-z_1'+z_2'$	$-x_1'+x_2'$ $y_1'-y_2'$ $-z_1'+z_2'$	$x_1'+x_2'$ $-y_1'-y_2'$ $z_1'+z_2'$	$2x_2'$ $\frac{1}{2}$ $\frac{1}{2}+2z_2'$	0 $\frac{1}{2}-2y_2'$ $\frac{1}{2}$	0 0 0	$2x_2'$ $-2y_2'$ $2z_2'$
x_2' $\frac{1}{2}-y_2'$ $\frac{1}{2}+z_2'$	$x_1'-x_2'$ $\frac{1}{2}+y_1'+y_2'$ $\frac{1}{2}-z_1'-z_2'$	$-x_1'-x_2'$ $\frac{1}{2}-y_1'+y_2'$ $\frac{1}{2}-z_1'-z_2'$	$-x_1'-x_2'$ $y_1'+y_2'$ $-z_1'-z_2'$	$x_1'-x_2'$ $-y_1'+y_2'$ $z_1'-z_2'$	0 $\frac{1}{2}+2y_2'$ $\frac{1}{2}$	$-2x_2'$ $2y_2'$ $-2z_2'$	$-2x_2'$ $2y_2'$ $-2z_2'$	0 0 0

Table 2.3

Cs⁺ atom as 0,38; 0,10; 0,16 and 0,60; 0,23; 0,50 respectively. Three cycles of least squares refinement using these values gave an R of 0,186 and a subsequent difference Fourier yielded the positions of all remaining non-

hydrogen atoms except some of the waters of crystallisation. Subsequent refinement was analogous to that of the dysprosium complex (I), except that in the final stages methylenic hydrogen atoms did appear in difference maps. The latter were therefore included in the final model using the technique of constrained refinement ($d(\text{C-H}) = 1,08 \text{ \AA}$) and refining their temperature factors as one single parameter. The water hydrogen atoms could not be located and were omitted. Details of the final refinement parameters are given in Table 2.1. Tables 2.4 and 2.5 show the final atomic coordinates and temperature factors for structures (I) and (II) respectively. Appendices A and B show the respective observed and calculated structure factors.

2.4 Description of the structures and discussion.

The molecular structures with atomic nomenclature of the two compounds appear in Figures 2.1 and 2.2. In complex (I), the structure around the central ion consists of a girdle of four oxygens (one each from the four acetate groups) on either side of which are bonded three water molecules and two nitrogen atoms effectively *trans* to each other. The coordination geometry can best be described by a quasi-mirror passing through Dy³⁺ and the two nitrogen atoms. In this approximation C(1) and C(2) of the puckered ethylenediamine ring are omitted.

In the Yb³⁺ complex, the eight-coordinated square

Table 2.4

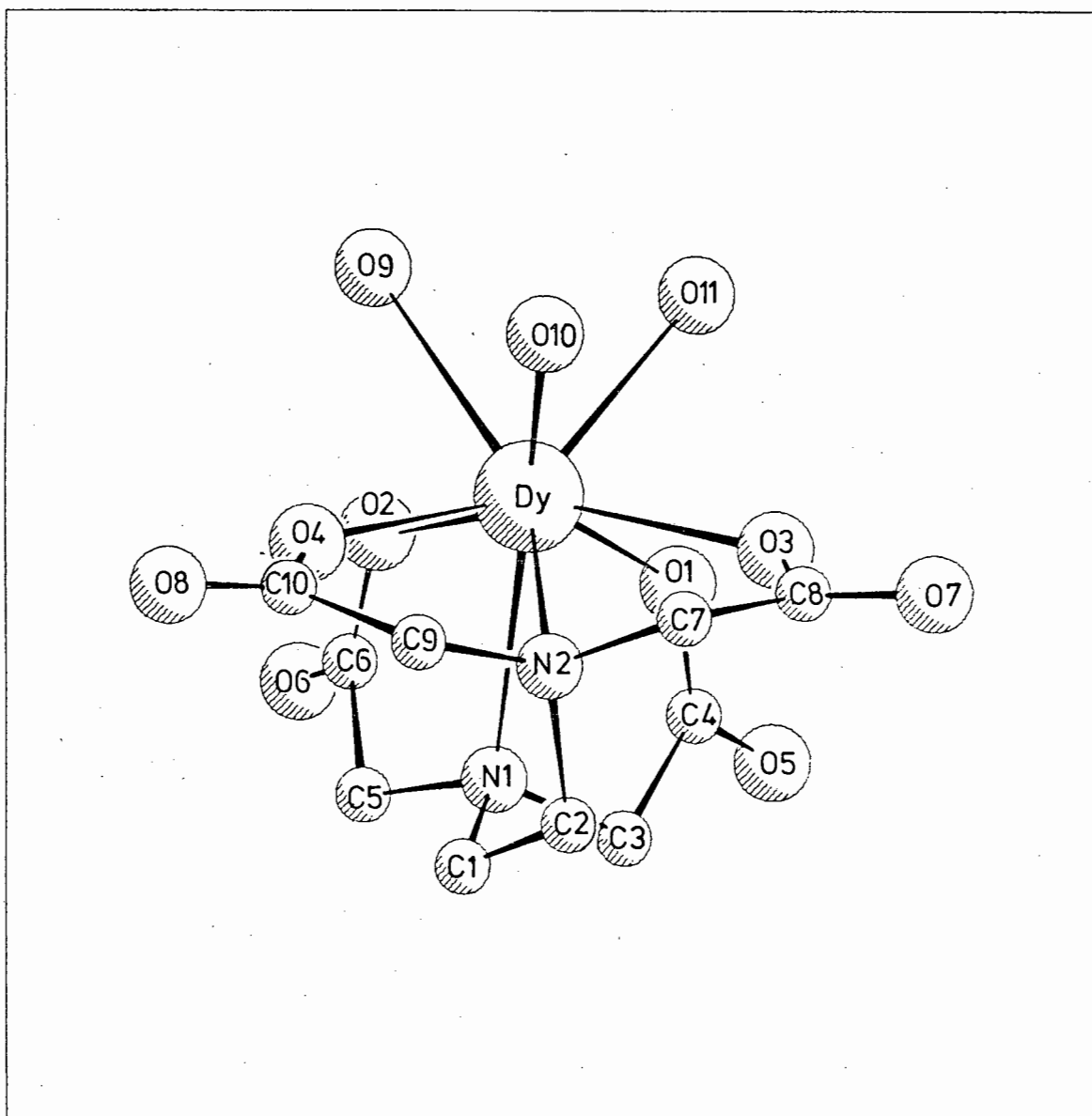
Fractional atomic coordinates ($\times 10^4$) and temperature factors ($\text{\AA}^2 \times 10^3$) with e.s.d.'s for $\text{Na}^+ [\text{DyEDTA}(\text{3H}_2\text{O})]^- \cdot 5\text{H}_2\text{O}$ (I)						
	x	y	z	U		
Dy	823(0)	1553(0)	0	†		
Na	1372(5)	1321(3)	3203(9)	54(3)		
O(1)	963(7)	1822(4)	1862(12)	28(4)		
O(2)	-290(8)	1677(4)	790(13)	41(4)		
O(3)	2029(8)	1682(4)	380(12)	38(4)		
O(4)	173(8)	1640(4)	-1555(13)	33(4)		
O(5)	1054(9)	2307(5)	3001(15)	53(5)		
O(6)	-1008(9)	2089(5)	1575(15)	63(5)		
O(7)	3103(7)	1766(4)	-211(13)	40(4)		
O(8)	68(8)	1703(4)	-3391(13)	39(5)		
O(9)	22(7)	980(3)	-92(13)	33(3)		
O(10)	1341(9)	1104(4)	-1168(13)	36(5)		
O(11)	1301(8)	1065(4)	1260(12)	29(4)		
N(1)	558(8)	2283(4)	150(18)	35(5)		
N(2)	1468(8)	1939(5)	-1523(14)	25(5)		
C(1)	753(12)	2477(8)	-899(22)	36(7)		
C(2)	1459(11)	2352(6)	-1252(19)	34(6)		
C(3)	929(13)	2453(8)	1072(22)	37(8)		
C(4)	974(12)	2163(7)	2042(21)	41(7)		
C(5)	-217(11)	2337(6)	318(18)	36(6)		
C(6)	-501(12)	2016(6)	936(20)	38(6)		
C(7)	2216(12)	1807(6)	-1547(20)	37(6)		
C(8)	2458(11)	1768(6)	-404(17)	27(5)		
C(9)	1151(11)	1854(6)	-2622(22)	36(6)		
C(10)	411(10)	1727(5)	-2515(24)	32(5)		
W(1)	634(8)	390(4)	1089(13)	47(5)		
W(2)	1472(11)	985(6)	5643(19)	85(7)		
W(3)	1212(12)	1783(7)	4603(20)	102(8)		
W(4)	194(14)	1121(8)	3460(24)	132(10)		
W(5)	2594(17)	2154(9)	5720(29)	193(14)		
† Anisotropic parameters in the form $T = \exp[-2\pi^2(u_{11}h^2a^{*2} + u_{22}k^2b^{*2} + u_{33}l^2c^{*2} + 2u_{12}hka^*b^* + 2u_{13}hla^*c^* + 2u_{23}klb^*c^*) \times 10^3]$						
Dy	u_{11} 13(0)	u_{22} 32(1)	u_{33} 23(1)	u_{23} 1(1)	u_{13} -2(1)	u_{12} -1(1)

Table 2.5

Fractional atomic coordinates ($\times 10^4$) and temperature factors ($\text{\AA}^2 \times 10^3$) with e.s.d.'s for $\text{Cs}^+[\text{YbEDTA}(\text{2H}_2\text{O})]^- \cdot 3\text{H}_2\text{O}$ (II)						
	x	y	z	U		
Yb	3758(0)	964(0)	1549(0)	++		
Cs	6039(0)	2308(1)	5008(0)	++		
O(1)	4122(5)	77(7)	2834(4)	26(1)		
O(2)	2651(5)	-873(6)	922(4)	23(1)		
O(3)	3611(5)	3310(7)	1957(4)	23(1)		
O(4)	4007(5)	1225(7)	-299(4)	24(1)		
O(5)	3965(5)	503(8)	4068(4)	33(2)		
O(6)	1133(6)	-2197(8)	693(4)	35(2)		
O(7)	3173(7)	5687(9)	1651(5)	46(2)		
O(8)	3227(5)	698(7)	-1008(4)	30(2)		
O(9)	5051(5)	-906(6)	1577(4)	22(1)		
O(10)	5516(5)	2011(7)	1972(4)	31(2)		
N(1)	2098(6)	860(7)	2041(4)	20(2)		
N(2)	2198(6)	2302(8)	561(4)	20(2)		
C(1)	1308(7)	2003(11)	1651(5)	27(2)		
C(2)	1142(8)	2070(11)	746(6)	29(2)		
C(3)	2458(7)	1116(10)	2935(6)	25(2)		
C(4)	3614(6)	-503(10)	3331(5)	20(2)		
C(5)	1632(8)	-665(11)	1879(6)	29(2)		
C(6)	1798(7)	-1295(10)	1084(6)	24(2)		
C(7)	2516(8)	3892(10)	634(6)	30(2)		
C(8)	3125(7)	4366(10)	1487(5)	23(2)		
C(9)	2118(6)	1726(10)	-262(5)	21(2)		
C(10)	3187(7)	1185(9)	-338(5)	19(2)		
W(1)	-1403(6)	3180(9)	809(5)	44(2)		
W(2)	-76(7)	608(10)	3591(6)	55(2)		
W(3)	8682(8)	822(10)	1954(7)	65(3)		
++ Anisotropic parameters in the form $T = \exp[-2\pi^2(u_{11}h^2a^{*2} + u_{22}k^2b^{*2} + u_{33}l^2c^{*2} + 2u_{12}hka^*b^* + 2u_{13}hla^*c^* + 2u_{23}klb^*c^*) \times 10^3]$						
	u_{11}	u_{22}	u_{33}	u_{23}	u_{13}	u_{12}
Yb	21(0)	12(0)	10(0)	1(0)	2(0)	0
Cs	34(0)	25(0)	31(0)	10(0)	8(0)	2(0)

Table 2.5.1

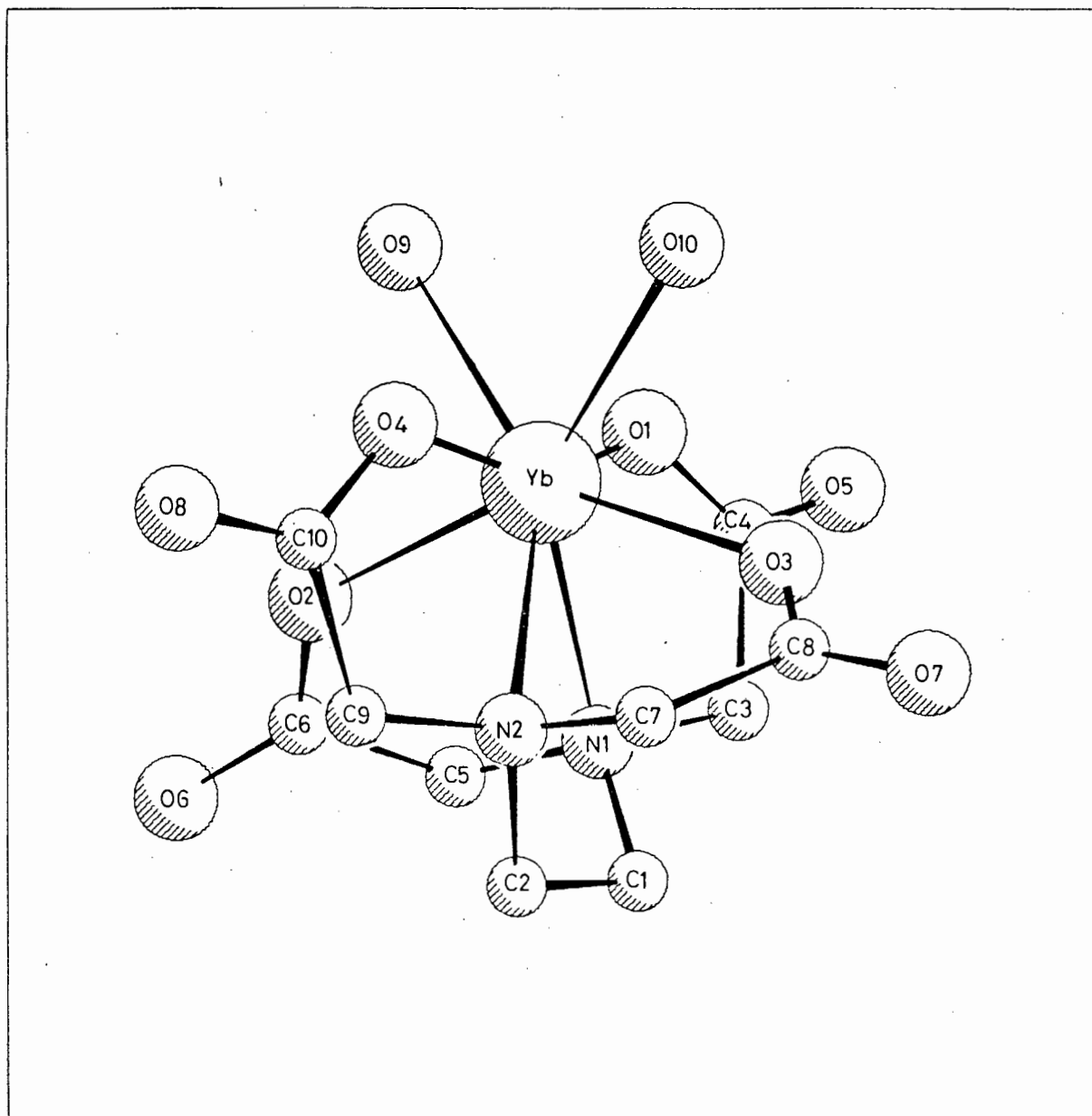
Hydrogen Atoms: Fractional atomic coordinates ($\times 10^3$) for complex (II)			
	x	y	z
H(11)	55	175	176
H(12)	159	307	191
H(21)	61	298	49
H(22)	79	104	47
H(31)	245	230	305
H(32)	190	56	320
H(51)	202	-139	238
H(52)	78	-62	182
H(71)	302	410	24
H(72)	179	456	44
H(91)	155	81	-39
H(92)	183	261	-70



Complex (I)

Fig 2.1

antiprismatic geometry (Figure 2.3) shows effectively two-fold symmetry about an axis bisecting the angle $N(1)-Yb-N(2)$. A certain amount of puckering in the five chelate rings of the EDTA moiety is evident in both compounds, but bond lengths and angles (Table 2.6) measured were comparable to those found in similar metal ion-EDTA crystal studies.^{2.9,2.10} Hence in $Li[FeEDTA(H_2O)]$, the $114,2^\circ$ encountered at a methylene carbon atom and the $115,6^\circ$ at a carboxylate carbon atom represent examples of maximum angular strain in the chelate rings for

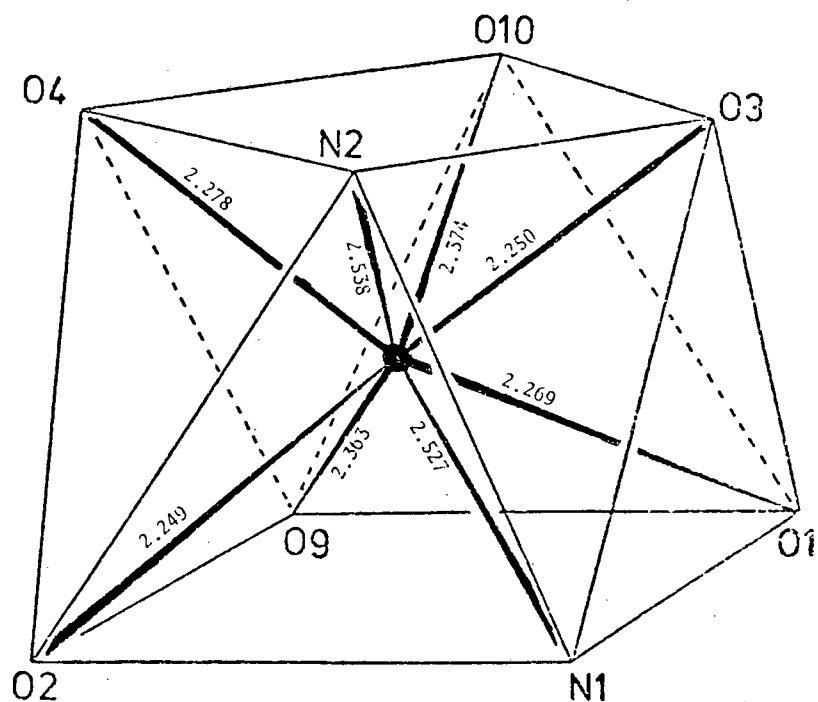
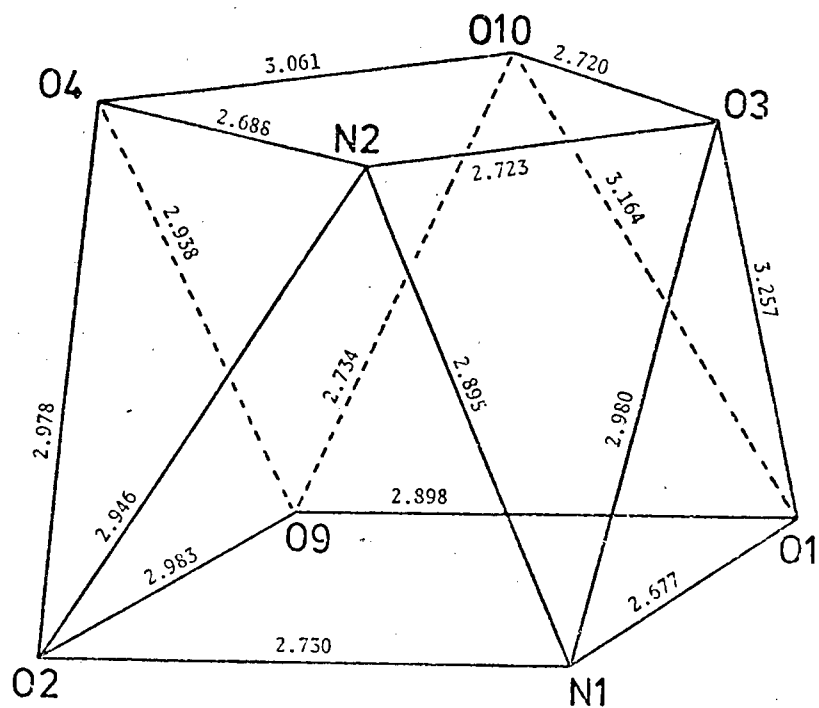


Complex (II)

Fig 2.2

this compound; analogous deviations from ideal tetrahedral and sp^2 geometry in the present study are $111,9^\circ$ and $115,5^\circ$ respectively in complex (I) and $114,1^\circ$ and $115,2^\circ$ respectively in complex (II).

Assuming that the nature of the cation does not radically influence coordination to the central lanthanide ion, the decreasing atomic radius of the latter is manifested in the



Square antiprismatic geometry about the Yb^{3+} ion.

Fig 2.3

Table 2.6

Intramolecular bond lengths (\AA) and selected bond angles ($^\circ$) with e.s.d.'s for the complex anions		
	$[\text{DyEDTA}(\text{3H}_2\text{O})]^-$	$[\text{YbEDTA}(\text{2H}_2\text{O})]^-$
M-O(1)	2,464(14)	2,269(6)
M-O(2)	2,396(15)	2,249(6)
M-O(3)	2,420(14)	2,250(6)
M-O(4)	2,285(15)	2,278(6)
M-O(9)	2,553(12)	2,363(6)
M-O(10)	2,351(16)	2,374(6)
M-O(11)	2,483(14)	
M-N(1)	2,642(16)	2,527(7)
M-N(2)	2,614(17)	2,538(7)
N(1)-C(1)	1,493(31)	1,469(11)
C(1)-C(2)	1,498(30)	1,511(12)
N(2)-C(2)	1,496(25)	1,503(11)
N(1)-C(3)	1,458(31)	1,492(11)
C(3)-C(4)	1,561(35)	1,556(11)
O(5)-C(4)	1,277(25)	1,219(11)
O(1)-C(4)	1,229(25)	1,278(10)
N(1)-C(5)	1,523(25)	1,493(11)
C(5)-C(6)	1,466(29)	1,553(13)
O(6)-C(6)	1,276(26)	1,233(11)
O(2)-C(6)	1,280(24)	1,271(11)
N(2)-C(7)	1,520(26)	1,484(11)
C(7)-C(8)	1,469(30)	1,513(13)
O(7)-C(8)	1,269(23)	1,220(11)
O(3)-C(8)	1,298(22)	1,289(11)
N(2)-C(9)	1,496(30)	1,484(10)
C(9)-C(10)	1,505(27)	1,507(11)
O(8)-C(10)	1,254(28)	1,248(11)
O(4)-C(10)	1,289(28)	1,289(10)
O(1)-M-O(4)	139,7(5)	155,6(2)
O(2)-M-O(3)	139,0(5)	137,7(2)
O(1)-M-O(11)	70,4(5)	
O(2)-M-O(9)	67,6(5)	80,6(2)

continued...../

Table 2.6 continued/

O(4)-M-O(10)	80,2(5)	82,3(2)
O(9)-M-N(1)	131,5(4)	129,2(2)
O(10)-M-N(2)	74,1(5)	121,6(2)
O(9)-M-O(10)	72,3(5)	70,5(2)
O(9)-M-O(11)	72,6(5)	
O(10)-M-O(11)	75,0(4)	
N(1)-M-N(2)	68,2(5)	69,7(2)
M-N(1)-C(1)	110,0(14)	111,5(5)
N(1)-C(1)-C(2)	109,8(19)	111,9(7)
N(2)-C(2)-C(1)	111,1(19)	110,8(7)
M-N(2)-C(2)	110,5(13)	112,8(5)
M-N(1)-C(3)	111,2(14)	107,3(5)
N(1)-C(3)-C(4)	109,5(20)	111,9(7)
O(5)-C(4)-C(3)	115,5(21)	118,6(7)
O(1)-C(4)-C(3)	120,7(22)	115,3(7)
M-O(1)-C(4)	123,0(15)	122,9(5)
M-N(1)-C(5)	108,8(11)	107,9(5)
N(1)-C(5)-C(6)	109,9(18)	110,4(7)
O(6)-C(6)-C(5)	116,2(21)	118,0(8)
O(2)-C(6)-C(5)	122,3(21)	115,7(8)
M-O(2)-C(6)	120,9(14)	125,9(6)
M-N(2)-C(7)	107,8(12)	104,7(5)
N(2)-C(7)-C(8)	108,3(18)	114,1(8)
O(7)-C(8)-C(7)	119,2(19)	118,5(8)
O(3)-C(8)-C(7)	120,5(18)	115,2(8)
M-O(3)-C(8)	121,4(12)	124,4(5)
M-N(2)-C(9)	109,1(11)	107,4(5)
N(2)-C(9)-C(10)	111,9(21)	112,4(7)
O(8)-C(10)-C(9)	116,8(24)	118,5(7)
O(4)-C(10)-C(9)	119,2(23)	117,9(7)
M-O(4)-C(10)	125,5(13)	119,7(5)

expulsion of a water molecule from the inner coordination sphere on changing from Dy^{3+} to Yb^{3+} . Hoard *et al*^{2.6} report that for $[\text{LaEDTA}(\text{3H}_2\text{O})]^-$, the displacement of La^{3+} from the mean plane of the four complexed oxygen atoms of EDTA is 0,60 Å on the side away from the nitrogen atoms. In complex (I), this distance is 0,48 Å, a decrease of 0,12 Å, and illustrates the enhanced ability of EDTA to envelope more effectively the smaller cation. In complex (II) this trend is continued to such an extent as to cause the rejection of one water molecule with consequent change in coordination number.

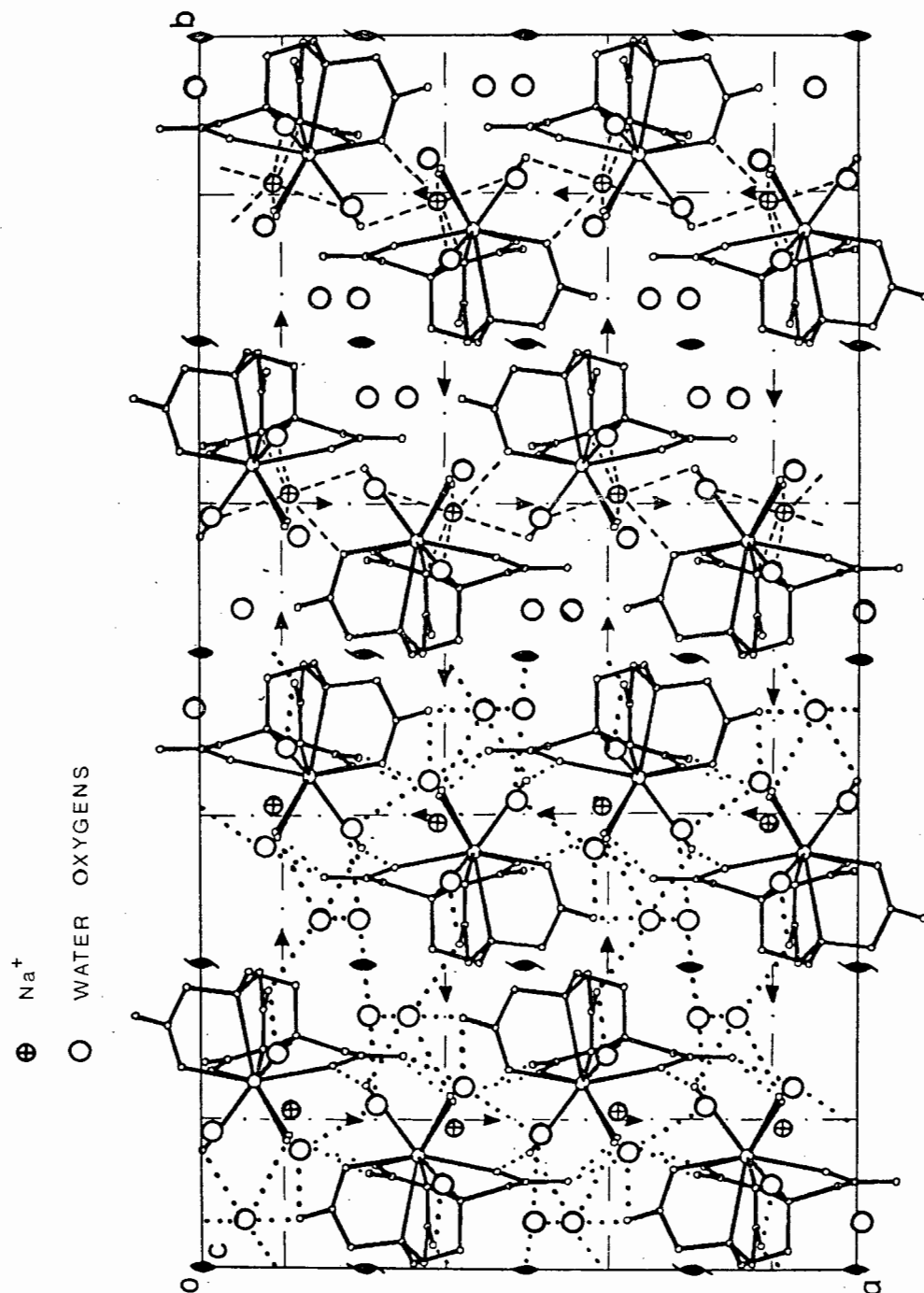
Bell^{2.11} discusses the largely electrostatic interaction between lanthanide ion and ligand which causes increasing stability as the size of Ln^{3+} decreases. Thus,^{2.6} the average of the four Ln^{3+} - O bond lengths is 2,507 Å in $[\text{LaEDTA}(\text{3H}_2\text{O})]^-$, 2,319 Å in complex (I) and 2,262 Å in complex (II). The average Ln^{3+} - N and Ln^{3+} - OH_2 bond lengths in the three structures show similar shortening, having average values of 2,755 Å, 2,628 Å and 2,533 Å respectively for the former type of bond, and 2,580 Å, 2,462 Å and 2,369 Å for the latter. (Standard deviations are 0,005 - 0,006 Å for individual Ln - O, Ln - N and Ln - OH_2 bond lengths in $[\text{LaEDTA}(\text{3H}_2\text{O})]^-$, 0,012 - 0,017 Å for the same bonds in complex (I) and 0,006 - 0,007 Å for these bonds in complex (II)).

2.5 Packing patterns and close contacts.

The Dy^{3+} complex (I) crystallises in the space group Fdd2 . O...O close contacts in the range 2,47(5) Å to 2,98(2) Å (Table 2.7) are shown by dotted lines in the left

Table 2.7

Close intramolecular and intermolecular approaches (Å) with e.s.d.'s for complex (I)			
Symmetry code: (i) $\frac{1}{2} + x, \quad y, \quad \frac{1}{2} + z$			
(ii) $\frac{1}{4} + x, \quad \frac{1}{4} - y, \quad \frac{1}{4} + z$			
(iii) $\frac{1}{4} + x, \quad \frac{1}{4} - y, \quad -\frac{3}{4} + z$			
(iv) $\frac{1}{4} - x, \quad \frac{1}{4} + y, \quad \frac{1}{4} + z$			
(v) $\frac{1}{2} - x, \quad \frac{1}{2} - y, \quad z$			
O(11)....O(1)	2,85(2)	W(5)....O(6 ⁱ)	2,90(4)
O(9)O(2)	2,76(2)	W(2)....O(6 ⁱⁱ)	2,78(3)
O(11)....O(3)	2,81(2)	O(7)....O(2 ⁱⁱⁱ)	2,76(3)
O(10)....O(4)	2,99(2)	O(7)....O(10 ⁱⁱ)	2,69(2)
O(10)....O(9)	2,90(2)	O(3)....O(9 ⁱⁱ)	2,70(2)
O(9)O(11)	2,98(2)	O(11)....O(4 ⁱⁱ)	2,78(2)
O(10)....O(11)	2,95(2)	W(3)O(5)	2,70(3)
O(9)O(4)	2,95(2)	O(10)....O(8 ⁱⁱ)	2,63(2)
		W(1)O(9)	2,79(2)
		W(1)O(11)	2,72(2)
		O(5)W(1 ^{iv})	2,69(2)
		W(2)W(4 ⁱⁱ)	2,77(3)
		W(5)W(4 ⁱⁱ)	2,77(4)
		W(5)W(1 ⁱⁱ)	2,79(4)
		W(5)W(5 ^v)	2,47(5)



(001) projection of Dy³⁺ structure.

Fig. 2.4

half of the [001] projection shown in Figure 2.4 and represent probable H-bonding. A stronger indication of formal H-bonds was not possible due to the uncertainty of the hydrogen positions on any of the water molecules. As regards $\text{Na}^+\dots\text{O}$ close contacts, the environment of the Na^+ atom can best be described in terms of its position at the centre of a highly distorted octahedron, depicted by broken lines in the right half of Figure 2.4. The $\text{Na}^+\dots\text{O}$ distance varies from 2,40 Å to 2,54 Å and agrees favourably with similar results obtained by Fallon and Gatehouse,^{2.12} who measured distorted octahedral $\text{Na}^+\dots\text{O}$ distances varying between 2,35 Å and 2,48 Å in sodium acetate trihydrate, and Cameron, Mannan and Rahman,^{2.13} where the $\text{Na}^+\dots\text{O}$ distance ranged between 2,35(1) Å and 2,56(1) Å in sodium μ -triethylenetetraaminehexaacetatodi-(oxo-vanadium(IV)) decahydrate.

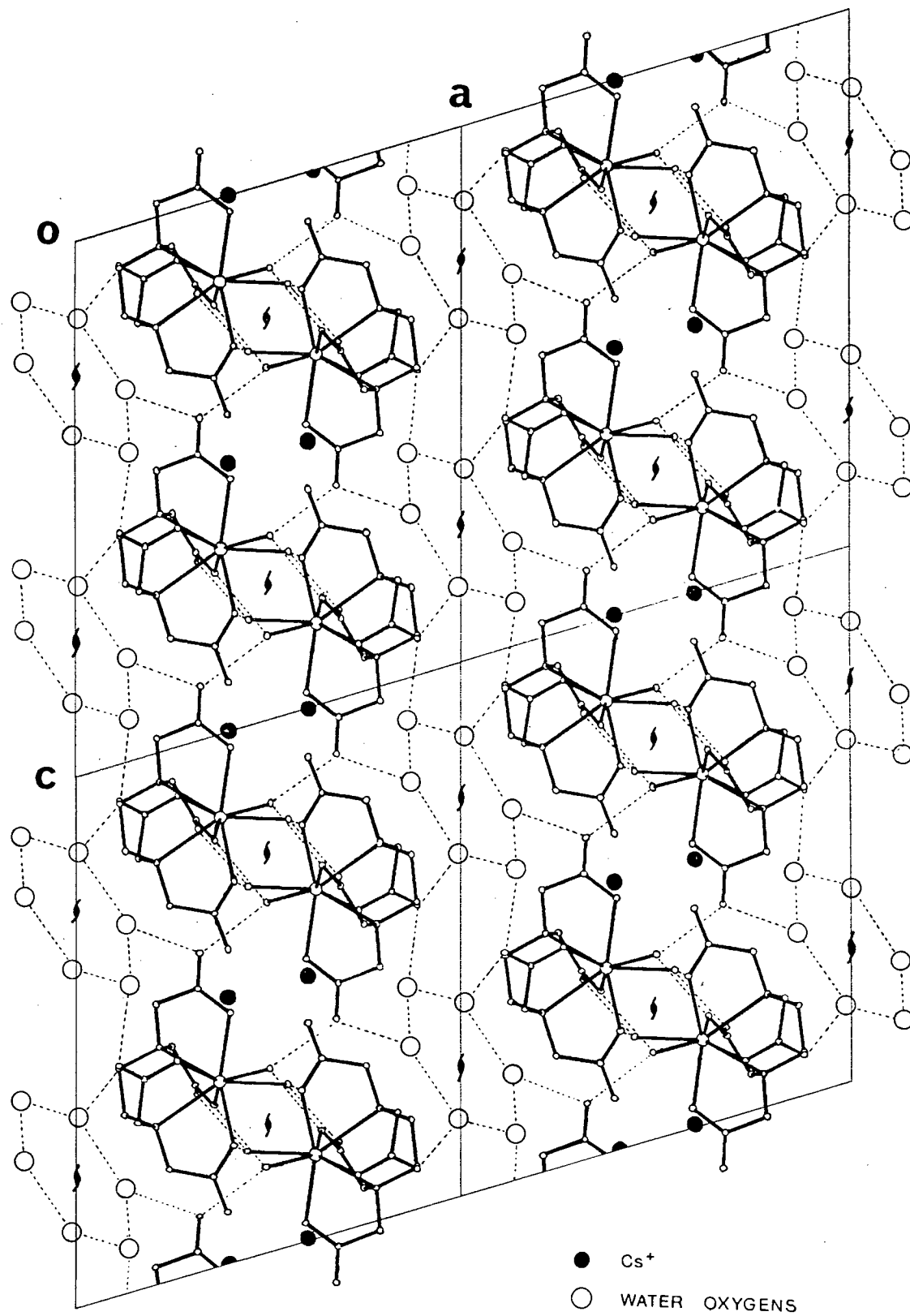
The packing of the Yb^{3+} complex (II) is shown by the four unit cells as a [010] projection in Figure 2.5. The channels of water molecules in the bc-plane are extensively H-bonded (Table 2.8), the $\text{O}\dots\text{O}$ close contacts being in the range 2,69(1) Å to 3,16(1) Å. Gupta and Mahata^{2.14} found that in caesium hydrogen acetylenedicarboxylate monohydrate, the Cs^+ atom was nine-fold coordinated to surrounding oxygen atoms, the $\text{Cs}^+\dots\text{O}$ distance varying from 3,31(3) Å to 3,40(2) Å. In the present study the Cs^+ atom is irregularly surrounded by six O atoms, the close contacts having values less than 3,40 Å.

In the Yb^{3+} complex (II) the change in the crystal packing from $\underline{\text{Fdd2}}$ to $\underline{\text{P2}}_1/\underline{\text{c}}$ is probably influenced by at least three factors, which are not necessarily independent:

- (i) the loss of a water molecule concomitant with the decreasing size of the central ion as the coordi-

Table 2.8

Close intramolecular and intermolecular approaches (Å) with e.s.d.'s for complex (II)			
Symmetry code:			
	(i)	$1 - x, \frac{1}{2} + y, \frac{1}{2} - z$	
	(ii)	$1 - x, -y, -z$	
	(iii)	$-x, -y, -z$	
	(iv)	$-x, -\frac{1}{2} + y, \frac{1}{2} - z$	
	(v)	$1 + x, y, z$	
0(9)0(1)	2,90(1)	0(10)....0(1 ⁱ)	2,80(1)
0(10)....0(1)	3,16(1)	0(3)0(9 ⁱ)	2,71(1)
0(9)0(2)	2,98(1)	W(1)....0(6 ⁱⁱⁱ)	2,85(1)
0(10)....0(3)	2,72(1)	0(6)....W(2 ^{iv})	2,87(1)
0(9)0(4)	2,94(1)	0(7)....0(10 ⁱ)	2,76(1)
0(10)....0(4)	3,06(1)	0(9)....0(8 ⁱⁱ)	2,69(1)
0(10)....0(9)	2,73(1)	W(3)0(8 ⁱⁱ)	2,88(1)
		W(3)W(1 ^v)	2,88(1)
		W(2)W(1 ^{iv})	2,89(1)
		W(3)W(2 ^v)	2,82(1)



(010) projection of Yb³⁺ structure.

Fig 2.5

nation number of the Ln^{3+} changes from 9 to 8 results in a different geometry of the ytterbium complex anion;

- (ii) the different numbers of waters of crystallisation;
- (iii) the large size of the Cs^+ cation.

It is recognised that a substitution of the much smaller Na^+ or K^+ ion for Cs^+ in the ytterbium complex might help in resolving the relative importance of the proposed factors governing the crystal packing pattern for this compound as mentioned above.

CHAPTER 3

THE CRYSTAL STRUCTURES OF A PENTA-COORDINATE PHOSPHORUS COMPOUND: 3,4,8,9-DIBENZ-5,7-DIMETHYL-1-PHENYL-2,6,10,11-TETRAOXATRICYCLO[5,3,1,0^{1,5}] PHOSPHA-UNDECANE AND ITS HYDROLYSIS PRODUCT: PHENYL-1-HYDROXY-1-(2'-HYDROXYPHENYL)-ETHYLIDYNYL PHOSPHINIC ACID

ABSTRACT

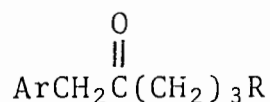
Reaction of PhPCl_2 with $o\text{-HOC}_6\text{H}_4\text{COCH}_3$ gives an unusual tricyclic tetraoxaphospha-undecane. Crystals of this compound are monoclinic, $P2_1/c$, with $a = 13,874(7)$, $b = 7,924(4)$, $c = 16,572(8)$ Å, $\beta = 94,5(2)^\circ$, $z = 4$. The geometry around the phosphorus atom is distorted trigonal bipyramidal, with the two five-membered and one six-membered rings all in apical-equatorial positions. It is postulated that permutational isomerisation in solution is unlikely for this compound due in part to its molecular rigidity.

Crystals of the disubstituted phosphinic acid are monoclinic, $P2_1/n$, with $a = 13,886(7)$, $b = 7,378(4)$, $c = 15,280(8)$ Å, $\beta = 116,9(2)^\circ$, $z = 4$. The phosphorus atom has the expected tetrahedral environment. $O\dots O$ close contacts less than $2,76(1)$ Å indicate hydrogen bonding which extends throughout the lattice.

2.1 Introduction

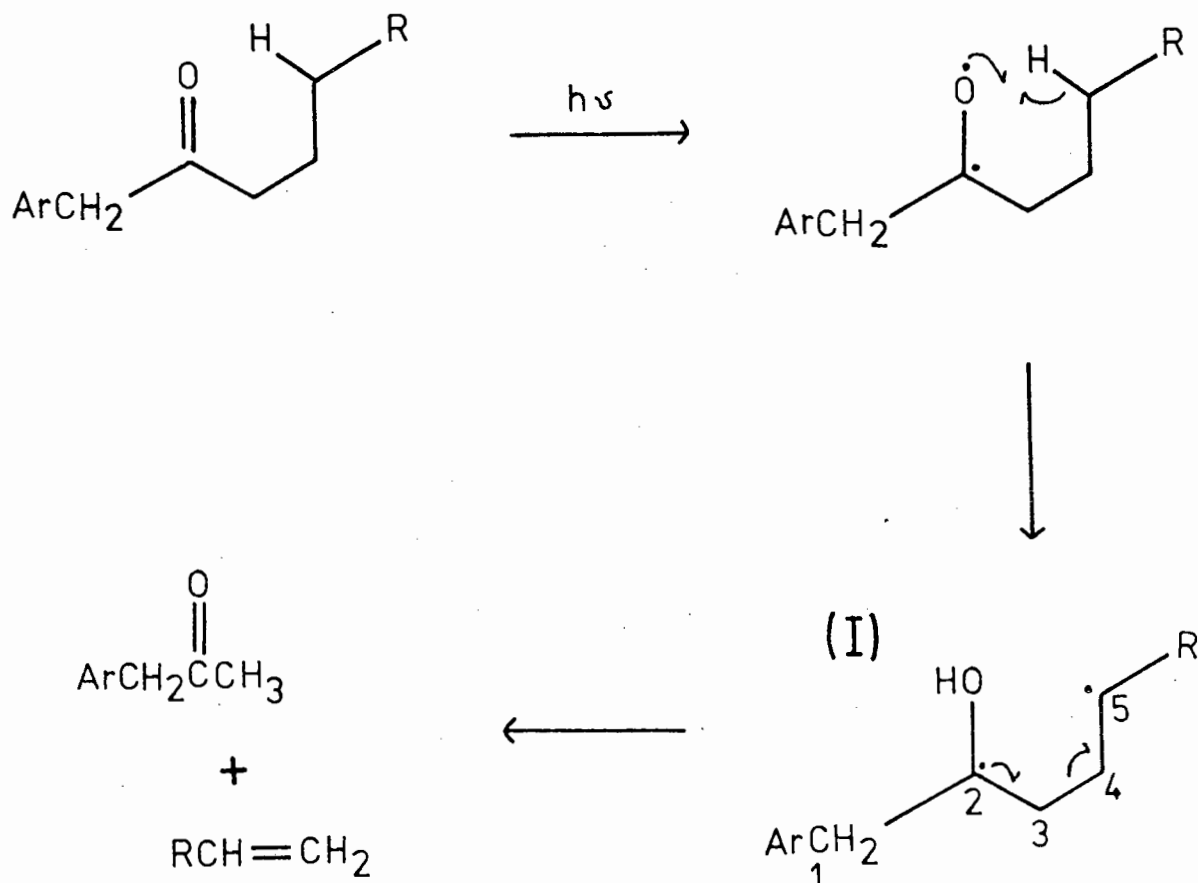
The importance of photochemical reactions in synthetic organic chemistry is well established. An understanding of the molecular orbital excitations involved has proved to be one of the major challenges in this field. Inherent in any mechanistic study of this type of reaction is the need for techniques such as labelling experiments, stereochemistry, kinetics and the trapping of intermediates which besides accounting for the observed phenomena can make useful predictions about how the reaction will respond to changes in substrate or conditions. The famous Diels-Alder cycloaddition process has been characterised and understood on this basis.

Work in our laboratories has recently been directed towards studies relating to compounds such as



(Ar = aryl and R = H or alkyl or aryl), where under light of appropriate wavelength, an $n \rightarrow \pi^*$ excitation of the carbonyl group usually leads to chemical degradation via a postulated diradical intermediate (I, Scheme 3.1). This is generally known as a Norrish Type II process,^{3.1, 3.2} and requires a hydrogen atom on the carbon atom γ to the carbonyl group.^{3.3}

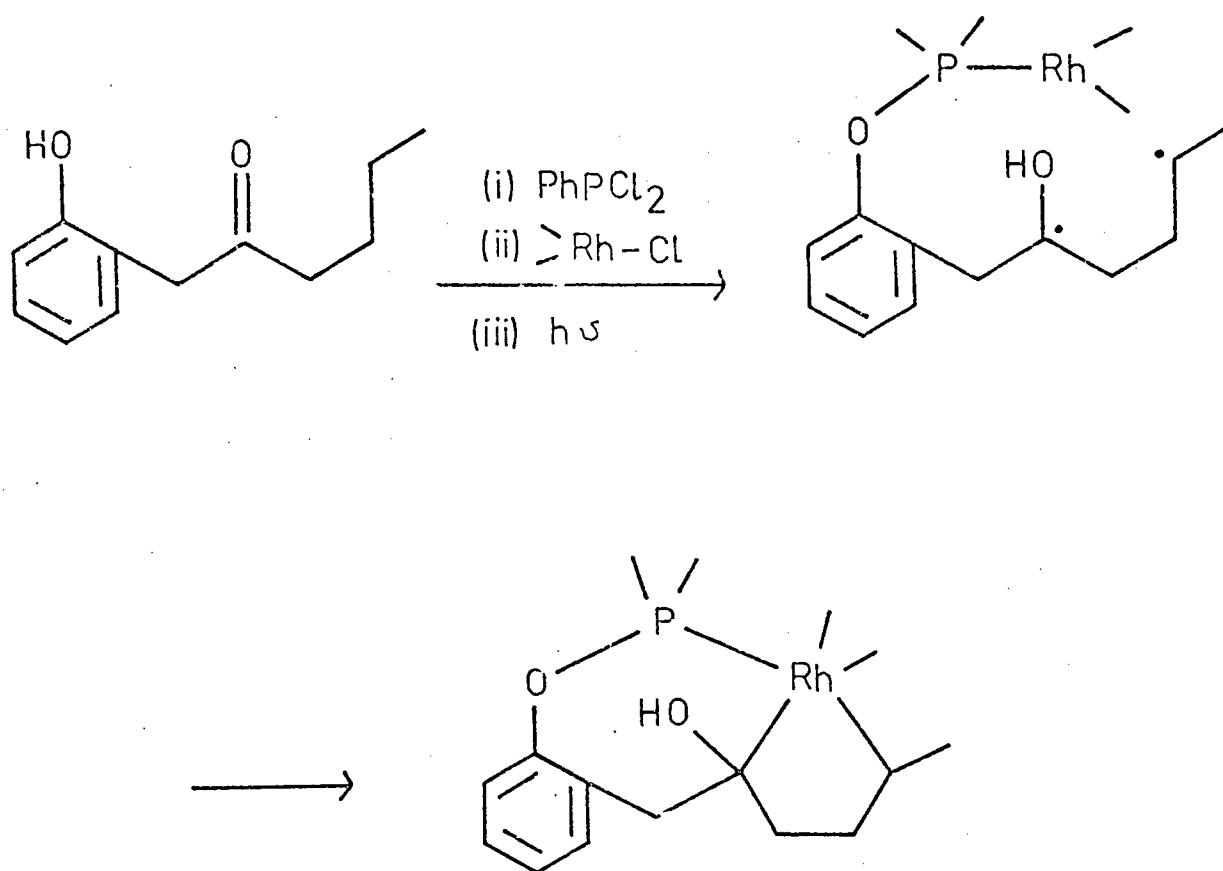
In addition to the two products shown below, ring closure of the biradical intermediate (I) can occur to yield the cyclobutanol as a minor product. Now Rh(I) and Fe(0) have the property of being able to insert into carbon-carbon bonds and have a capacity to undergo oxidative addition.^{3.4} If insertion



Scheme 3.I

of one of these metal atoms between C(2) and C(5) of the excited intermediate (I) could be induced and the product isolated, the existence of such a biradical would be substantiated. However, since the biradical will be short-lived and the concentrations used in photochemistry are of a very low order, the probability of any interaction occurring between the biradical and the free metal derivative would be extremely small. A possible solution would be prior coordination of the metal derivative to the organic species before irradiation, facilitating the proposed trapping of the biradical during

photolysis by the metal atom because of its close proximity. In order to minimise the interaction between the keto group and the metal atom, and to place the metal atom stereochemically in a more favourable position for bond formation to occur, a phosphorus atom could form a bridge between the two moieties (Scheme 3.2).



Scheme 3.2

This atom, being a 'soft' ligand, might be expected to form a strong metal-ligand bond and thus a stable complex with a 'soft' metal such as Rh(I) .^{3.5}

In preliminary investigations along these lines, reaction of PhPCl_2 with two equivalents of $o\text{-HOC}_6\text{H}_4\text{COCH}_3$ in triethylamine gave a crystalline product of stoichiometry $\text{PhP}(\text{OC}_6\text{H}_4\text{COCH}_3)_2$. The absence of $\text{C}=\text{O}$ stretching peaks in the infrared spectrum of this compound and the presence of two methyl resonances in its NMR spectrum with one split into a doublet ($J = 20 \text{ Hz}$) indicated that it was not the expected product. It was thus decided to solve the structure by x-ray analysis. The characterisation of the product as an unusual tricyclic tetra-oxaphospha-undecane (I) is herein described.

An attempt to recrystallise (I) using 'wet' acetone i.e. not completely dry, led to the formation of a new substance. Subsequent x-ray analysis on this compound, also described herein, confirms that (I) undergoes hydrolysis in forming the di-substituted phosphinic acid (II).

3.2 Crystal preparation and data collection.

12,62 g (0,1 mole) $o\text{-HOC}_6\text{H}_4\text{COCH}_3$ in 70 ml benzene was added dropwise at 0°C to a solution of 10,12 g (0,1 mole) NEt_3 and 8,95 g (0,05 mole) PhPCl_2 in 170 ml benzene. The reaction, carried out under N_2 , was stirred at 0°C for a further 3 hours and left overnight. After filtration to remove the amine hydrochloride, the filtrate was concentrated under reduced pressure. The crystalline product (I) was purified by recrystallisation from hot benzene. δ 1,65 (3H, d, J 20 Hz, 5 Me), 2,0 (3H, s, 7-Me), 6,7-7,7 (11H, m, ArH) and 8,0-8,4 (2H, m, ArH). (II) resulted from an attempted recrystallisation of (I) using acetone which had not been rigorously dried.

Oscillation and Weissenberg photographs taken of (I) and

(II) gave the crystallising space groups as $P2_1/c$ for the former and $P2_1/n$ for the latter.^{2.8} Crystal data and experimental details of the data collections are listed in Table 3.1.

3.3 Solution and refinement of the structures.

The structure of the tetraoxaphospha-undecane (I) was solved by the automatic centrosymmetric direct methods routine of the SHELX program system,^{1.1} in which an E map yielded the positions of all the heavy atoms. Some of the hydrogen atoms were located in subsequent Fourier maps. The final refinement was carried out with the heavy atoms treated anisotropically, the methyl H atoms being refined as rigid groups and the remaining H atoms being constrained at 1,08 Å from their respective C atoms, their positions dictated by the geometry of the molecule. The isotropic temperature factors of the H atoms were treated as two single parameters. Details of the final refinements are reported in Table 3.1.

The disubstituted phosphinic acid (II) was solved in an analogous manner, although hydroxyl hydrogen atoms could not be located in difference Fourier maps and were therefore omitted from the final model.

The positional and thermal parameters of the non-hydrogen atoms are presented in Table 3.2 for compound (I) and Table 3.3 for compound (II). The hydrogen atom positions are listed in Tables 3.2.1 and 3.3.1 respectively. Observed and calculated structure factors appear in Appendices C and D.

Table 3.1

Crystal data and experimental and refinement parameters for the crystal structures of the phospho-undecane (I) and its hydrolysis product (II).		
Crystal data	(I)	(II)
Molecular formula	C ₂₂ H ₁₉ O ₄ P	C ₁₄ H ₁₅ O ₄ P
<u>Mr</u>	378,4	278,2
Space group	P2 ₁ /c	P2 ₁ /n
<u>a</u>	13,874(7)Å	13,886(7)Å
<u>b</u>	7,924(4)	7,378(4)
<u>c</u>	16,572(8)	15,280(8)
β	94,5(2) ^o	116,9(2) ^o
<u>D_m</u>	1,36 Mg M ⁻³	1,20 Mg m ⁻³
<u>D_c</u>	1,38 for z=4	1,22 for z=4
μ (MoK α)	0,133 mm ⁻¹	0,159 mm ⁻¹
<u>F(000)</u>	792	584
<u>Data Collection</u>		
Crystal dimensions	0,38x0,50x0,88 mm	0,14x0,16x0,20mm
Scan mode	ω -2 θ	ω -2 θ
Scan width	1,0 ^o θ	1,2 ^o θ
Scan speed	0,03 ^o θ s ⁻¹	0,04 ^o θ s ⁻¹
Range scanned (2 θ)	6-50 ^o	6-44 ^o
Stability of standard reflections	3,4%	1,4%
Number of reflections collected	3257	1880
Number of 'observed' reflections	2463 with $\underline{I}(\text{rel}) > 2\sigma\underline{I}(\text{rel})$	1140 with $\underline{I}(\text{rel}) > 2\sigma\underline{I}(\text{rel})$
<u>Final refinement</u>		
Number of variables	252	177
$R = \sum F_o - F_c / \sum F_o $	0,049	0,058
$R_w = \sum w^{\frac{1}{2}} F_o - F_c / \sum w^{\frac{1}{2}} F_o $	0,065	0,050
Weighting scheme w	$(\sigma^2 F + 1,34 \times 10^{-2} F^2)^{-1}$	$(\sigma^2 F)^{-1}$
U (aromatic H)	0,067 Å ²	0,085 Å ²
U (methyl H)	0,050	0,076

Table 3.2

Non hydrogen atoms. Fractional atomic coordination ($\times 10^4$) and anisotropic temperature factors ($\text{\AA} \times 10^3$) with e.s.d.'s for compound (I).

Thermal parameters are of the form: $T = \exp[-2\pi^2(U_{11}h^2a^{*2} + U_{11}k^2b^{*2} + U_{33}l^2c^{*2} + 2U_{12}hka^*b^* + 2U_{13}hla^*c^* + 2U_{23}klb^*c^*) \times 10^3]$

	x	y	z	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
P	2463(1)	828(1)	1680(0)	29(1)	23(1)	34(1)	1(0)	1(0)	3(0)
O(2)	1440(1)	273(3)	1059(1)	31(1)	32(1)	44(1)	-4(1)	-4(1)	6(1)
O(6)	3764(1)	-1529(2)	1913(1)	29(1)	20(1)	59(1)	-1(1)	-3(1)	-2(1)
O(10)	2933(1)	1575(3)	885(1)	37(1)	32(1)	37(1)	4(1)	2(1)	6(1)
O(11)	3457(1)	1190(2)	2318(1)	32(1)	22(1)	33(1)	-1(1)	2(1)	3(1)
C(3)	1562(2)	-1291(4)	717(2)	31(2)	28(2)	38(2)	-6(1)	3(1)	-1(1)
C(4)	2284(2)	-2279(4)	1098(2)	29(2)	22(2)	39(2)	1(1)	5(1)	-5(1)
C(5)	2737(2)	-1436(4)	1844(2)	31(2)	22(2)	41(2)	5(1)	0(1)	0(1)
C(7)	4205(2)	122(4)	2112(2)	30(2)	20(1)	43(2)	-2(1)	0(1)	-2(1)
C(8)	4588(2)	864(3)	1355(2)	31(2)	22(1)	42(2)	-8(1)	4(1)	0(1)
C(9)	3929(2)	1676(3)	811(2)	35(2)	23(2)	35(2)	-4(1)	5(1)	-1(1)
C(12)	2322(2)	-2104(4)	2612(2)	50(2)	37(2)	38(2)	10(1)	1(1)	-9(1)
C(13)	4939(2)	-96(4)	2830(2)	36(2)	31(2)	51(2)	6(1)	-11(1)	-1(1)
C(1A)	1756(2)	2254(4)	2245(2)	34(2)	27(2)	40(2)	-2(1)	6(1)	-2(1)
C(2A)	1918(3)	2388(5)	3085(2)	60(2)	50(2)	41(2)	-2(2)	5(2)	17(2)
C(3A)	1360(3)	3458(6)	3511(2)	74(3)	65(3)	46(2)	-8(2)	14(2)	19(2)
C(4A)	652(3)	4429(5)	3111(2)	54(2)	48(2)	64(2)	-15(2)	16(2)	10(2)
C(5A)	497(2)	4328(4)	2274(2)	41(2)	37(2)	68(2)	-5(2)	1(2)	8(2)
C(6A)	1031(2)	3229(4)	1843(2)	31(2)	29(2)	58(2)	-1(1)	1(1)	6(1)
C(4B)	2465(2)	-3879(4)	818(2)	43(2)	28(2)	42(2)	-1(1)	2(1)	1(1)
C(5B)	1905(3)	-4476(5)	142(2)	53(2)	38(2)	52(2)	-12(2)	-3(2)	3(2)
C(6B)	1175(3)	-3477(5)	-228(2)	63(2)	45(2)	40(2)	-13(2)	-10(2)	2(2)
C(7B)	998(2)	-1877(5)	54(2)	46(2)	49(2)	42(2)	-3(2)	-9(1)	7(2)
C(8C)	5562(2)	895(4)	1210(2)	36(2)	35(2)	52(2)	-9(1)	8(1)	-2(1)
C(9C)	5865(3)	1738(5)	541(2)	44(2)	56(2)	57(2)	-14(2)	20(2)	-9(2)
C(10C)	5191(3)	2577(5)	17(2)	63(3)	49(2)	43(2)	-6(2)	19(2)	-14(2)
C(11C)	4222(3)	2545(4)	157(2)	52(2)	39(2)	38(2)	-2(1)	9(2)	-2(2)

Table 3.2.1

H atoms. Fractional atomic coordinates ($\times 10^3$) for compound (I)			
	x	y	z
H(121)	155	-192	258
H(122)	265	-145	314
H(123)	248	-344	266
H(131)	550	-97	270
H(132)	458	-54	335
H(133)	525	113	296
H(2A)	248	165	340
H(3A)	148	354	416
H(4A)	21	526	345
H(5A)	-5	511	196
H(6A)	89	314	119
H(4B)	303	-465	111
H(5B)	204	-572	-9
H(6B)	74	-396	-75
H(7B)	43	-110	-24
H(8C)	609	27	162
H(9C)	662	174	42
H(10C)	543	325	-50
H(11C)	370	320	-25

Table 3.3

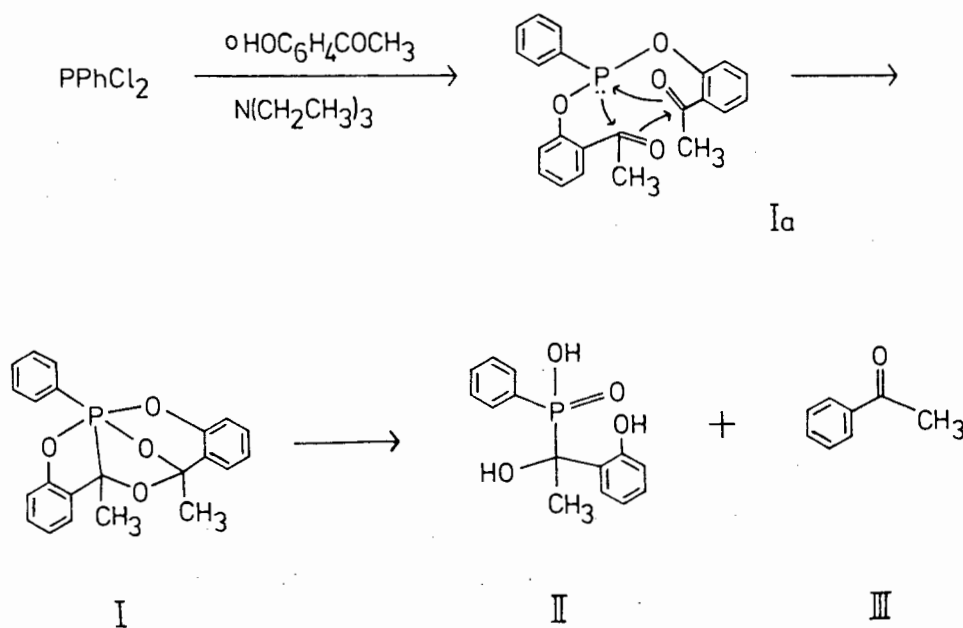
Non Hydrogen Atoms. Fractional atomic coordinates ($\times 10^4$) and anisotropic temperature factors ($\text{\AA}^2 \times 10^3$) with e.s.d.'s for compound(II).									
Thermal parameters are of the form: $T = \exp[-2\pi^2(U_{11}h^2a^{*2}+U_{22}k^2b^{*2}+U_{33}l^2c^{*2}+2U_{12}hka^*b^*+2U_{13}hla^*c^*+2U_{23}klb^*c^*) \times 10^3]$									
	x	y	z	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
P	6521(1)	584(2)	2397(1)	39(1)	30(1)	74(1)	1(1)	29(1)	1(1)
O(1)	7664(3)	1236(4)	2735(3)	39(2)	36(2)	91(2)	1(2)	33(2)	0(2)
O(2)	6194(3)	-941(4)	1597(3)	55(2)	32(2)	63(2)	-9(2)	28(2)	-5(2)
O(3)	4510(3)	3254(6)	3098(3)	71(2)	85(3)	88(3)	-24(2)	-47(2)	-8(2)
O(4)	5911(3)	3911(4)	2491(3)	57(2)	31(2)	82(2)	-11(2)	33(2)	-3(2)
C(1)	5542(4)	2441(7)	1789(4)	44(3)	33(3)	72(3)	4(3)	29(2)	5(2)
C(2)	5594(4)	3020(7)	856(4)	62(3)	46(3)	82(3)	9(3)	42(2)	0(2)
C(3)	4414(4)	1840(7)	1618(4)	34(2)	35(3)	77(3)	14(3)	28(2)	8(2)
C(4)	3811(4)	751(7)	797(4)	41(3)	48(3)	80(3)	-3(3)	20(2)	-1(2)
C(5)	2822(5)	61(8)	635(5)	53(3)	59(3)	90(3)	-6(3)	19(3)	-11(3)
C(6)	2411(5)	433(9)	1288(5)	41(3)	65(3)	114(3)	10(3)	36(3)	-5(3)
C(7)	2985(5)	1508(8)	2091(5)	50(3)	65(3)	104(3)	4(3)	47(2)	1(3)
C(8)	3988(4)	2208(8)	2279(5)	39(3)	49(3)	89(3)	11(3)	35(2)	6(2)
C(11)	6307(4)	-271(7)	3393(4)	33(2)	31(2)	67(3)	-2(2)	24(2)	-5(2)
C(21)	6919(5)	436(8)	4333(5)	65(3)	44(3)	73(3)	-7(3)	29(2)	-8(3)
C(31)	6698(5)	-145(9)	5090(5)	89(3)	81(3)	64(3)	-11(3)	31(3)	15(3)
C(41)	5914(5)	-1395(9)	4925(5)	84(3)	71(3)	88(3)	22(3)	54(3)	25(3)
C(51)	5322(5)	-2098(8)	4004(5)	55(3)	61(3)	105(3)	21(3)	41(3)	2(3)
C(61)	5523(4)	-1544(7)	3235(5)	51(3)	45(3)	72(3)	5(3)	30(2)	-2(2)

Table 3.3.1

H atoms. Fractional atomic coordinates ($\times 10^3$) for compound (II)			
	x	y	z
H(1)	532	192	33
H(2)	642	336	102
H(3)	508	419	55
H(4)	413	45	29
H(5)	237	-77	0
H(6)	164	-12	117
H(7)	265	181	259
H(21)	755	141	447
H(31)	716	40	582
H(41)	576	-184	552
H(51)	470	-309	388
H(61)	506	-211	251

3.4 Description of the structures and discussion.

A possible route for the formation of the phospho-undecane (I) would be initial reaction of two molecules of *o*-HOC₆H₄COCH₃ with PhPCl₂ to form the intermediate phosphonite I(a) (Scheme 3.3). Concerted intramolecular nucleophilic attack would then yield the tricyclic phospho-undecane. This postulated mechanism is supported by the report^{3,6} that PhP(OPh)₂ reacts with hexafluoroacetone via attack at the carbonyl carbon atom, although in this case the initial P-C-O bond rearranges to give a product containing a P-O-C bond.



Scheme 3.3

As a result of the mechanistic versatility of (I), a discussion using available data concerning the reaction pathway and possible intermediates leading to the hydrolysis products would be purely speculative. Presumably the process involves nucleophilic attack at the phosphorus atom followed by concerted cyclic proton transfer mechanisms.

The molecular structures with atomic nomenclature for compounds (I) and (II) appear in Figures 3.1 and 3.6 respectively, with bond lengths and angles for the two structures in Tables 3.4 and 3.5. The phospho-undecane (I) shows remarkable ability in accommodating the relatively crowded environment around the phosphorus atom, and this is probably due to the presence of five- and six-membered rings involving this atom. It has been mentioned^{3,8} that this factor plays a significant role in minimising prohibitive non-bonded interactions within pentacoordinated phosphorus containing molecules. The phosphorus atom in (I) lies at the centre of a distorted trigonal bipyramid (Figure 3.2), with the two five-membered rings and one six-membered ring all in apical-equatorial planes, and both carbon atoms equatorial. The apical-equatorial position for five-membered rings and the tendency for the more electronegative atoms to occupy apical rather than equatorial positions appears to be a general phenomenon in trigonal bipyramidal phosphorus.^{3,8}

In (I), the diapical bond axis is slightly distorted, with the angle $O(2)-P-O(11) = 174,9(0)^\circ$. The apical - equatorial bond angles range from $85,7(1)^\circ$ to $94,9(0)^\circ$, with the equatorial bond angles having values of $112,4(1)^\circ$, $117,4(1)^\circ$ and $130,1(1)^\circ$.

The ^1H NMR spectrum of a freshly prepared solution of (I) shows a singlet at 2,00 δ due to the protons on C(13) and a doublet at 1,65 δ ($J_{\text{HP}} = 20$ cps) due to the protons on C(12) (Figure 3.1). The spectrum run on the same solution after 24 hours contains, besides the original set of 3 lines, an additional set of 3 lines consisting also of a singlet and a doublet at values of 2,63 δ and 1,39 δ ($J_{\text{HP}} = 18$ cps) respectively. Given that the ^1H NMR spectrum of phenyl ethanone shows a singlet at 2,59 δ ,^{3.7} and assuming that in this reaction the introduction of the phosphoryl group has a shielding effect, the spectrum of the aged solutions of (I) can be interpreted in terms of the formation of the hydrolysis products (II) and (III) (Scheme 3.3).

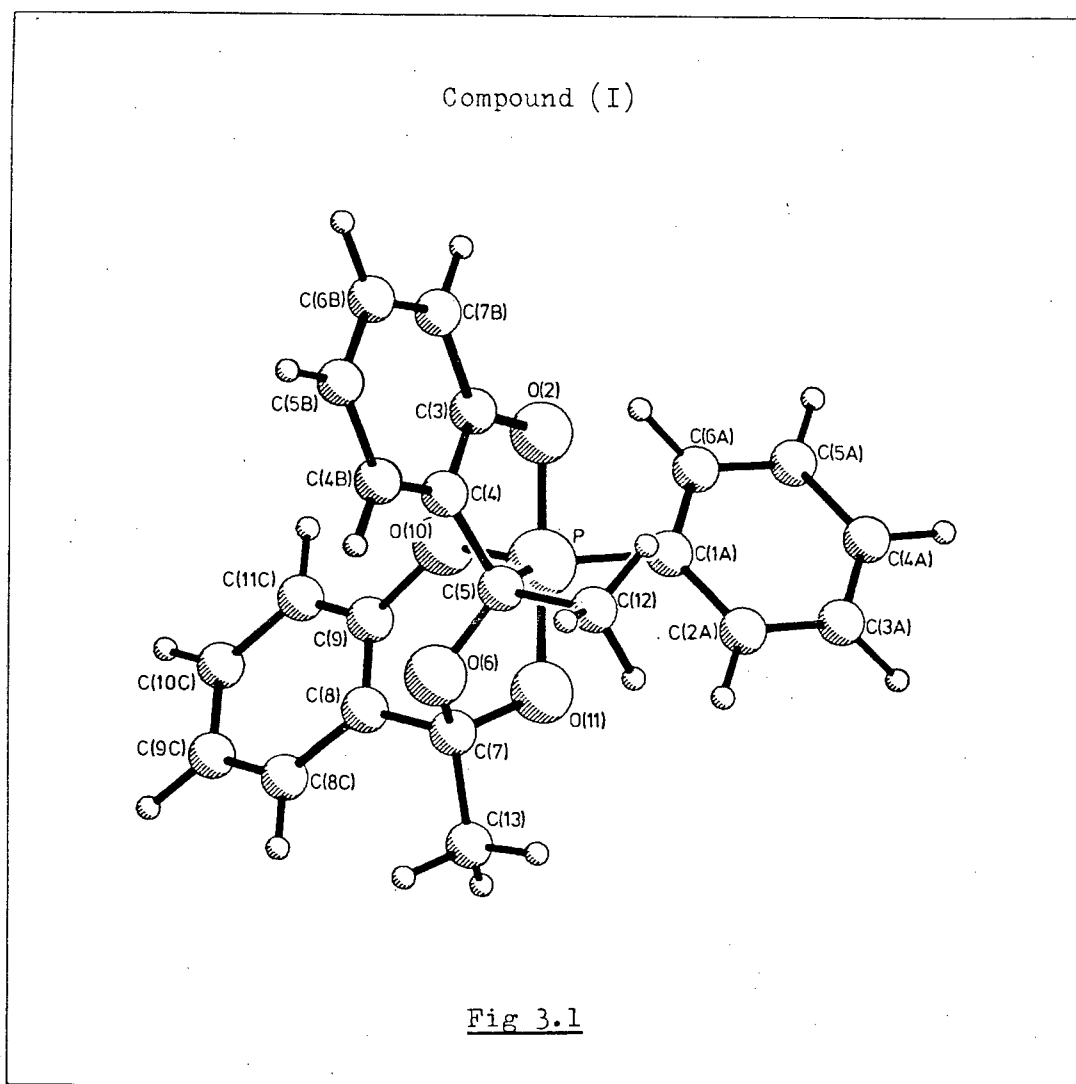


Table 3.4

Intramolecular bond lengths (\AA) and bond angles ($^\circ$) with e.s.d.'s for compound (I).			
P-O(2)	1,741(1)	C(6B)-C(7B)	1,380(2)
P-O(10)	1,626(1)	C(7B)-C(3)	1,378(2)
P-O(11)	1,696(1)	C(4)-C(5)	1,499(2)
P-C(5)	1,849(1)	C(5)-C(12)	1,533(2)
P-C(1A)	1,806(1)	C(5)-O(6)	1,421(2)
C(1A)-C(2A)	1,397(2)	O(6)-C(7)	1,471(2)
C(2A)-C(3A)	1,379(2)	C(7)-C(13)	1,512(2)
C(3A)-C(4A)	1,377(3)	C(7)-O(11)	1,402(2)
C(4A)-C(5A)	1,388(3)	C(7)-C(8)	1,519(2)
C(5A)-C(6A)	1,379(2)	C(8)-C(9)	1,391(2)
C(6A)-C(1A)	1,395(2)	C(8)-C(8C)	1,392(2)
O(2)-C(3)	1,379(2)	C(8C)-C(9C)	1,388(2)
C(3)-C(4)	1,383(2)	C(9C)-C(10C)	1,393(3)
C(4)-C(4B)	1,380(2)	C(10C)-C(11C)	1,384(2)
C(4B)-C(5B)	1,395(2)	C(11C)-C(9)	1,371(2)
C(5B)-C(6B)	1,390(2)	C(9)-O(10)	1,399(2)
O(2)-P-O(10)	88,5(1)	O(6)-C(7)-C(13)	108,5(1)
O(2)-P-O(11)	174,9(0)	O(11)-C(7)-C(13)	110,0(1)
O(10)-P-O(11)	94,9(0)	C(8)-C(7)-C(13)	115,8(1)
O(2)-P-C(5)	89,5(1)	O(6)-C(7)-C(8)	109,0(1)
O(10)-P-C(5)	112,4(1)	O(11)-C(7)-C(8)	106,3(1)
O(11)-P-C(5)	85,7(1)	C(7)-C(8)-C(9)	117,4(1)
O(2)-P-C(1A)	90,8(1)	C(7)-C(8)-C(8C)	123,9(1)
O(10)-P-C(1A)	117,4(1)	C(8C)-C(8)-C(9)	118,5(1)
O(11)-P-C(1A)	91,1(1)	O(10)-C(9)-C(8)	121,3(1)
C(1A)-P-C(5)	130,1(1)	O(10)-C(9)-C(11C)	117,1(1)
P-O(2)-C(3)	110,4(1)	C(8)-C(9)-C(11C)	121,6(1)
C(5)-O(6)-C(7)	111,6(1)	P-C(1A)-C(2A)	120,9(1)
P-O(10)-C(9)	123,7(1)	P-C(1A)-C(6A)	120,0(1)
P-O(11)-C(7)	109,1(1)	C(2A)-C(1A)-C(6A)	119,1(1)
O(2)-C(3)-C(4)	115,4(1)	C(1A)-C(2A)-C(3A)	120,3(2)

Table 3.4 continued

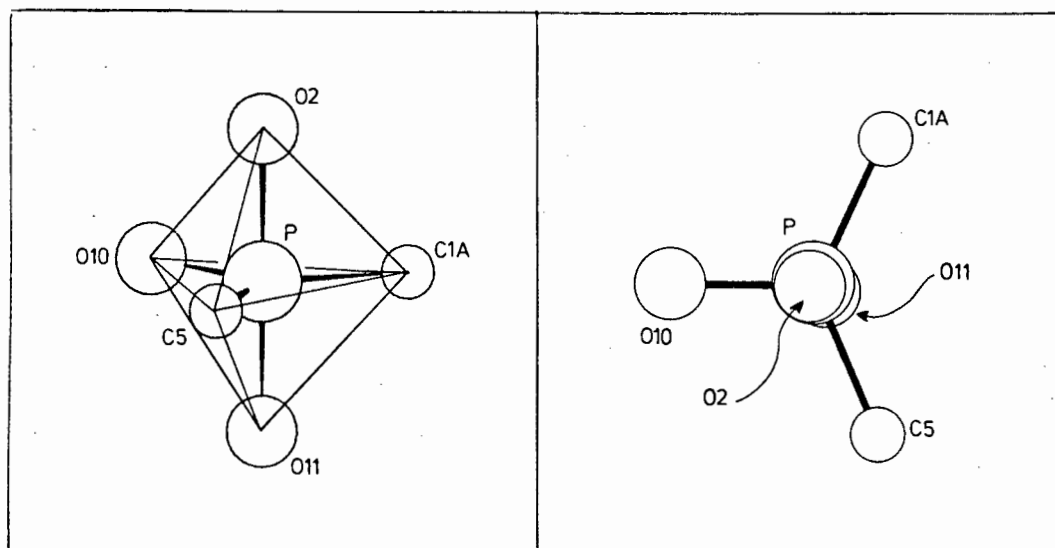
O(2)-C(3)-C(7B)	123,4(1)	C(2A)-C(3A)-C(4A)	120,4(2)
C(4)-C(3)-C(7B)	121,2(1)	C(3A)-C(4A)-C(5A)	119,8(1)
C(3)-C(4)-C(5)	111,4(1)	C(4A)-C(5A)-C(6A)	120,4(2)
C(3)-C(4)-C(4B)	120,8(1)	C(1A)-C(6A)-C(5A)	120,0(2)
C(4B)-C(4)-C(5)	127,7(1)	C(4)-C(4B)-C(5B)	118,5(1)
P-C(5)-O(6)	104,8(1)	C(4B)-C(5B)-C(6B)	120,1(1)
P-C(5)-C(4)	104,0(1)	C(5B)-C(6B)-C(7B)	121,1(1)
O(6)-C(5)-C(4)	113,3(1)	C(3)-C(7B)-C(6B)	118,3(2)
P-C(5)-C(12)	111,7(1)	C(8)-C(8C)-C(9C)	120,4(2)
O(6)-C(5)-C(12)	110,8(1)	C(8C)-C(9C)-C(10C)	119,8(1)
C(4)-C(5)-C(12)	111,7(1)	C(9C)-C(10C)-C(11C)	120,0(2)
O(6)-C(7)-O(11)	106,8(1)	C(9)-C(11C)-C(10C)	119,6(2)

Table 3.5

Intramolecular bond lengths (Å) and bond angles (°) with e.s.d.'s for compound (II)			
P-O(1)	1,509(3)	C(6)-C(7)	1,374(7)
P-O(2)	1,570(3)	C(7)-C(8)	1,389(7)
P-C(1)	1,857(5)	O(3)-C(8)	1,367(6)
P-C(11)	1,792(5)	C(8)-C(3)	1,409(7)
C(1)-C(2)	1,522(7)	C(11)-C(21)	1,399(7)
C(1)-O(4)	1,447(6)	C(21)-C(31)	1,391(7)
C(1)-C(3)	1,532(7)	C(31)-C(41)	1,361(8)
C(3)-C(4)	1,404(7)	C(41)-C(51)	1,371(8)
C(4)-C(5)	1,378(7)	C(51)-C(61)	1,387(7)
C(5)-C(6)	1,382(7)	C(61)-C(11)	1,374(6)
O(1)-P-O(2)	112,6(2)	C(3)-C(4)-C(5)	121,1(6)
O(1)-P-C(1)	110,8(2)	C(4)-C(5)-C(6)	120,0(6)
O(2)-P-C(1)	104,5(2)	C(5)-C(6)-C(7)	119,9(6)
O(1)-P-C(11)	111,9(2)	C(6)-C(7)-C(8)	121,4(6)
O(2)-P-C(11)	108,0(2)	O(3)-C(8)-C(3)	123,0(5)

Table 3.5 continued

C(1)-P-C(11)	108,6(2)	O(3)-C(8)-C(7)	117,8(6)
P-C(1)-O(4)	104,4(3)	C(3)-C(8)-C(7)	119,2(6)
P-C(1)-C(3)	109,5(3)	P-C(11)-C(21)	119,0(4)
O(4)-C(1)-C(3)	109,6(4)	P-C(11)-C(61)	120,9(5)
P-C(1)-C(2)	110,0(4)	C(21)-C(11)-C(61)	120,0(5)
O(4)-C(1)-C(2)	109,5(4)	C(11)-C(21)-C(31)	118,7(6)
C(2)-C(1)-C(3)	113,5(5)	C(21)-C(31)-C(41)	120,9(7)
C(1)-C(3)-C(8)	123,4(5)	C(31)-C(41)-C(51)	120,3(7)
C(1)-C(3)-C(4)	118,0(5)	C(41)-C(51)-C(61)	120,1(6)
C(4)-C(3)-C(8)	118,4(5)	C(51)-C(61)-C(11)	120,0(6)

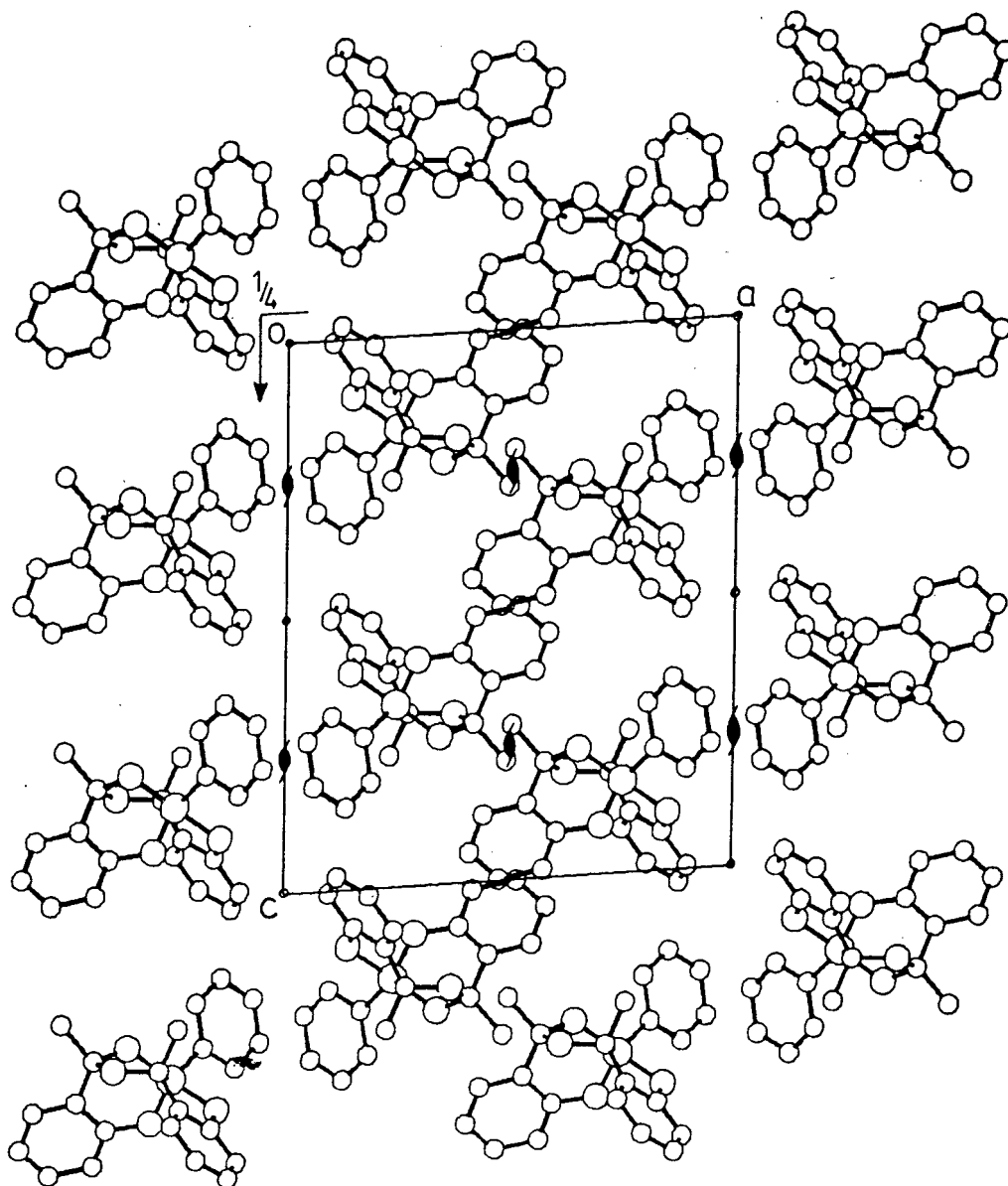


Trigonal bipyramidal geometry about the phosphorus atom in compound (I).

Fig 3.2

The phosphorus atom lies $0,01 \text{ \AA}$ from the equatorial plane. The phenyl groups have normal geometry. The two five-membered rings are slightly puckered, the P-C(5)-O(6)-C(7)-O(11) ring containing two tetrahedral carbon atoms showing the greater deviation from planarity. Torsion angles in the six-membered dioxaphosphorinene ring vary between 76° for P-O(11)-C(7)-C(8) and -10° for the C(7)-C(8)-C(9)-O(10). A projection of the unit cell looking down the b axis is shown in Figure 3.3.

It is well established that many pentacoordinate phosphorus compounds are able to change their ligand distributions by bond deformation processes which may be explained using the Berry pseudorotation^{3.9} or the turnstile rotation mechanisms.^{3.8} A schematic representation of the two mechanisms is depicted in Figure 3.4. In Berry pseudorotation, pairs of ligands move in planes perpendicular to each other, the planes being

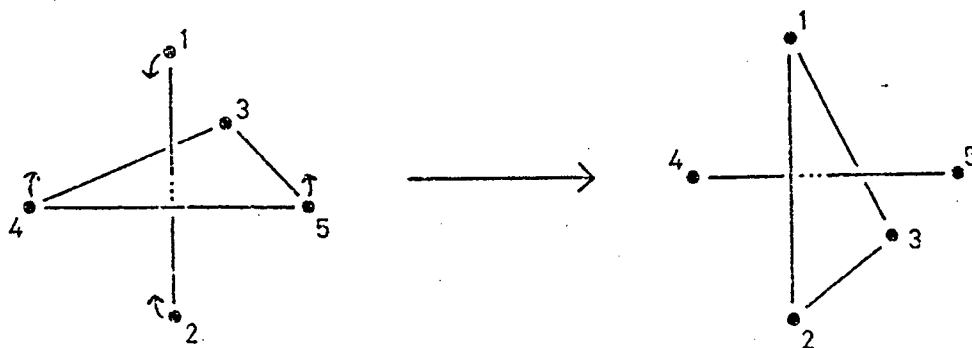


(010) projection of compound(I).

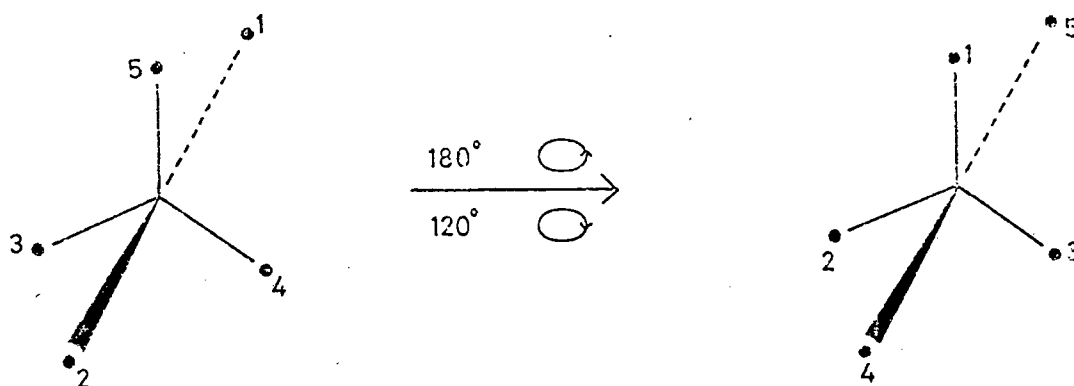
Fig 3.3

defined by the phosphorus atom, the corresponding pairs of ligands, and the fixed equatorial ligand or pivot. The nett result is an interchange of the two apical ligands into positions that are equatorial in the trigonal bipyramid, but

there is no internal rotation of some part of the molecule relative to another.



BERRY PSEUDOROTATION



TURNSTILE ROTATION

Fig 3.4

As regards turnstile rotation, referring to Figure 3.4, the orientation of the trigonal bipyramid emphasises that the apical ligand 1 and the equatorial ligand 5 constitute the pair, while the apical ligand 2 and the equatorial ligands 3 and 4 are the *trio* in this mechanism. The isomerism is achieved by first transposing the pair ligands (180° rotation), and then rotating the *trio* ligands 120° either clockwise or

anti-clockwise depending on which ligand of the *trio* is to remain equatorial. As can be verified by choosing alternative appropriate ligands as the pair, the example depicted in Figure 3.4 is only one of four equivalent turnstile rotations that accomplishes what one Berry pseudorotation does, when ligand 3 is the pivot.

The schematic representation of the movements of atoms taking place in the turnstile rotation mechanism as discussed is highly misleading in that their actual motion involves a series of angular compressions and expansions, tilting of the ligand pair, and small rotations of the pair relative to the *trio* in opposite directions in realising the same isomerisation. These movements are such as to minimise energy changes in the model situations, and is discussed more fully in the review article by Ramirez and Ugi.^{3.8}

Hence the turnstile rotation mechanism, and its extensions, the double and triple turnstile rotations, are essentially alternative processes that accomplish all that the Berry pseudorotation does, but in addition are better able to explain the permutational isomerisations observed in certain cyclic derivatives of pentacoordinated phosphorus compounds which Berry pseudorotation could not do without involving energetically unfavourable intermediates. Energy considerations make it less favourable for atom motions to occur which result in the formation of the intermediate, than other motions which result in the formation of the stable isomer.

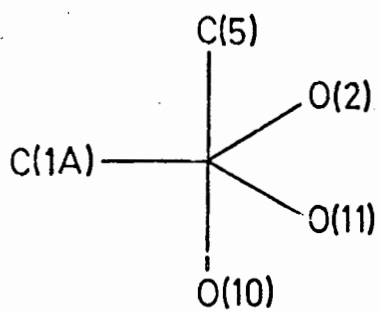
Five-coordinated phosphorus having trigonal bipyramidal symmetry, D_{3h} , can exist as 20 isomers, or 10 pairs of enantiomers, when all the ligands are different. Application

to the phospha-undecane (I) shows that, when one considers one of each of the enantiomeric pairs, as shown in Figure 3.5(a)-(j) only one is plausible. The other nine exhibit serious drawbacks to their possible formation due to one or more of the following factors:

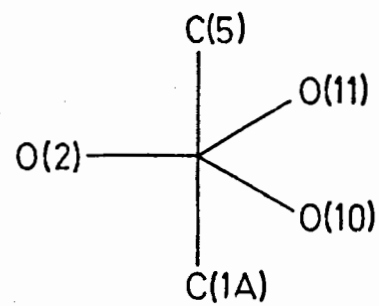
- (i) the plane of one or both five-membered rings lie in diequatorial or diapical positions,
- (ii) one or both of the apical positions are occupied by a carbon atom which has a lower electronegativity than an oxygen atom,
- (iii) serious steric hindrances exist due to the enforced orientation of bulky groups about the central phosphorus atom.

Hence, referring to Figure 3.5,

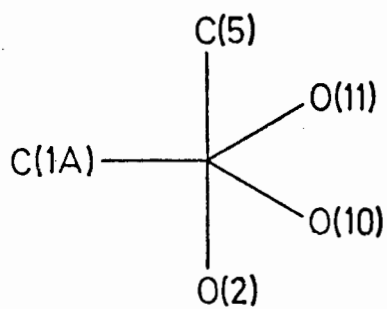
- (a) C(5) is apical. A model shows prohibitive close contacts involving the phenyl rings precluding this isomer on steric grounds alone; its formation is therefore considered improbable.
- (b) C(5) and C(1A) are both apical; its formation is again improbable.
- (c) C(5) is apical; the five-membered P-C(5)-C(4)-C(3)-O(2) ring is in a diapical plane; improbable.
- (d) C(5) is apical; the five-membered P-C(5)-O(6)-C(7)-O(11) ring is in a diapical plane; improbable.
- (e) C(1A) is apical; both five-membered rings are in the diequatorial plane; improbable.
- (f) The P-C(5)-O(6)-C(7)-O(11) ring is diequatorial; improbable.
- (g) The P-C(5)-C(4)-C(3)-O(2) ring is diequatorial; in addition the six-membered P-O(10)-C(9)-C(8)-C(7)-O(11) ring is diapical; improbable.



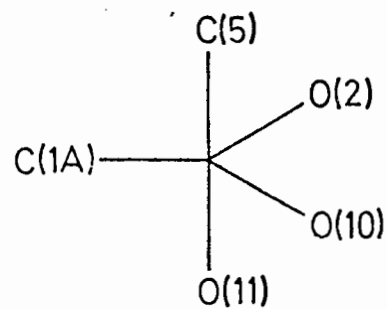
a



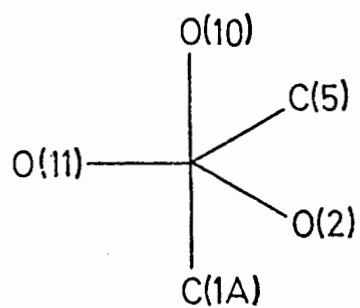
b



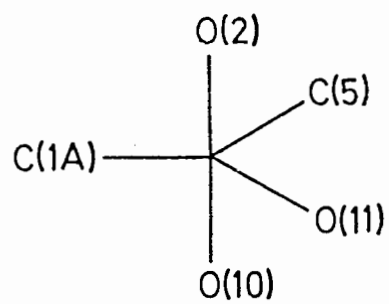
c



d



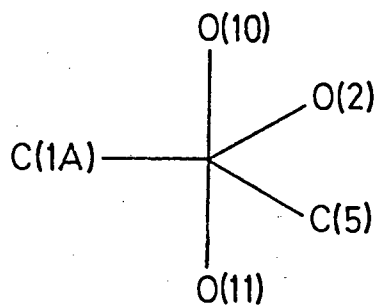
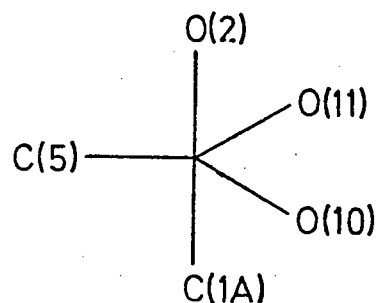
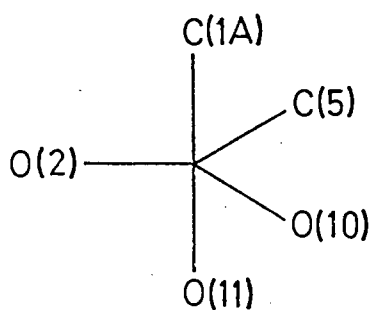
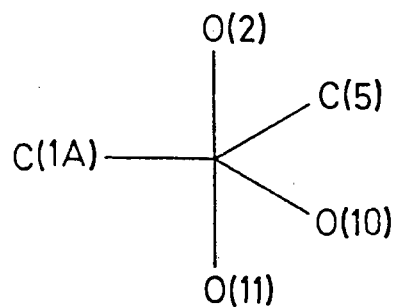
e



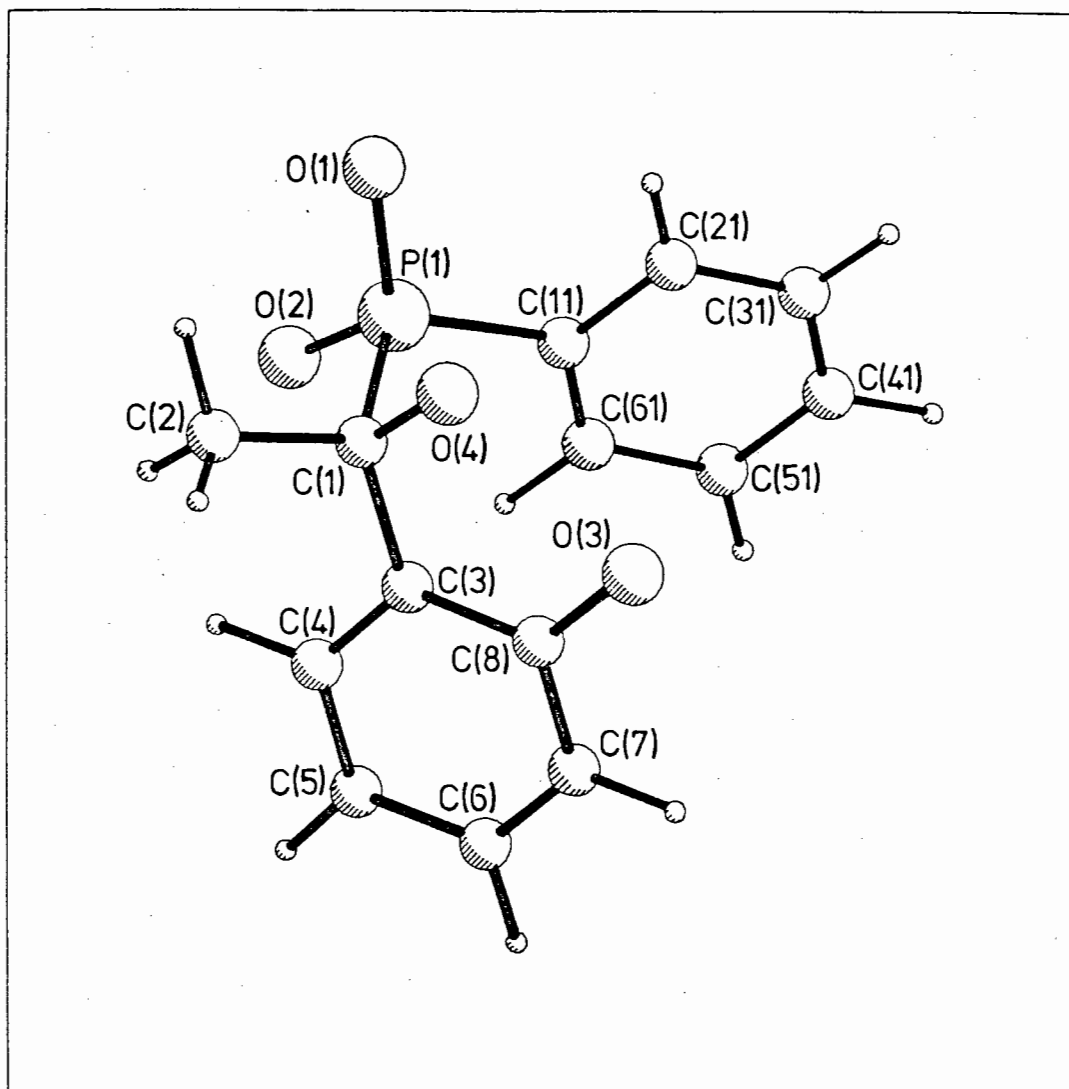
f

Enantiomeric forms of compound(I).

Fig 3.5

ghijFig 3.5 (contd.)

- (h) C(1A) is apical; the P-C(5)-O(6)-C(7)-O(11) ring is diequatorial; improbable.
- (i) C(1A) is apical; the P-C(5)-C(4)-C(3)-O(2) ring is diequatorial; improbable.
- (j) This isomer is found in the crystal structure; both apical positions are occupied by oxygen atoms, both five-membered rings are in apical-equatorial planes, and there are no serious steric hindrances.



Compound(II)

Fig 3.6

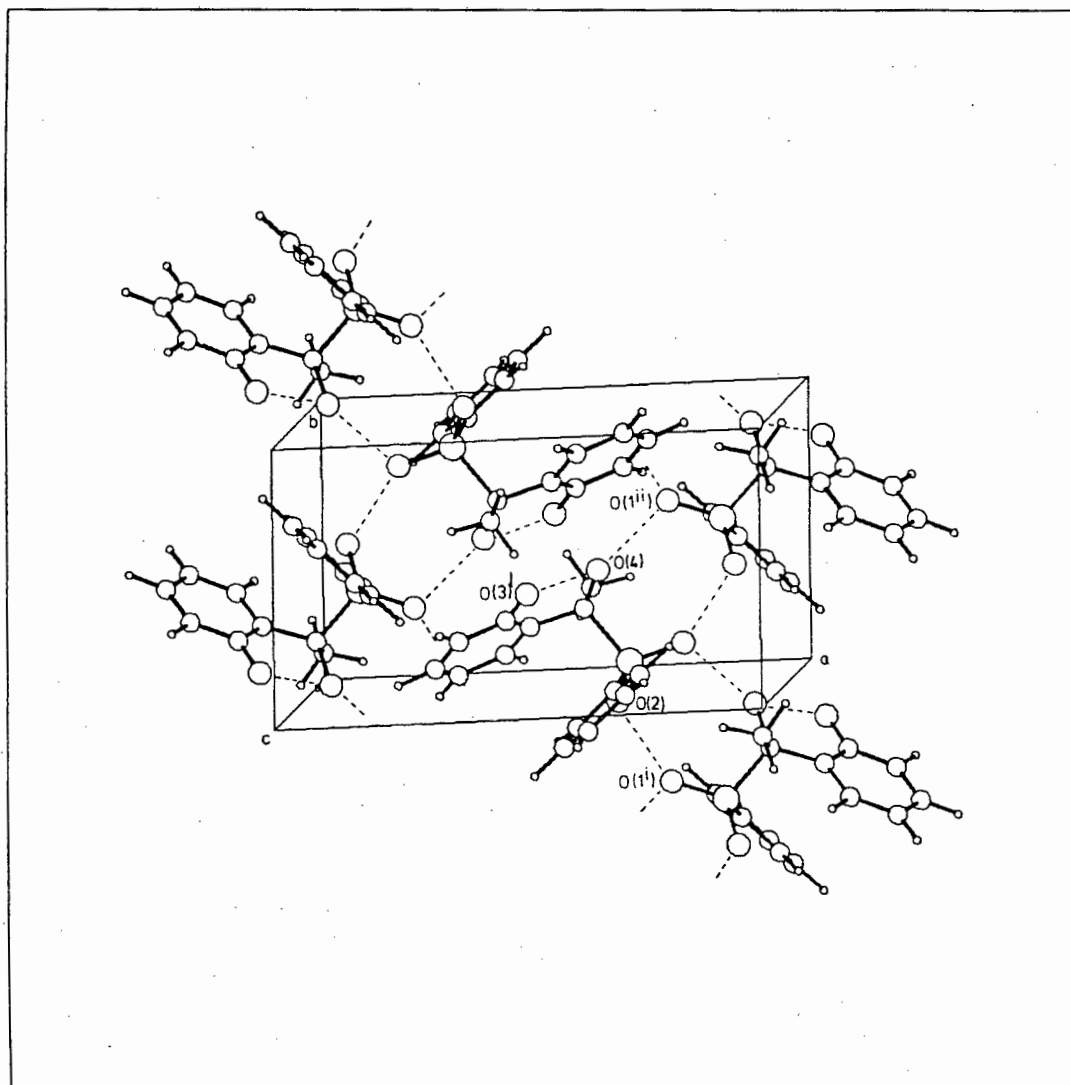
The molecular rigidity of (I) largely precludes any significant positional changes or rotations of a pair of ligands relative to a trio without rupture of bonds, and therefore the likelihood of permutational isomerism taking place in solution for this compound, either by Berry pseudorotation or by turnstile rotation mechanisms, is considered remote. This conclusion is supported by examination of the ^1H NMR spectrum of (I), whose absorption pattern is temperature independent throughout the range -60°C to 65°C .

The ligands around the phosphorus atom in the disubstituted phosphinic acid (II) are in the expected tetrahedral environment. The P-C(11) bond length of $1,792(5) \text{ \AA}$ agrees favourably with the average P-C bond length of $1,786(4) \text{ \AA}$ as determined in diphenylphosphinic acid.^{3.10} The P-C(1) bond length is a somewhat longer $1,857(5) \text{ \AA}$.

The presence of O...O close contacts (less than $2,76 \text{ \AA}$) are indicative of hydrogen bonding which extends throughout the lattice (Table 3.6). Although the hydroxyl hydrogens could not be located, the P-O(1) bond distance of $1,509(3) \text{ \AA}$, typical of P=O linkage, and the P-O(2) bond distance of $1,570(3) \text{ \AA}$, typical of P-OH linkage,^{3.11} allow a reasonable assessment to be made concerning the donor and acceptor oxygen atoms involved in the H-bonding scheme; O(1) acts as hydrogen acceptor as regards both the O(2)...O(1) and O(4)...O(1) intermolecular bonds, the hydrogen in the O(3)...O(4) intramolecular bond being donated by O(3). This is illustrated in Figure 3.7.

Table 3.6

Close intramolecular and intermolecular approaches (Å) with e.s.d.'s for compound (II)		
Symmetry code: (i) $1\frac{1}{2} - x, -\frac{1}{2} + y, \frac{1}{2} - z$		
(ii) $1\frac{1}{2} - x, \frac{1}{2} + y, \frac{1}{2} - z$		
O(2) O(1 ⁱ)		2,536(5)
O(4) O(1 ⁱⁱ)		2,756(6)
O(3) O(4)		2,551(7)



Unit cell contents of compound(II) showing probable H-bonding

Fig 3.7

CHAPTER 4

CRYSTAL STRUCTURE AND MOLECULAR CONFORMATION OF 2,3-DICYCLOPROPYL-1,4-NAPHTHOQUINONE

ABSTRACT

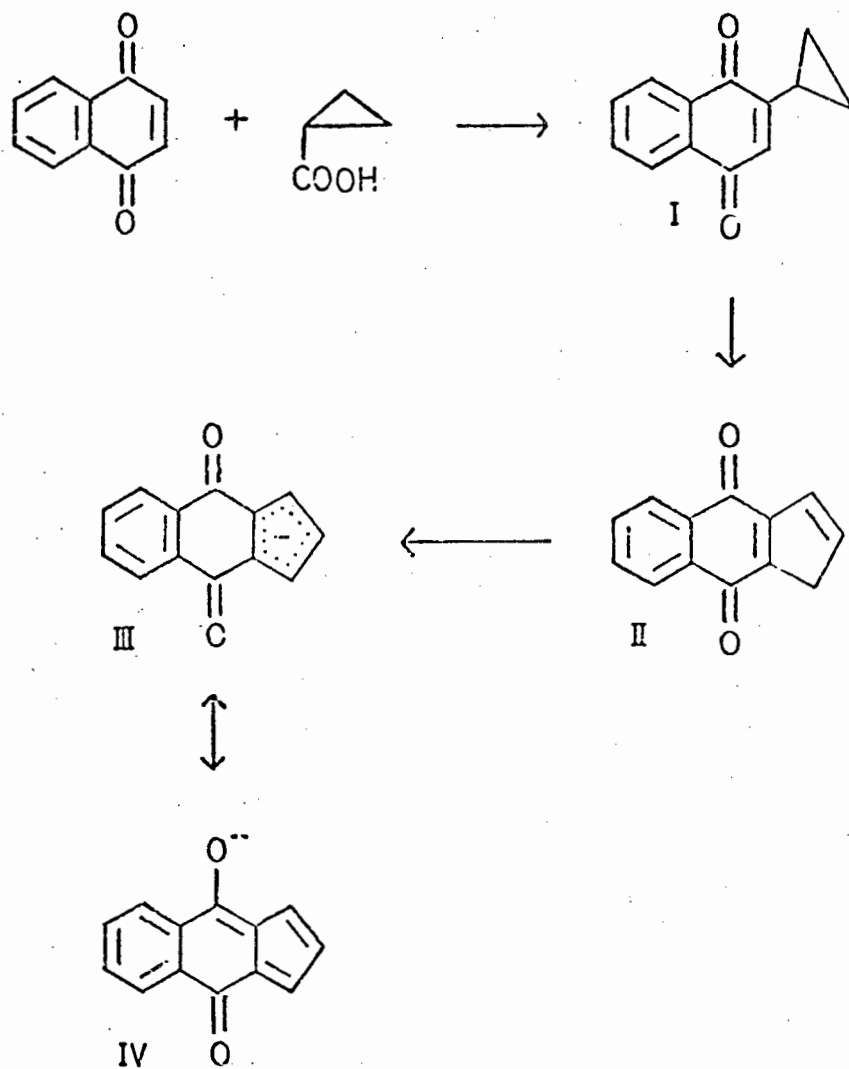
Crystals of the title compound are orthorhombic with $\underline{a} = 8,483(4)$, $\underline{b} = 18,249(9)$, $\underline{c} = 15,889(8)$ Å, space group \underline{Pbca} , $\underline{z} = 8$. The quinone ring shows distinct distortions from planarity. The solution NMR spectrum of the compound at various temperatures suggest a measure of restricted rotation about the cyclopropyl - quinonoid bonds. This is partially substantiated by a conformational analysis using the method of atom pair potentials.

4.7 Introduction

A molecule satisfying the so-called Hückel rule (or $4n + 2$ rule) is well known to have an unusually high stability.^{4.1} Thus the considerable stability possessed by the cyclopentadienide ion has been ascribed to it having six π electrons ($n = 1$). The synthesis of benz[f]indene-4,9-quinone (II) was attempted with a view to study the chemistry of its anion (III), which is isoelectronic with anthroquinone. The anion, in addition to delocalisation of the negative charge about the five-membered ring, could also be stabilised by the quinone functional group in forming the canonical form (IV). The route that was proposed in the attempted synthesis of (III) was as shown in Scheme 4.1.^{4.2}

Reaction of 1,4-naphthoquinone with cyclopropyl carboxylic acid in the presence of silver ions and peroxodisulphonate would yield 2-cyclopropyl-1,4-naphthoquinone (I). The thermal and photochemical isomerisations of vinylcyclopropanes to cyclopentenes are well established processes and similar treatment of (I) would hopefully yield the benz[f]indene-4,9-quinone. Reaction of this with base would then afford the desired anion.

An early fraction in the chromatographed products of the first stage in the above reaction scheme yielded 2,3-dicyclopropyl-1,4-naphthoquinone in the form of yellow crystals. The interesting nature of the ^1H NMR spectrum at room temperature of this by-product, which exhibited a highly symmetrical pattern about the methylene and methine proton absorption region, prompted further investigations as regards its structure and conformation.



Scheme 4.1

4.2 Crystal preparation and data collection

Ammonium peroxodisulphonate (6,2g) in water (50 ml) was added at a bath temperature of 80-85°C to a stirred mixture of cyclopropyl carboxylic acid (0,95 g), silver nitrate (1g) and naphthoquinone (1,58 g) in water (25 ml) and cyclohexane (25 ml) over a period of 45 minutes. The reaction mixture was stirred for 10 minutes after addition was completed, and then worked up by cooling and extracting with ether. The ether extract was washed with 10% sodium bicarbonate solution until excess acid had been removed, and then once with water. The ethereal solution was dried with sodium sulphate and evaporated off under reduced pressure. The resulting brown oil (1,87 g) was chromatographed on a silica column using 2½ - 10% ethyl acetate/light petroleum ether as eluant. An early fraction afforded 2,3-dicyclopropyl-1,4-naphthoquinone as yellow needles (0,62 g, 26%). δ 1,08 (8H, d, J 8Hz, cyclopropyl CH₂), 2,00 (2H, quintet, J 8Hz, cyclopropyl CH), 7,65 (2H, m, 6- and 7-H) and 8,08 (2H, m, 5- and 8-H). A later fraction afforded the 2-cyclopropyl-1,4-naphthoquinone analogue as the major product (0,96 g, 48,5%).

Preliminary oscillation Weissenberg and precession photographs gave the crystallising space group as Pbca, the conditions limiting possible reflections being^{2.8}

$$okl : k = 2n$$

$$hol : l = 2n$$

$$hko : h = 2n$$

$$hoo : h = 2n$$

$$oko : k = 2n$$

$$ool : l = 2n$$

Crystal data and experimental details of the data collection are listed in Table 4.1.

Table 4.1

Crystal data and experimental and refinement parameters for the structure analysis		
<u>Crystal data</u>	<u>Data collection</u>	<u>Final refinement</u>
Molecular formula	Crystal dimensions	Number of variables
$C_{16}H_{14}O_2$	0, 1x0, 1x0, 5mm	106
M_r	Scan mode	$R = \Sigma F_o - F_c / \Sigma F_o $
238	$\omega - 2\theta$	0,096
Space group	Scan width	$R_w = \Sigma w^{1/2} F_o - F_c / \Sigma w^{1/2} F_o $
$Pbca$	1, 1 $^\circ\theta$	0,071
a	Scan speed	Weighting scheme
8,483(4) \AA	0,03 $^\circ \theta \text{ s}^{-1}$	$w = (\sigma^2 F)^{-1}$
b	Range scanned(2 θ)	U (aromatic H)
18,249(9)	6 $^\circ - 40^\circ$	U (methine H)
c	Stability of standard reflections	U (methylene H)
15,889(8)	0,84%	
V	No. of reflections collected	
2459,7 \AA^3	1358	
D_m	No. of 'observed' reflections	
1,27Mg m $^{-3}$	489 with I(rel)	
Dc	>2 σ I(rel)	
μ (MoK α)		
0,047 mm $^{-1}$		
F(000)		
1008		

4.3 Solution and refinement of the structure

The structure was solved by the automatic centrosymmetric direct-methods routine of the SHELX program system,^{1.1} in which an E map yielded the positions of all the heavy atoms. Due to the paucity of observed reflections, the structure refinement proceeded with isotropic temperature factors for the non-H atoms; a subsequent difference Fourier indicated the positions of most of the hydrogen atoms. These were then included in the model by the technique of constrained refinement, with $d(\text{C-H}) = 1,08 \text{ \AA}$, their positions being dictated by the geometry of the molecule. The positions of the two methine hydrogens H(21) and H(31) derived in this way, however, were deemed unsatisfactory because they gave rise to unacceptable angles between the C-(methylene H) bond and adjacent C-C bonds. The methine hydrogen atoms were therefore omitted from a structure factor calculation and their positions found in a difference electron density map. All H atoms in the cyclopropyl rings were finally allowed to refine with a simple bond length constraint of $1,08 \text{ \AA}$ from their parent atoms. The final geometry of the cyclopropyl moieties compared favourably with that previously found in N,N-dimethyl-2-phenylcyclopropylamine hydrochloride^{4.3} and α -cyano-3-phenoxybenzyl-*cis*-3-(2,2-dibromovinyl)-2,2-dimethylcyclopropanecarboxylate^{4.4} in which no constraints had been applied to the hydrogens. The isotropic temperature factors of all the H atoms were treated as three single parameters, whose final values are reported in Table 4.1. Final atomic parameters, principal bond lengths and angles are shown in Tables 4.2, 4.3 and 4.4 respectively. Observed and calculated structure factors appear in Appendix E.

Table 4.2

Fractional atomic coordinates ($\times 10^4$) and temperature factors ($\text{\AA}^2 \times 10^3$) and their e.s.d.'s in parenthesis				
	x	y	z	U
C(1)	23(16)	3167(7)	2039(8)	46(4)
C(2)	29(14)	2879(7)	2922(8)	39(4)
C(21)	-668(16)	2143(8)	3074(8)	53(4)
C(22)	356(18)	1574(9)	3475(11)	75(5)
C(23)	18(19)	1509(9)	2554(11)	71(4)
C(3)	535(14)	3315(7)	3563(9)	41(4)
C(31)	276(18)	3088(8)	4433(9)	60(5)
C(32)	17(19)	3629(8)	5178(9)	65(5)
C(33)	1531(20)	3205(8)	5133(10)	72(5)
C(4)	1358(16)	4024(8)	3382(8)	56(5)
C(41)	1624(15)	4242(6)	2474(9)	43(4)
C(5)	2551(16)	4845(7)	2300(8)	56(4)
C(6)	2717(17)	5049(6)	1451(8)	64(5)
C(7)	1988(17)	4649(7)	828(9)	70(5)
C(8)	1076(18)	4036(8)	983(9)	67(5)
C(81)	911(15)	3829(7)	1843(8)	42(4)
O(1)	-761(10)	2841(5)	1512(6)	70(3)
O(4)	1854(10)	4425(5)	3930(6)	81(4)

Table 4.2.1

Hydrogen atoms.	Fractional atomic coordinates ($\times 10^3$).		
	x	y	z
H(21)	-192	219	320
H(221)	-20	119	390
H(222)	159	168	358
H(231)	-114	132	235
H(232)	94	157	209
H(31)	-5	254	464
H(321)	-83	337	559
H(322)	-5	420	502
H(331)	150	272	553
H(332)	256	353	499
H(5)	309	516	280
H(6)	347	551	129
H(7)	210	484	19
H(8)	54	373	48

Table 4.3

Bond lengths (Å) and their e.s.d.'s			
O(1)-C(1)	1,223(13)	C(3)-C(4)	1,497(15)
C(1)-C(2)	1,498(12)	O(4)-C(4)	1,212(13)
C(2)-C(21)	1,488(16)	C(4)-C(41)	1,513(16)
C(21)-C(22)	1,495(19)	C(41)-C(5)	1,381(14)
C(21)-C(23)	1,536(19)	C(5)-C(6)	1,406(15)
C(22)-C(23)	1,496(21)	C(6)-C(7)	1,377(14)
C(2)-C(3)	1,362(14)	C(7)-C(8)	1,383(16)
C(3)-C(31)	1,460(17)	C(8)-C(81)	1,423(14)
C(31)-C(32)	1,556(19)	C(41)-C(81)	1,393(15)
C(31)-C(33)	1,554(19)	C(1)-C(81)	1,458(15)
C(32)-C(33)	1,501(20)		

Table 4.4

Selected bond angles (°) and their e.s.d.'s.			
O(1)-C(1)-C(2)	118,2(1,3)	C(31)-C(33)-C(32)	61,2(1,0)
C(21)-C(2)-C(3)	122,1(1,3)	C(32)-C(31)-C(33)	57,7(0,9)
C(31)-C(3)-C(4)	119,8(1,2)	H(21)-C(21)-C(2)	110,4(6,2)
O(4)-C(4)-C(41)	118,3(1,3)	H(21)-C(21)-C(22)	123,0(6,3)
C(4)-C(41)-C(5)	119,0(1,4)	H(221)-C(22)-C(21)	117,2(6,5)
C(41)-C(5)-C(6)	117,4(1,3)	H(222)-C(22)-C(23)	110,9(7,2)
C(5)-C(6)-C(7)	120,3(1,3)	H(231)-C(23)-C(21)	93,0(6,6)
C(6)-C(7)-C(8)	123,5(1,4)	H(232)-C(23)-C(22)	121,5(6,7)
C(7)-C(8)-C(81)	116,2(1,4)	H(31)-C(31)-C(3)	125,7(6,5)
C(8)-C(81)-C(1)	118,4(1,3)	H(31)-C(31)-C(32)	108,7(6,4)
C(21)-C(22)-C(23)	61,8(1,0)	H(321)-C(32)-C(31)	105,6(6,8)
C(21)-C(23)-C(22)	59,1(1,0)	H(322)-C(32)-C(33)	122,2(6,8)
C(22)-C(21)-C(23)	59,1(0,9)	H(331)-C(33)-C(31)	106,8(6,7)
C(31)-C(32)-C(33)	61,1(1,0)	H(332)-C(33)-C(32)	114,5(6,7)

4.4 Description of the structure and discussion

Figure 4.1 shows the molecule viewed perpendicular to the two aromatic rings. A distinctive feature of the structure is the considerable distortion from planarity occurring in the quinone ring, as shown by the endocyclic torsion angles given in Figure 4.2.

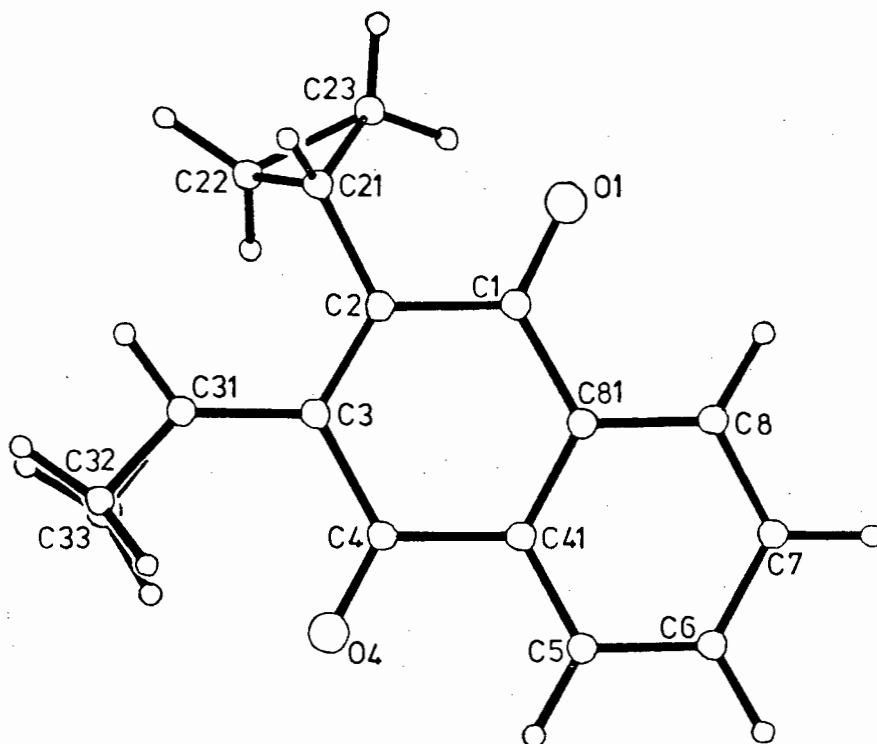


Fig 4.1

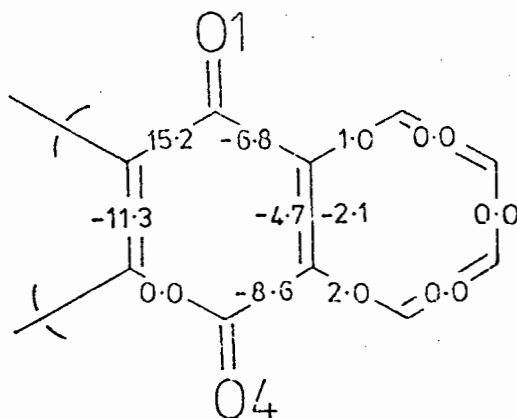


Fig 4.2

This is probably due to the intramolecular close contacts C(23)...C(1) of 3,13 Å and C(23)...O(1) of 3,01 Å. Thus C(1) and O(1) are displaced 0,14 Å and 0,39 Å respectively from the least squares plane containing C(2), C(3), C(4), C(41) and C(81). There are no significant intermolecular close contacts.

A conformational study of the compound was carried out by calculating the van der Waal's energy using empirical atom pair potentials. This method of evaluating molecular energies is well established and has recently been reviewed by Kitaigorodsky^{4.5} and Mirsky^{4.6}. The coefficients of the atom-atom potentials are of the form

$$u(r) = a \exp(-br)/r^d - c/r^6$$

where r is the distance between any pair of atoms and the coefficients a, b, c and d are those given by Giglio,^{4.7} such that $u(r)$ is evaluated in calories when r is in Å. These potential curves were derived primarily to give good agreement for the calculation of molecular positions in crystal structures and the energy values mean little in an absolute sense. No account was taken of possible torsional potentials for the σ bonds, partial atomic charges or dipole interactions. The program EENY^{4.8} calculates molecular coordinates as defined by appropriate torsion angles and sums the energy for all non-bonded pairs of atoms whose separation is dependent on the torsion angles.

In the present study, if an idealised structure in which the aromatic and quinonoid rings are exactly planar and the cyclopropyl moieties are identical is assumed, and the molecule is divided into three residues linked by two torsion angles:

τ_1 [C(1)-C(2)-C(21)-H(21)] and τ_2 [C(4)-C(3)-C(31)-H(31)] as shown in Figure 4.3, then for equal but opposite values of τ_1 and τ_2 , the molecule possesses C_s symmetry, the mirror plane bisecting residue 1 and lying perpendicular to the aromatic and quinonoid rings.

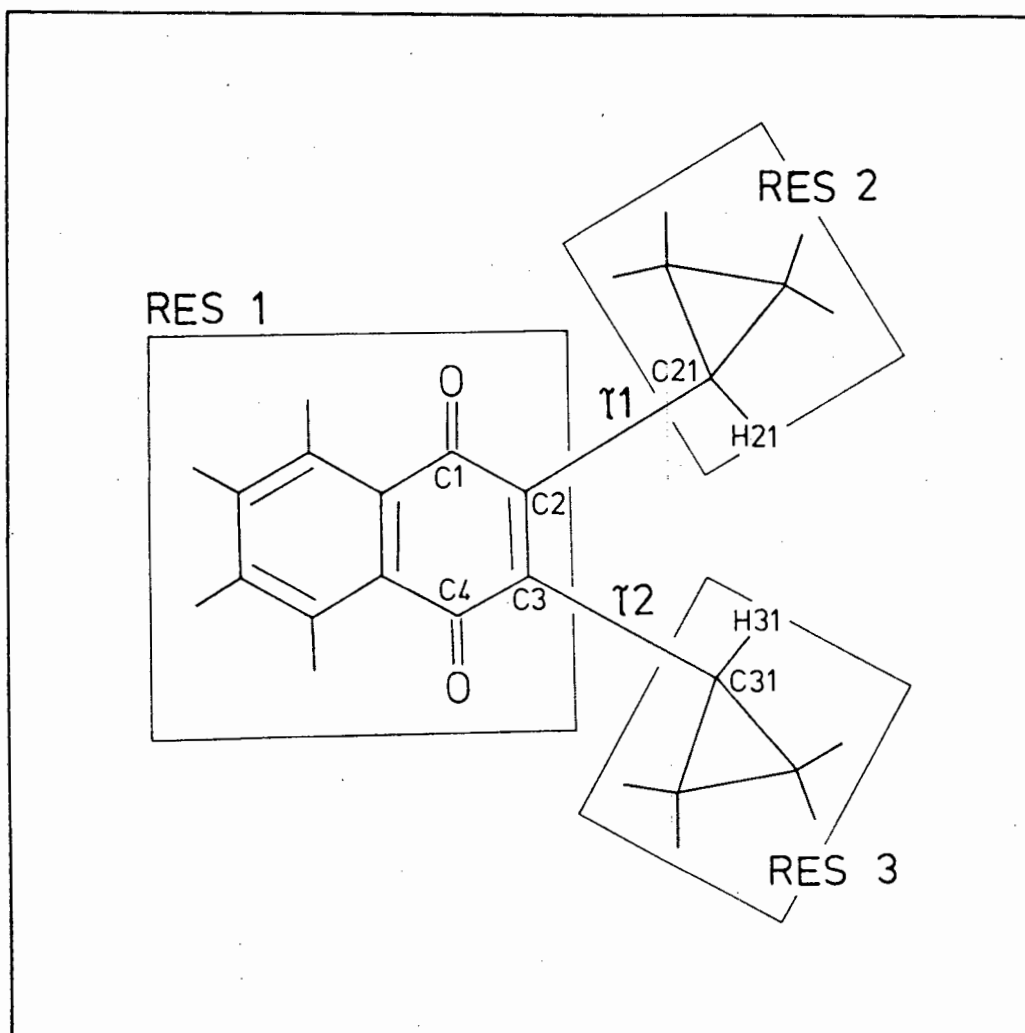


Fig 4.3

Hence any conformational arrangement defined by torsion angles τ_1 and τ_2 which gives rise to an energy minimum will have an equivalent arrangement at values of $-\tau_1$ and $-\tau_2$, which is its mirror image. The atomic coordinates used in the energy

analysis however were not for the idealised molecule, but those obtained from the refined crystal structure. An energy map was calculated with both the torsion angles varying over the complete angular range $0-360^\circ$ in 10° increments. This map, shown in Figure 4.4, yielded six energy minima labelled A,B,C and A',B',C'. These are displayed as a three-dimensional energy surface in Figure 4.5 for greater clarity.^{4.10}

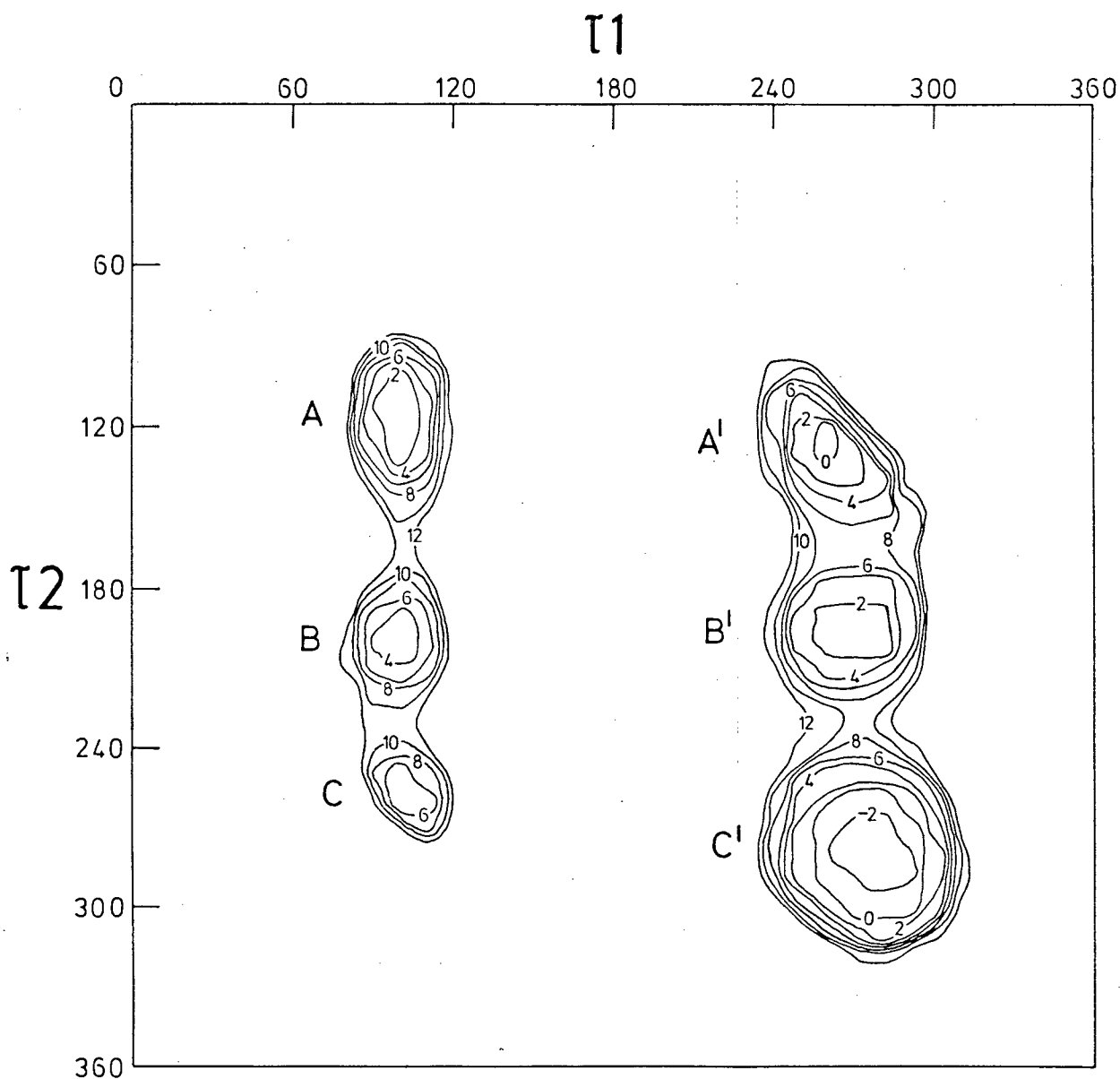


Fig 4.4

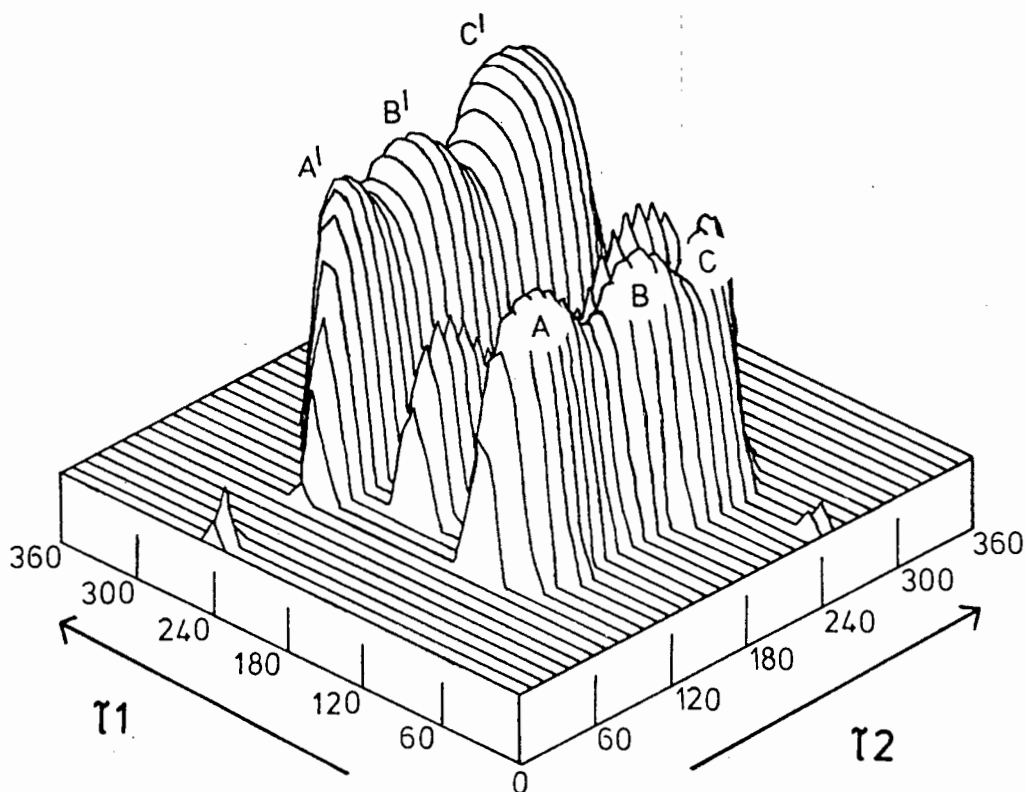
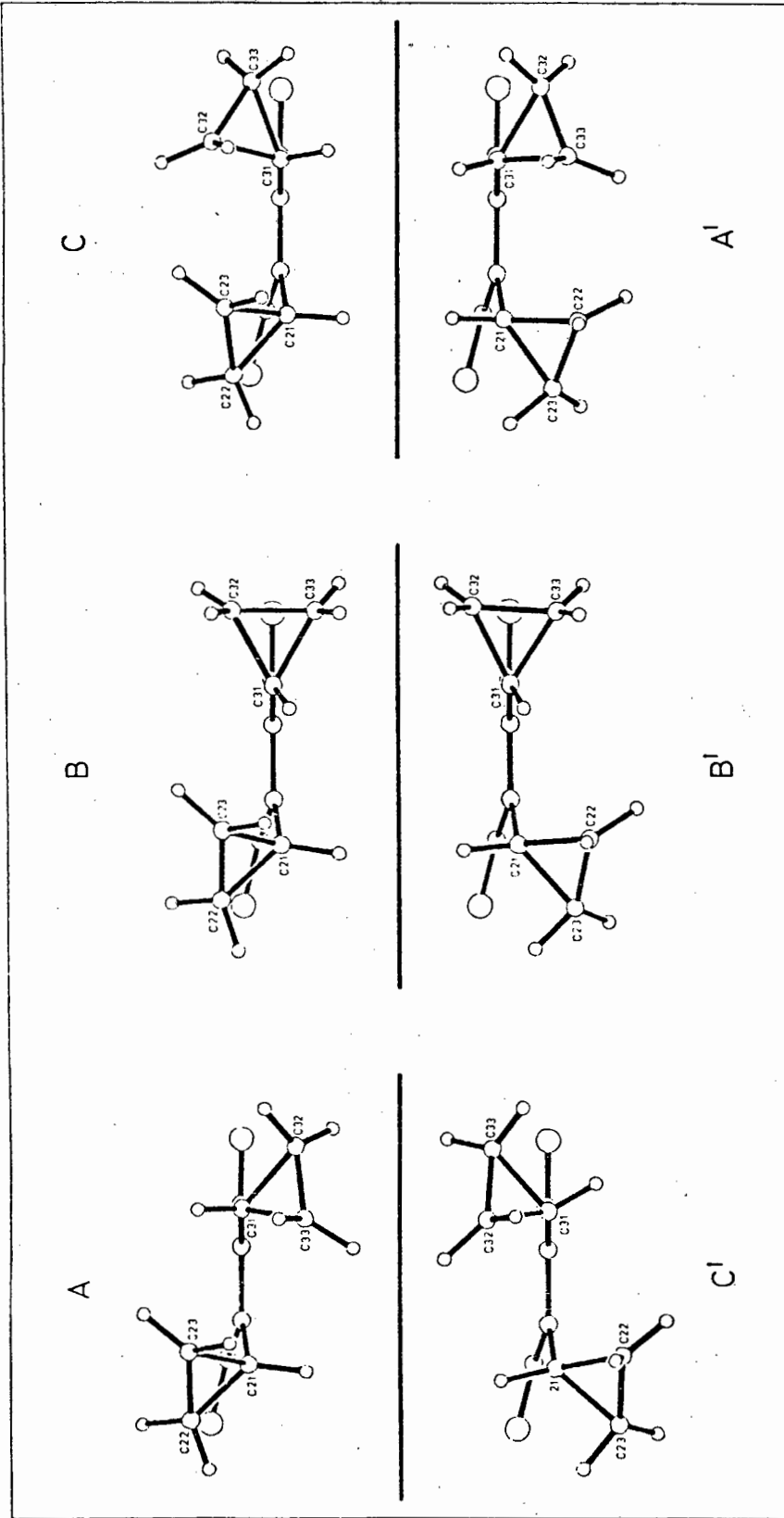


Fig 4.5

The torsion angles at the exact minima were obtained using the method of steepest descent incorporated in the program, and these are shown in Table 4.5 together with calculated energies at these positions. Energy barriers between the different conformations are of the order of 40 kJ mol^{-1} .

The peaks obtained correspond to the molecular conformations shown in Figure 4.6, which do indeed display mirror image pairs, although mirror symmetry is inexact. The conformation in the crystal structure with $\tau_1 = 271,3^\circ$ and $\tau_2 = 195,4^\circ$



Molecular conformations corresponding to the energy minima. The view is along a line joining the midpoints of C(2)—C(3) and C(6)—C(7). Aromatic H atoms and selected C atoms have been omitted for clarity.

Fig 4.6

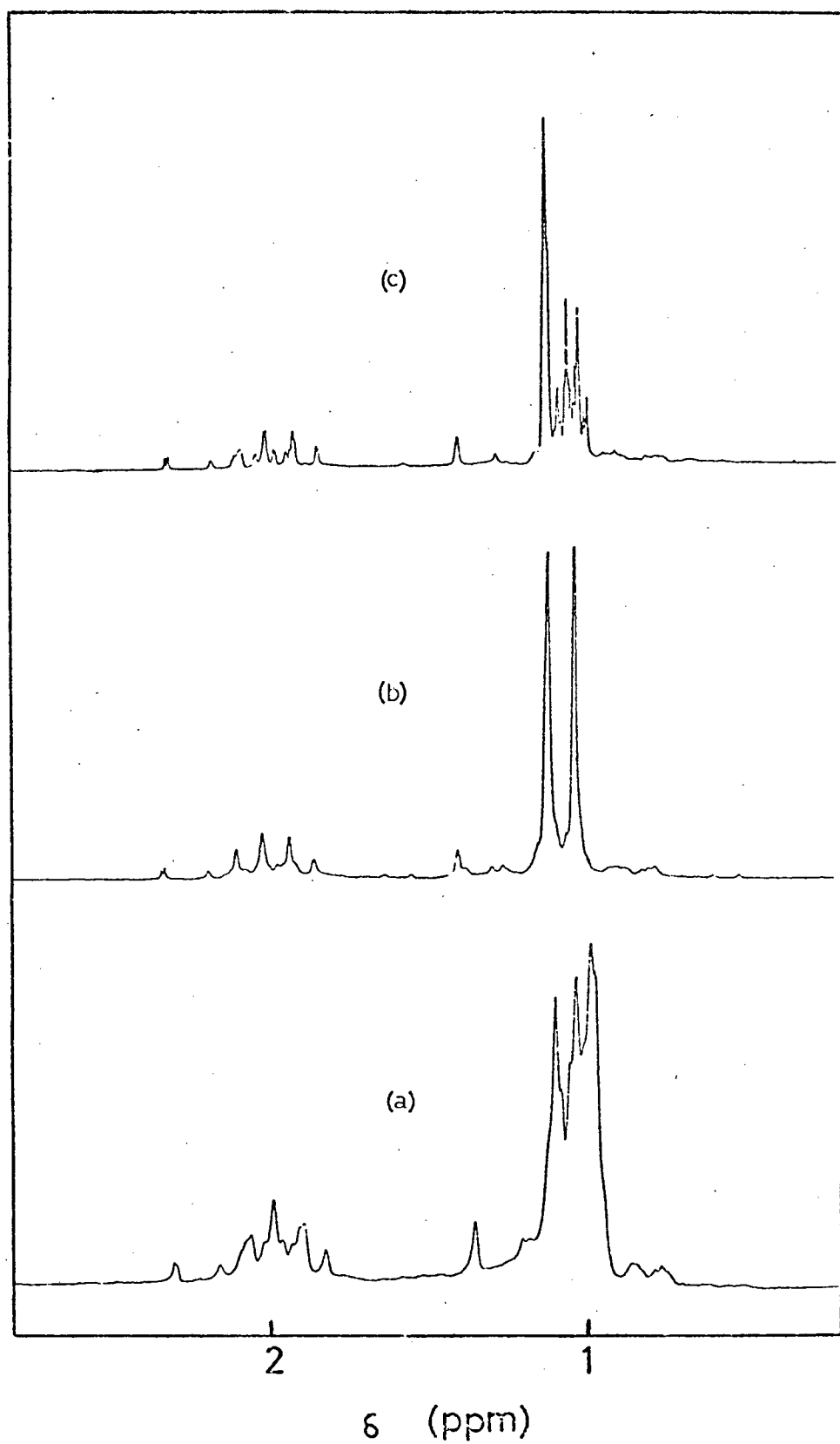
Table 4.5

Torsion angles (degrees) and energies (kcal) for potential energy minima						
	A	B	C	A'	B'	C'
τ_1	100,1	97,8	105,3	261,0	268,7	276,6
τ_2	114,4	198,8	259,2	127,6	195,5	284,8
μ (min)	-,4	2,5	3,7	-,8	,3	-3,3

is gratifying close to that of B' in Figure 4.6, where $\tau_1 = 268,7^\circ$ and $\tau_2 = 195,5^\circ$.

As was mentioned above, the room temperature ^1H NMR spectrum of 2,3 dicyclopropyl-1,4-naphthoquinone exhibits a highly symmetrical pattern around the methylene and methine proton absorption region. On heating or cooling, the simple doublet observed collapses and the true complexity of the spectrum becomes evident (Figure 4.7). In general, a reversible temperature dependence in the ^1H NMR spectrum of a compound can be the result of one or more of the following:

- (i) Rate processes involving intermolecular proton transfers.
- (ii) Rotations about single bonds with partial double bond character or intramolecular close contacts causing rotational restrictions.
- (iii) Inversion of carbocyclic and heterocyclic rings.
- (iv) Valence isomerisations and intramolecular rearrangements.
- (v) Temperature sensitive solvent interactions.



Methine and methylene ^1H NMR spectrum of compound at (a) -35°C
(b) 28°C and (c) 76°C

Fig 4.7

In the present study this dependence is most likely due to measurable differences in energy between molecular conformations, described in terms of the torsion angles τ_1 and τ_2 , such that molecules spend a variable length of time in any one given conformation relative to another. As the temperature changes, so too does the population distribution, and the ^1H NMR spectrum, which sees the average of the conformations, is accordingly affected. Hence 'free' rotation about the C(2)-C(21) and C(3)-C(31) bonds is restricted in the sense that the barriers separating the energy minima are sufficiently large that the molecule spends more of its time in those of lowest energy, yet sufficiently small that interconversion between minima is fast on the NMR time scale.

Although the protons lying *cis* to the methine proton (Figure 4.8) are thermodynamically equivalent, they are in general magnetically non-equivalent as the couplings HaHb and HaHb' are observed to differ. The analogous also holds for those protons lying *trans* to the methine proton. Thus the spectrum of the cyclopropyl protons constitutes a complex AA' BB' C spin system, the appearance of which is determined by the signs and magnitudes of these couplings.

Figure 4.8 shows a schematic representation of the methine and methylene chemical shift changes considered to be taking place as a function of temperature. At -35°C , the overall environment of the cyclopropyl rings is such that the chemical shifts of the Ha and Hb protons are different. The ^1H NMR spectrum in this region hence appears as the nett resultant of a complicated absorption pattern, of the AA' BB' C type discussed above, incorporating both vicinal and

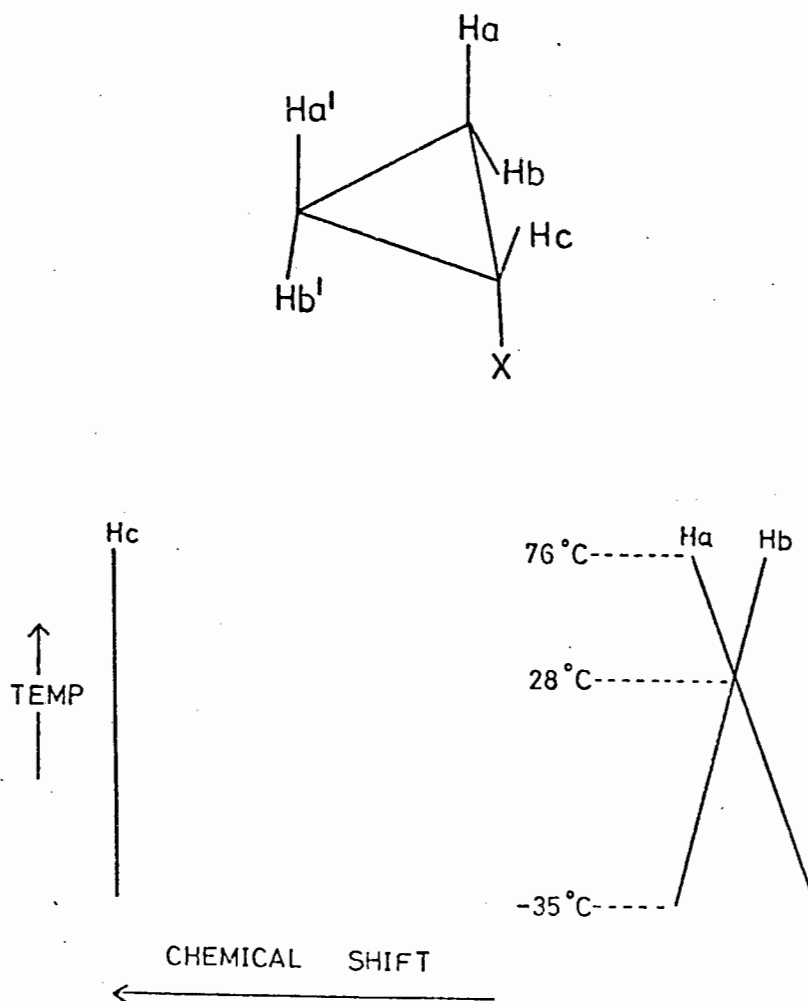


Fig 4.8

geminal coupling, and whose interpretation cannot be explained in terms of first order splitting rules. At room temperature, the conformational population distribution is such that the chemical shift difference between the Ha and Hb protons becomes negligible i.e. $\delta\text{Ha} = \delta\text{Hb}$ 'by accident',^{4.9} and the spectrum degenerates into a deceptively simple spectrum of the A A' A'' A''' C type with a quintet/doublet pattern as observed. The splitting in the multiplets is here an average of the coupling constants involved. As the temperature continues to rise, the chemical shift differences are no longer zero (the chemical shifts of the cyclopropyl protons vary characteristically with the orientation of substituents), and the pattern again becomes complex.

Whether in fact the proton resonances actually 'cross over', as is implied in Figure 4.8, or merely change direction can only be ascertained by a complete analysis of the spectra. This detailed analysis, if possible, could hopefully also yield information as to the nature of the conformational minima, ideally matching those obtained from solid state coordinates, and their relative populations as a function of temperature.

APPENDIX A

Observed and calculated structure factors for $\text{Na}^+[\text{DyEDTA}(\text{3H}_2\text{O})]^- \cdot 5\text{H}_2\text{O}$

H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC					
4	0	0	752	731	0	0	0	97	87	14	16	0	97	81	2	20	0	306	341	3	5	1	367	376
8	0	0	69	63	10	8	0	626	627	16	16	0	179	153	8	20	0	126	135	5	5	1	119	99
12	0	0	603	584	14	8	0	510	501	2	16	0	236	226	10	20	0	276	297	7	5	1	265	309
16	0	0	178	187	2	10	0	555	550	4	16	0	444	463	12	20	0	199	190	9	5	1	616	566
20	0	0	166	181	4	10	0	180	180	8	10	0	343	344	4	30	0	243	271	11	5	1	314	315
4	2	0	744	790	6	10	0	839	844	10	10	0	157	158	6	30	0	345	380	13	5	1	239	246
6	2	0	616	590	8	10	0	378	354	14	18	0	77	32	8	30	0	234	241	15	5	1	444	439
8	2	0	510	514	10	10	0	289	303	16	18	0	419	430	10	30	0	143	146	17	5	1	254	254
10	2	0	258	279	12	10	0	63	96	8	20	0	291	290	8	30	0	264	255	19	5	1	166	165
12	2	0	156	171	14	10	0	311	300	2	20	0	127	133	4	32	0	183	212	1	7	1	413	450
16	2	0	592	592	16	10	0	122	128	4	20	0	319	342	8	32	0	210	197	3	7	1	224	207
18	2	0	206	209	18	10	0	459	462	6	20	0	100	127	4	34	0	333	339	5	7	1	487	436
20	2	0	242	323	8	12	0	892	859	8	20	0	221	239	6	34	0	302	301	7	7	1	505	509
2	4	0	821	855	2	12	0	293	324	10	20	0	222	237	8	36	0	256	264	9	7	1	481	464
4	4	0	210	217	4	12	0	170	130	12	20	0	440	440	2	36	0	202	205	11	7	1	241	236
6	4	0	138	142	6	12	0	154	169	14	20	0	200	208	4	36	0	142	119	13	7	1	383	382
8	4	0	99	69	8	12	0	250	248	16	20	0	110	123	3	1	1	554	507	15	7	1	150	147
10	4	0	322	322	10	12	0	347	655	2	22	0	271	330	5	1	1	563	548	17	7	1	261	261
12	4	0	345	352	12	12	0	454	449	4	22	0	167	190	7	1	1	348	332	19	7	1	282	282
14	4	0	434	419	14	12	0	271	297	6	22	0	442	433	9	1	1	676	632	1	9	1	123	110
20	4	0	214	219	16	12	0	171	153	8	22	0	197	196	11	1	1	389	362	3	9	1	678	709
2	6	0	271	301	4	14	0	662	692	10	22	0	143	184	13	1	1	284	280	5	9	1	379	365
4	6	0	160	142	6	14	0	459	459	12	22	0	75	91	15	1	1	356	356	7	9	1	550	528
6	6	0	730	685	8	14	0	589	591	14	22	0	169	225	17	1	1	270	282	9	9	1	545	507
8	6	0	63	51	10	14	0	64	62	16	22	0	177	181	19	1	1	205	207	11	9	1	358	349
10	6	0	419	406	14	14	0	87	78	8	24	0	74	53	3	3	1	84	83	13	9	1	369	373
14	6	0	198	206	16	14	0	351	348	2	24	0	467	437	5	3	1	562	565	15	9	1	195	187
16	6	0	292	285	18	14	0	116	150	4	24	0	165	204	7	3	1	684	688	17	9	1	277	242
18	6	0	452	454	8	16	0	712	748	10	24	0	438	455	9	3	1	183	154	19	9	1	224	237
20	6	0	193	192	2	16	0	360	366	14	24	0	318	295	11	3	1	380	401	1	11	1	634	657
0	8	0	263	240	4	16	0	104	96	2	26	0	227	256	13	3	1	366	340	3	11	1	548	551
2	8	0	553	604	8	16	0	376	359	6	26	0	469	480	17	3	1	312	312	5	11	1	355	359
4	8	0	306	306	10	16	0	130	156	14	26	0	207	215	19	3	1	279	290	7	11	1	440	421
6	8	0	362	330	12	16	0	566	567	0	26	0	191	154	1	5	1	461	495	9	11	1	528	520
H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC					
11	11	1	233	218	5	19	1	293	297	7	27	1	125	133	20	2	2	117	121	2	12	2	149	155
13	11	1	255	233	7	19	1	363	388	9	27	1	254	240	2	4	2	250	220	4	12	2	571	569
15	11	1	334	340	9	19	1	133	117	11	27	1	212	221	4	4	2	449	438	6	12	2	498	478
17	11	1	189	190	11	19	1	263	265	13	27	1	124	123	6	4	2	605	579	8	12	2	467	467
19	11	1	157	169	13	19	1	280	262	1	29	1	227	249	8	4	2	666	572	10	12	2	279	250
1	13	1	553	547	15	19	1	159	143	5	29	1	266	265	10	4	2	403	394	14	12	2	217	223
3	13	1	156	165	17	19	1	184	177	7	29	1	207	205	12	4	2	189	176	16	12	2	204	208
5	13	1	319	371	1	21	1	169	174	11	29	1	210	214	14	4	2	212	217	18	12	2	348	345
7	13	1	366	377	3	21	1	565	594	1	31	1	121	159	16	4	2	336	339	0	14	2	127	140
9	13	1	218	236	5	21	1	95	119	3	31	1	266	270	18	4	2	320	322	2	14	2	682	697
11	13	1	409	346	7	21	1	209	239	5	31	1	139	166	20	4	2	196	216	4	14	2	150	122
13	13	1	340	351	9	21	1	315	310	7	31	1	127	156	8	6	2	649	674	6	14	2	95	84
17	13	1	319	321	11	21	1	137	128	9	31	1	217	213	2	6	2	130	935	8	14	2	121	186
19	13	1	256	260	13	21	1	141	143	11	31	1	154	169	4	6	2	451	445	10	14	2	539	518
1	15	1	441	457	15	21	1	336	336	1	33	1	145	159	6	6	2	71	54	12	14	2	218	231
3	15	1	442	445	1	23	1	377	392	3	33	1	212	246	8	6	2	557	516	14	14	2	277	293
5	15	1	344	357	3	23	1	170	150	5	33	1	197	218	10	6	2	239	252	16	14	2	155	138
7	15	1	312	309	5	23	1	313	314	7	33	1	185	172	12	6	2	533	511	2	16	2	417	412
9	15	1	376	346	7	23	1	292	187	9	33	1	182	168	14	6	2	248	254	4	16	2	132	139
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7	1	9	250	246	7	11	9	181	183	3	25	9	131	128	0	14	10	272	236	9	9	11	194	177					
9	1	9	259	251	9	11	9	331	315	2	0	10	286	315	2	14	10	349	336	1	11	11	130	139					
11	1	9	229	218	11	11	9	178	175	6	0	10	528	549	10	14	10	322	302	3	11	11	352	348					
13	1	9	155	139	13	11	9	159	153	10	0	10	195	205	2	16	10	205	190	5	11	11	167	156					
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9	7	9	199	219	1	19	9	277	265	12	6	10	306	325	3	5	11	282	290	2	6	12	253	233					
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H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC					
0	12	12	252	249	2	12	12	222	225																				

APPENDIX B

Observed and calculated structure factors for $\text{Cs}^+[\text{YbEDTA}(\text{2H}_2\text{O})]^- \cdot 3\text{H}_2\text{O}$

H	K	L	F _O	F _C	H	K	L	F _O	F _C	H	K	L	F _O	F _C	H	K	L	F _O	F _C	H	K	L	F _O	F _C
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7	0	0	75	-88	3	3	0	287	-324	11	5	0	26	23	9	8	0	48	-47	11	1	1	22	-20
8	0	0	229	257	4	3	0	68	72	12	5	0	34	-34	1	9	0	78	80	12	1	1	53	-50
9	0	0	160	-174	5	3	0	122	140	13	5	0	40	36	2	9	0	101	-108	13	1	1	69	64
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12	0	0	110	-102	7	3	0	203	228	1	6	0	172	174	4	9	0	15	-12	-14	2	1	94	41
13	0	0	108	103	8	3	0	64	-69	2	6	0	13	9	5	9	0	64	-70	-13	2	1	68	-87
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-3	5	16	75	-75	-5	2	17	126	125	-3	5	17	83	88	2	2	18	28	-31	-7	2	19	86	-89
-2	5	16	54	63	-4	2	17	94	-94	-2	5	17	70	-73	-9	3	18	59	-52	-6	2	19	83	83
-1	5	16	22	-23	-3	2	17	22	21	-1	5	17	23	16	-8	3	18	123	124	-5	2	19	32	-33
0	5	16	14	-14	-2	2	17	54	51	0	5	17	62	59	-7	3	18	128	-134	-4	2	19	32	-29
H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
-3	2	19	86	90	-6	3	19	15	-3	-6	0	20	28	23	-2	0	20	34	-39	-4	1	20	54	59

APPENDIX C

Observed and calculated structure factors for $C_{22}H_{19}O_4P$.

H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
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4	0	0	6	-5	1	3	0	29	-30	8	5	0	19	-20	-5	1	1	16	-15	6	2	1	16	17
5	0	0	12	11	2	3	0	3	3	9	5	0	12	-12	-4	1	1	61	-63	7	2	1	3	2
6	0	0	66	-53	3	3	0	21	20	10	5	0	5	6	-3	1	1	46	-48	9	2	1	40	-35
7	0	0	8	8	4	3	0	44	-41	11	5	0	21	21	-2	1	1	59	-63	10	2	1	20	21
8	0	0	40	40	5	3	0	49	-46	0	6	0	5	-5	-1	1	1	65	70	12	2	1	14	-13
9	0	0	9	-8	6	3	0	3	-4	2	6	0	17	18	1	1	1	109	-121	13	2	1	9	-5
10	0	0	31	-31	7	3	0	21	22	3	6	0	17	16	2	1	1	68	68	14	2	1	14	14
11	0	0	10	-9	8	3	0	31	29	4	6	0	57	-58	3	1	1	49	49	-14	3	1	5	-6
12	0	0	20	19	9	3	0	47	-46	5	6	0	18	-19	5	1	1	13	12	-13	3	1	4	4
14	0	0	11	-10	10	3	0	14	-15	6	6	0	13	13	6	1	1	5	-3	-12	3	1	4	2
2	1	0	4	-4	11	3	0	22	21	7	6	0	20	20	7	1	1	63	62	-11	3	1	9	-8
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4	1	0	20	-18	13	3	0	8	-8	9	6	0	17	-17	9	1	1	40	-39	-9	3	1	13	13
5	1	0	31	28	0	4	0	41	-43	10	6	0	38	39	10	1	1	22	-21	-8	3	1	6	7
7	1	0	32	-28	1	4	0	6	6	12	6	0	14	-13	11	1	1	8	7	-7	3	1	9	-10
8	1	0	19	-17	2	4	0	42	41	1	7	0	6	5	12	1	1	5	6	-6	3	1	36	-36
9	1	0	19	17	3	4	0	57	-55	2	7	0	13	-12	13	1	1	5	2	-5	3	1	6	-5
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12	1	0	7	-8	5	4	0	24	22	4	7	0	6	-5	-12	2	1	31	-32	-3	3	1	14	11
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H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
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APPENDIX D

Observed and calculated structure factors for $C_{14}H_{15}O_4P$.

H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
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4	0	0	38	-37	0	4	0	10	-12	11	0	1	10	-8	1	2	1	44	43	2	4	1	13	12
6	0	0	40	-38	1	4	0	20	20	-12	1	1	14	-15	2	2	1	97	96	3	4	1	12	-12
10	0	0	27	-27	3	4	0	5	6	-11	1	1	7	8	3	2	1	12	12	5	4	1	9	-6
12	0	0	7	-7	4	4	0	5	4	-9	1	1	19	-21	4	2	1	23	-23	6	4	1	24	-24
1	1	0	92	-99	5	4	0	7	-7	-8	1	1	61	63	5	2	1	36	-35	7	4	1	30	31
2	1	0	54	54	6	4	0	22	20	-7	1	1	37	-37	6	2	1	18	-19	8	4	1	6	-3
3	1	0	109	111	7	4	0	29	29	-6	1	1	49	-49	8	2	1	10	-9	10	4	1	22	23
4	1	0	26	27	10	4	0	9	-5	-5	1	1	53	-53	10	2	1	12	11	-9	5	1	16	-15
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1	2	0	33	-31	0	6	0	9	-9	4	1	1	23	22	-1	3	1	28	26	0	5	1	17	-18
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8	3	0	18	-16	7	0	1	5	2	-1	2	1	33	-34	0	4	1	26	-26	-4	7	1	23	23
H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
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-9	4	8	6	-5	2	1	9	11	-11	-6	4	9	24	-23	-9	2	10	17	11	0	5	10	6	-7
-7	4	8	8	-8	6	1	9	14	14	-5	4	9	21	24	-7	2	10	14	14	1	5	10	6	6
-6	4	8	27	29	-12	2	9	10	-10	-4	4	9	3	-8	-5	2	10	9	-9	-11	0	11	9	-8
-5	4	8	24	24	-11	2	9	15	15	-3	4	9	11	14	-3	2	10	23	-25	-7	0	11	6	7
H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
-5	0	11	13	-14	-1	2	11	15	15	2	0	12	7	5	1	3	12	6	1	-9	4	13	9	-6
-1	0	11	32	33	1	2	11	6	-6	-9	1	12	11	11	-7	4	12	17	-17	-6	4	13	7	-7
1	0	11	7	-7	3	2	11	10	-9	-7	1	12	9	-9	-6	4	12	10	9	-17	0	14	8	-7
3	0	11	13	-13	-11	3	11	6	-7	-6	1	12	9	9	-2	4	12	7	3	-10	0	14	13	13
5	0	11	8	10	-10	3	11	14	14	-4	1	12	10	-10	-4	5	12	6	-5	-8	0	14</		

APPENDIX E

Observed and calculated structure factors for $C_{16}H_{14}O_2$.

H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC	H	K	L	FO	FC
2	4	0	152	152	0	4	0	20	17	5	7	1	13	-12	1	3	2	13	-15	3	12	2	18	17
4	0	0	61	62	4	0	0	18	17	0	1	1	79	-77	3	3	2	64	68	3	14	2	26	29
6	0	0	24	24	4	14	0	18	-14	0	8	8	16	-13	3	3	2	28	-31	0	16	2	15	15
2	2	1	108	-111	2	2	1	107	109	2	2	1	51	-53	0	0	2	11	-7	2	16	14	14	14
4	2	2	23	-23	4	4	1	10	-12	4	4	1	20	-19	0	0	2	28	-27	2	16	13	13	-7
6	2	2	18	-19	5	1	1	25	27	1	1	1	37	-38	4	4	2	46	-46	1	1	3	65	63
2	2	2	200	-200	2	2	2	43	-44	2	2	2	20	-20	3	3	2	47	-48	3	3	3	35	36
4	2	2	21	-18	4	2	2	35	-16	6	1	1	17	-15	4	4	2	36	-34	4	1	1	54	56
6	2	2	14	-7	5	1	1	20	-43	1	1	1	23	-22	4	4	2	12	-9	3	1	1	12	-15
2	4	3	14	-49	2	2	3	43	-43	6	1	1	15	-17	2	2	5	28	-26	5	1	1	12	-9
4	4	4	47	-49	2	2	3	20	-22	1	1	1	13	-12	2	2	5	13	-11	5	6	1	12	13
6	4	4	127	-126	3	3	3	13	-10	2	2	1	43	-40	3	3	5	25	-25	0	2	2	63	60
2	4	4	41	-41	3	3	1	24	-21	0	1	1	14	-10	4	4	5	20	-20	0	2	2	15	-10
4	4	4	20	-23	5	3	1	18	-20	2	2	1	14	-11	1	1	6	38	-36	4	4	2	136	-134
6	4	4	90	-91	5	3	1	81	-81	2	2	1	18	-16	2	2	6	17	-18	3	3	3	25	25
2	6	6	93	-91	0	4	4	42	-43	3	1	1	13	-11	4	4	6	65	-64	5	3	3	27	-27
4	6	6	45	-45	2	4	4	88	-92	5	3	1	19	-20	5	5	6	19	-20	5	3	3	15	-17
6	6	6	92	-95	3	4	4	46	-47	2	2	2	52	-50	1	2	6	48	-47	0	3	3	27	-26
2	6	6	14	-16	4	4	1	41	-42	3	0	0	63	-62	2	2	7	36	-37	5	4	3	21	18
4	6	6	33	-37	5	4	1	17	-18	4	0	0	25	-23	3	7	2	16	-18	1	4	3	27	26
6	6	6	50	-50	5	5	1	22	-22	6	0	0	13	-11	4	7	2	12	-16	2	4	3	30	-33
2	8	8	26	-25	2	2	5	82	-83	7	0	0	22	-19	0	6	2	16	-12	3	4	3	40	-38
4	8	8	35	-37	2	2	5	19	-16	1	1	2	41	-42	1	8	2	55	-54	3	4	3	31	-35
6	8	8	27	-22	6	0	5	21	-18	2	1	2	35	-34	2	8	2	36	-37	3	5	3	46	-47
2	4	9	16	-12	4	6	1	92	-92	3	1	2	30	-31	5	8	2	19	-18	3	5	3	19	-16
4	9	9	14	-13	2	6	1	41	-40	4	1	2	11	-6	6	8	2	16	-19	0	6	3	27	-26
6	9	9	47	-44	3	6	1	50	-49	5	1	2	29	-32	4	9	2	23	-22	2	6	3	39	-37
2	10	10	17	-17	4	6	1	54	-57	0	2	2	15	-15	0	10	2	26	-26	3	6	3	17	-16
4	10	10	19	-21	4	6	1	27	-29	2	2	2	61	-61	1	10	2	30	-29	3	6	3	45	-45
6	10	10	16	-13	6	6	1	40	-37	2	2	2	97	-96	1	11	2	15	-15	4	6	3	13	-9
2	12	12	20	-23	2	7	1	12	-10	3	2	2	11	-17	2	11	2	15	-15	5	6	3	24	-26
4	12	12	33	-37	3	7	1	12	-16	4	2	2	16	-16	0	12	2	12	-7	2	7	3	12	-8
6	12	12	19	-24	3	7	1	28	-30	5	2	2	30	-42	2	12	2	17	-15	2	7	3	19	-22
7	7	3	23	24	4	5	4	39	-40	1	5	5	28	-29	3	0	6	37	-37	2	1	7	29	-28
0	8	8	13	13	0	6	4	41	-41	2	5	5	28	-30	4	0	6	14	-14	3	1	7	23	-22
3	8	3	18	-29	1	6	4	13	-13	3	5	5	19	-23	3	1	6	21	-19	4	1	7	27	-32
5	8	3	22	-26	4	6	4	13	-5	4	5	5	51	-18	0	2	6	111	-109	0	2	7	19	-17
7	8	3	23	-26	3	7	4	14	-14	0	6	5	19	-49	3	2	6	33	-35	3	2	7	47	-44
9	8	3	20	-19	4	7	4	18	-23	3	4	2	26	-26	4	2	6	11	-10	3	2	7	43	-42
9	9	3	18	-19	5	8	4	12	-14	4	6	7	20	-22	1	3	6	70	-69	3	2	7	18	-15
5	9	3	26	-31	2	9	4	12	-16	2	7	5	22	-21	3	3	6	18	-13	2	3	7	10	-17
6	9	3	13	-15	1	9	4	18	-20	4	7	5	18	-18	0	4	6	21	-24	3	4	7	13	-18
10	9	3	15	-15	3	9	4	15	-24	0	8	5	19	-18	2	4	6	8	-8	1	4	7	12	-13
11	9	3	15	-12	5	10	4	25	-22	1	8	5	17	-14	1	5	6	63	-63	3	4	7	12	-14
14	9	3	14	-13	5	11	4	13	-11	2	8	5	22	-23	2	5	6	71	-69	4	4	7	15	-16
14	10	3	20	-20	2	12	4	12	-19	3	8	5	18	-18	3	5	6	33	-38	3	5	7	35	-32
0	16	3	17	-16	3	12	4	23	-25	4	8	5	13	-12	5	5	6	12	-2	3	5	7	19	-15
0	16	4	20	-18	4	12	4	13	-13	6	8	5	13	-16	6	5	6	15	-13	6	5	7	13	-8
2	17	4	32	-34	4	13	4	12	-7	3	9	5	13	-15	0	6	6	34	-35	0	6	7	42	-43
3	17	4	18	-20	3	13	4	26	-27	3	9	5	13	-11	3	6	6	13	-11	3	6	7	23	-21
4	17	4	45	-47	4	13	4	18	-17	4	9	5	15	-10	1	7	6	22	-20	2	6	7	30	-32
4	18	4	23	-24	4	14	4	15	-14	5	9	5	20	-23	0	8	6	30	-27	3	6	7	13	-11
6	18	4	39	-41	2	14	4	14	-11	2	10	5	18	-20	1	8	6	24	-19	4	6	7	14	-16
6	19	4	25	-25	1	15	4	79	-31	1	11	5	13	-4	1	9	6	24	-23	4	6	7	14	-7
2	20	4	47	-46	0	2	5	44	-45	3	11	5	31	-34	0	10	6	23	-26	1	7	7	34	-31
2	20	4	18	-17	1	2	5	13	-11	3	11	5	14	-23	1	10	6	15	-10	2	7	7	20	-22
2	22	4	16	-16	2	2	5	34	-33	0	12	5	16	-16	1	11	6	28	-32	3	7	7	13	-21
3	22	4	25	-26	4	2	5	20	-16	4	12	5	12	-12	4	11	6	17	-19	4	7	7	18	-18
3	23	4	18	-14	5	2	5	12	-11	5	12	5	28	-25	5	11	6	18	-11	5	7	7	14	-12
4	23	4	61	-65	6	2	5	15	-9	4	12	5	15	-15	4	12	6	15	-9	5	8	7	15	-10
4	24	4	12	-5	2	3	5	11	-10	3	13	5	13	-12	1	13	6	18	-23	3	8	7	14	-10
4	24	4	29	-28	3	3	5	26	-25	2	14	5	18	-17	2	13	6	22	-21	3	8	7	19	-21
4	24	4	18	-20	4	3	5	21	-24	0	16	5	18	-21	3	13	6	18	-19	1	9	7	19	-22
4	24	4	10	-5	0	4	5	48	-48	0	16	6	48	-47	1	15	6	22	-21	2	9	7	29	-29
4	24	4	16	-14	1	4	5	42	-43	1	0	6	67	-66	2	15	6	18	-15	3	9	7	17	-15
6	24	4	12	-14	2	1	5	18	-17	2	0	6	68	-70	1	1	7	37	-34	3	10	7	15	-13
1	10	7	13	-10	4	5	8	15	-5	3	5	9	23	-18	0	8	10	12	-9	4	2	12	15	-9
4	10	7	21	-22	0	6	8	15	-16	5	5	9	17	-10	3	8	10	14	-14	1	2	12	26	-22
1	11	7	20	-21	5	6	8	21	-20	0	6	9	29	-25	3	9	10	16	-11	2	3	12	18	-21
0	12	7	17	-17	1	7	8	15	-11	0	6	9	25	-24	0	10	10	13	-5	0	4	12	22	-22
1	12	7	21	-18	2	7	8	14	-20	3	6	9	13	-10	1	10	10	14	-13	1	4	12	21	-23
2	12	7	14	-6	3	7	8	34	-33	5	6	9	20	-20	2	10	10	27	-26	2	4	12	16	-13
0	13	8	30	-31	2	8	8	19	-20	4	7	9	22	-22										

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SOLUTION EQUILIBRIA OF THE METHYLDOPA-COPPER(II) SYSTEM
AND THE
CRYSTAL AND MOLECULAR STRUCTURES OF SELECTED COMPOUNDS

A thesis submitted to
THE UNIVERSITY OF CAPE TOWN
in fulfilment of the requirements for the degree of
DOCTOR OF PHILOSOPHY

by

M. ROBERT W. WRIGHT

Department of Physical Chemistry,
University of Cape Town,
Rondebosch, 7700,
Republic of South Africa.

August, 1979

SOLUTION EQUILIBRIA OF THE METHYLDOPA-COPPER(II) SYSTEM

ABSTRACT

The system copper(II) ions in aqueous solution in the presence of L-3-(3',4'-dihydroxyphenyl)-2-methylalanine (methyldopa), at 25°C and $I = 0,150 \text{ mol dm}^{-3} \text{ Na}^+ \text{ClO}_4^-$ has been characterised using glass electrode potentiometry. Both mononuclear and oligonuclear complexes were found to occur, the former including a series of successively deprotonated species in the higher ligand to metal ratio titrations, the latter including a cyclic dimer in the 1:1 ligand to metal solutions. Interpretation of the results was facilitated by (i) ultra-violet/visible spectrophotometry of solutions containing copper(II), methyldopa or alanine or catechol at various pH's, (ii) ^1H NMR measurements of methyldopa in $^2\text{H}_2\text{O}$ solutions, (iii) glass electrode potentiometric measurements on alanine and catechol in the presence of copper(II) ions. Formation constants are given for 3 proton - and 11 copper(II) - complexes of methyldopa, 2 proton and 2 copper(II) complexes of alanine and 1 proton and 2 copper(II) complexes of catechol.

PACKING PATTERNS IN LANTHANIDE-EDTA COMPLEXES: CRYSTAL AND MOLECULAR STRUCTURES OF SODIUM TRIAQUOETHYLENE-DIAMINETETRAACETATODYSPROSIUM(III) PENTAHYDRATE AND CAESIUM DIAQUOETHYLENEDIAMINETETRAACETATOYTTERBIUM(III) TRIHYDRATE

ABSTRACT

$\text{Na}^+ [\text{DyEDTA}(3\text{H}_2\text{O})]^- \cdot 5\text{H}_2\text{O}$ crystallises in the space group Fdd2 with $\underline{a} = 19,54(1)$, $\underline{b} = 35,40(2)$, $\underline{c} = 12,116(6)$ Å, $\underline{z} = 16$. $\text{Cs}^+ [\text{YbEDTA}(2\text{H}_2\text{O})]^- \cdot 5\text{H}_2\text{O}$ crystallises in the space group $\text{P2}_1/\text{c}$ with $\underline{a} = 12,935(6)$, $\underline{b} = 8,997(5)$, $\underline{c} = 17,22(1)$ Å, $\beta = 106,7(2)^\circ$, $\underline{z} = 4$. The structures were determined by Patterson and Fourier methods and refined by full-matrix least-squares to a final R of 0,045 and 0,050 respectively. The decrease in ionic radius from Dy^{3+} to Yb^{3+} results in a change in coordination number of the Ln^{3+} ion from 9 to 8 with concomitant loss of a water molecule. The changed geometry of the lanthanide complex gives rise to a different space group.

THE CRYSTAL STRUCTURES OF A PENTA-COORDINATE PHOSPHORUS
COMPOUND: 3,4,8,9-DIBENZ-5,7-DIMETHYL-1-PHENYL-2,6,10,11-
TETRAOXATRICYCLO[5,3,1,0^{1,5}] PHOSPHA-UNDECANE AND ITS
HYDROLYSIS PRODUCT: PHENYL-1-HYDROXY-1-(2'-HYDROXYPHENYL)-
ETHYLIDYNYL PHOSPHINIC ACID

ABSTRACT

Reaction of PhPCl_2 with $o\text{-HOC}_6\text{H}_4\text{COCH}_3$ gives an unusual tricyclic tetraoxaphospha-undecane. Crystals of this compound are monoclinic, $P2_1/c$, with $a = 15,874(7)$, $b = 7,924(4)$, $c = 16,572(8)$ Å, $\beta = 94,5(2)^\circ$, $z = 4$. The geometry around the phosphorus atom is distorted trigonal bipyramidal, with the two five-membered and one six-membered rings all in apical-equatorial positions. It is postulated that permutational isomerisation in solution is unlikely for this compound due in part to its molecular rigidity. Crystals of the disubstituted phosphinic acid are monoclinic, $P2_1/n$, with $a = 13,886(7)$, $b = 7,378(4)$, $c = 15,280(8)$ Å, $\beta = 116,9(2)^\circ$, $z = 4$. The phosphorus atom has the expected tetrahedral environment. O...O close contacts less than $2,76(1)$ Å indicate hydrogen bonding which extends throughout the lattice.

CRYSTAL STRUCTURE AND MOLECULAR CONFORMATION OF 2,3-DICYCLOPROPYL-1,4-NAPHTHOQUINONE

ABSTRACT

Crystals of the title compound are orthorhombic with $a = 8,483(4)$, $b = 18,249(9)$, $c = 15,889(8)$ Å, space group $Pbca$, $z = 8$. The quinone ring shows distinct distortions from planarity. The solution NMR spectrum of the compound at various temperatures suggest a measure of restricted rotation about the cyclopropyl - quinonoid bonds. This is partially substantiated by a conformational analysis using the method of atom pair potentials.